



# Using collision-induced dissociation to constrain sensitivity of ammonia chemical ionization mass spectrometry (NH<sub>4</sub><sup>+</sup>-CIMS) to oxygenated volatile organic compounds

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**Abstract.** Chemical ionization mass spectrometers (CIMS) routinely detect hundreds of oxidized organic compounds in the atmosphere. A major limitation of these instruments is the uncertainty in their sensitivity to many of the detected ions. We describe the development of a new high-resolution time-of-flight chemical ionization mass spectrometer that operates in one of two ionization modes: using either ammonium ion ligand switching reactions as NH<sub>4</sub><sup>+</sup>-CIMS or proton transfer reactions  
15 as PTR-MS. Switching between the modes can be done within two minutes. The NH<sub>4</sub><sup>+</sup>-CIMS mode of the new instrument has sensitivities of up to 67,000 dcps ppbv<sup>-1</sup> (duty cycle corrected ion counts per second/parts per billion by volume) and detection limits between 1 and 60 pptv at 2σ for a 1s integration time for numerous oxygenated volatile organic compounds. We present a mass spectrometric voltage scanning procedure based on collision-induced dissociation that allows us to determine the stability of ammonium-organic ions detected by the NH<sub>4</sub><sup>+</sup>-CIMS. Using this procedure, we can effectively  
20 constrain the sensitivity of the ammonia chemical ionization mass-spectrometer to a wide range of detected oxidized volatile organic compounds for which no calibration standards exist. We demonstrate the application of this procedure by quantifying the composition of secondary organic aerosols in a series of laboratory experiments.



## 1 Introduction

Understanding the photochemical oxidation of volatile organic compounds (VOC) in the atmosphere is crucial for estimating their contribution to the formation of secondary organic aerosols (SOA) and tropospheric ozone, key components of photochemical smog (Atkinson, 2000; Shrivastava et al., 2017). Identification and quantification of VOCs have remained an analytical challenge due to the complexity of multigenerational chemical systems and high variability in VOC concentrations in the atmosphere.

Chemical ionization mass spectrometry (CIMS) has become an important analytical tool for measurements of organic molecules in the atmosphere. Reagent ions are typically produced by glow discharge (Hansel et al., 1995) or a radioactive ion-source (Blake et al., 2004). These ions subsequently react with analyte molecules by ligand switching, reactive electron transfer, or proton transfer and form product ions which are later detected by a mass spectrometer. Many modern CIMS instruments use time-of-flight mass spectrometers (ToF-MS) which have high mass resolving power and simultaneous detection of all ions. Some of the benefits of CIMS include high sensitivity, fast time response, linearity, and reproducibility. A variety of reagent ions can be used to detect different classes of VOC. Nitrate ion CIMS has been used to detect highly oxidized organic molecules as well as sulfuric acid (Berresheim et al., 2000; Jokinen et al., 2012). Iodide adduct CIMS and acetate CIMS (both negative ion polarity) have played a key role in the measurement of carboxylic acids (Lee and Lopez-Hilfiker et al., 2014; Bertram et al., 2011).  $\text{CF}_3\text{O}^-$  CIMS has been used to measure specific classes of VOC such as hydroperoxides (Crouse et al., 2006). Protonated water clusters have been used to detect a broad range of chemical compounds containing oxygen, nitrogen and sulfur (Lindinger et al., 1998, Yuan et al., 2017). Recently, two new proton transfer reaction time-of-flight mass spectrometers have been developed: the PTR3 (Breitenlechner et al., 2017) and the VOCUS PTR-TOF (Krechmer et al., 2018). Using  $\text{H}_3\text{O}^+$  reagent ions, both instruments show sensitivities exceeding 10,000 cps  $\text{ppbv}^{-1}$  (counts per second/parts per billion by volume) for select compounds. Detection efficiency and sensitivity of CIMS instruments depend critically on both the reagent ion and the measured sample molecule (Hytinen et al., 2017).

CIMS instruments have been also used for analyzing submicrometer particulate organic matter. Hellen et al. (2008) equipped the inlet of the PTR-MS instrument with a denuder to remove the gas phase organics and a heater to vaporize the aerosol particles. Similarly, Eichler et al. (2015) introduced the CHARON-PTR-ToF-MS setup that transmits particles with a 75-90% efficiency. FIGAERO-HRToF-CIMS (Lopez-Hilfiker et al., 2014) uses a new filter inlet for thermal desorption of ambient submicron particles.

In this work, we describe the use of protonated ammonia molecules (ammonium,  $\text{NH}_4^+$ ) for soft ionization of analyte molecules. Ammonium has been previously used as a CIMS reagent ion. Lindinger et al. (1998) showed that proton transfer reactions can be utilized to softly ionize VOCs yielding product ions  $\text{VOC}\cdot\text{H}^+$ :



The proton transfer reaction (1) is exothermic for molecules that have proton affinities higher than of ammonia (854  $\text{kJ mol}^{-1}$ ) and is therefore more selective than the reaction with traditional hydronium ions as proton donors (proton affinity of water



is  $691 \text{ kJ mol}^{-1}$ ). Blake et al. (2006) showed that numerous VOCs can be detected through an association reaction of analyte molecules with ammonium clusters ( $\text{NH}_4^+$  and  $\text{NH}_4^+(\text{NH}_3)$ ):



5 where M is a third-body molecule. Shen et al. (2009) used these methods for on-line detection of the explosive triacetone triperoxide (TATP).

Most recently, Hansel et al. (2018) showed that ammonium-water clusters can be utilized for soft ionization of organic compounds via exothermic ligand switching reaction:



10 Hansel et al. used a modified version of the PTR3 instrument (called  $\text{NH}_4^+$ -CI3-ToF) to detect first generation peroxy radicals and closed-shell products from ozonolysis of cyclohexene and achieved sensitivities of up to  $28,000 \text{ cps ppbv}^{-1}$  for these compounds. However, the enhanced reaction time and increased pressure (4 ms and 80 mbar comparing to 0.1 ms and 2 mbar for PTR-MS instruments operated under standard conditions, respectively) raise the probability of reverse ligand switching reactions, which make it difficult to estimate sensitivities of the  $\text{NH}_4^+$ -CI3-ToF to species that cannot be calibrated  
15 directly.

In this study, we present a new instrument that is equipped with three similar corona discharge ion sources and currently can be operated in two different modes: (1) ligand switching reactions from adduct ions  $\text{NH}_4^+(\text{H}_2\text{O})_n$ , ( $n=0,1,2$ ) ( $\text{NH}_4^+$ -CIMS) and (2) proton transfer reactions with  $\text{H}_3\text{O}^+(\text{H}_2\text{O})_n$ , ( $n=0,1$ ) ions (PTR-MS). The instrument is a modified version of the PTR3 with a helical tripole reaction chamber and a long time-of-flight mass spectrometer (Tofwerk AG, Switzerland), and it  
20 can be used for measurements of organic molecules in both gas and particle phases. Here we discuss the performance of the new instrument and compare the two detection modes. We demonstrate a mass spectrometric voltage scanning procedure which is based on collision-induced dissociation that allows for the determination of the stability of detected ammonium-organic clusters. With this technique, we can experimentally estimate sensitivities of the  $\text{NH}_4^+$ -CIMS to the vast array of oxygenated organic compounds without their direct calibration in a matter of minutes. Finally, we present how this  
25 procedure can be applied to the measurement of organic aerosol composition in laboratory experiments.

## 2 $\text{NH}_4^+$ -CIMS instrument description

The instrument developed in this work is based on the PTR3, which is described in detail by Breitenlechner et al. (2017). Here, we summarize the basic operating principle and describe the two major design changes made to the original design. The schematic drawing of the  $\text{NH}_4^+$ -CIMS instrument is shown in Fig. 1.

30 Reagent ions are generated in a corona discharge region and are extracted using a source drift region as indicated by red arrows in Fig. 1. The reaction chamber uses a tripole electrode configuration and is operated at typical pressures between 50 and 70 mbar. Unlike many other PTR instruments, there is no axial electric field accelerating ions towards the exit of the



reaction chamber. Therefore, the reaction time is exclusively determined by the flow velocity of the sampled gas in the axial direction, leading to a typical reaction time of 3 ms. The LToF mass spectrometer with mass resolution  $m/\Delta m$  of up to 8000 allows for the separation of the components with the same nominal mass.

The first major instrument design change consists of replacing the single ion source by three ion sources, one active at a time. Currently, we use two sources: one for producing  $\text{H}_3\text{O}^+(\text{H}_2\text{O})_n$ , ( $n=0,1$ ) reagent ions (as PTR-MS, called  $\text{H}_3\text{O}^+$  mode), and another for producing  $\text{NH}_4^+(\text{H}_2\text{O})_n$ , ( $n = 0,1,2$ ) reagent ions (as  $\text{NH}_4^+$ -CIMS, called  $\text{NH}_4^+$  mode).  $\text{NH}_4^+(\text{H}_2\text{O})_n$  ions are produced in the corona discharge ion source from  $\text{NH}_3$  and  $\text{H}_2\text{O}$ . Constant flow (20 sccm) of ammonia and water vapour is added to the ion source region from the head-space of a 10% solution of ammonium hydroxide in water. Fig. 1 shows the instrument in the  $\text{NH}_4^+$  mode with the active ion source on the left, while the two other ion sources (depicted as a single ion source on the right) are inactive. The innermost source drift plate of the active ion source and the innermost source drift plates of both inactive sources generate an electric field perpendicular to the tripole axis. In addition, another component of the electric field is generated parallel to the tripole axis by biasing the electric potential at the secondary orifice relative to the tripole offset potential. Fig. 1 illustrates the resulting electric field in this transfer region. This geometry allows for effective ion guiding from the active ion source to the centre of the reaction tripole chamber. Compared to single-source designs, separate ion sources allow for faster switching between reagent ion species. As shown in Fig. S1, switching from the  $\text{H}_3\text{O}^+$  mode to the  $\text{NH}_4^+$  mode occurs within one minute, while the reverse switching from the  $\text{NH}_4^+$  mode to the  $\text{H}_3\text{O}^+$  mode can be done within two minutes.

The second major design change consists of replacing the straight tripole electrode rods with a helix. Simulations of ion trajectories in the original tripole showed that ions are lost mostly by exiting the device through spaces between the rods, rather than by collisions with the rods themselves, probably due to inhomogeneous effective potentials generated by the tripole radio frequency (RF) fields (Breitenlechner et al., 2017). The helical structure effectively averages these inhomogeneities, increasing the ion transmission efficiency and therefore the overall instrument performance.

The instrument can be used for measurements of organic molecules in both the gas and particle phases. During particle phase measurements, sampled air passes through a gas phase denuder (Ionicon Analytik GmbH, Austria) that removes the gas phase organics and then through a thermal desorption region heated to 180°C that vaporizes the aerosol particles.

### 3 $\text{NH}_4^+$ -CIMS instrument performance

Multiple reagent ions are observed in the mass spectrum of this instrument in the  $\text{NH}_4^+$  mode, including ammonium-water clusters  $\text{NH}_4^+(\text{H}_2\text{O})_n$ , ( $n=0,1,2$ ) and ammonium ammonia dimers  $\text{NH}_4^+(\text{NH}_3)$ . Humidity of the sampled air only slightly affects the distribution of the reagent ions, as shown in Fig. S2.

A series of laboratory experiments were performed to obtain instrument sensitivities to various organic compounds as a function of relative humidity. Table 1 shows sensitivities to 16 compounds measured using a liquid calibration unit (LCU, Ionicon Analytik GmbH, Austria) at 10% RH and 20 °C. The LCU quantitatively evaporates aqueous standards into the gas



stream. 16 standards were prepared gravimetrically or volumetrically, depending on the compound, with aqueous volume mixing ratios of compounds ranging between 2 and 6 ppmv.  $10 \mu\text{l min}^{-1}$  flow of each of these solutions were then evaporated into a humidified gas stream of synthetic air (9 slpm) resulting in calibration standards containing 1-2 ppbv of each calibrated component. In Table 1 we also present sensitivities calculated in duty-cycle-corrected counts per second/parts per billion by volume (dcps/ppbv, normalized to  $m/z = 100$ ). The duty cycle correction compensates for the mass-dependent extraction efficiency into the time-of-flight mass spectrometer:  $\text{dcps}(i) = \text{cps}(i) \cdot \sqrt{100/m_i}$ . The extraction frequency of the ToF was set at 14 kHz. Limits of detection are calculated for a 1 s integration time as two standard deviations of measured background divided by derived sensitivity. Sensitivity to each compound was measured at 10%, 30%, 50% and 70% RH at 20°C.

10 Signals of  $\text{NH}_4^+$ -VOC clusters decrease as humidity of the sampled air increases, as shown in Fig. 2. Increased reaction time (3 ms) and elevated pressure (60 mbar) in the reaction chamber, compared to the conventional PTR-MS instruments (0.1 ms and 2.3 mbar, respectively), promote equilibrium between forward and backward ligand switching reactions (3). Hence, under humid conditions, excess water vapour favours formation of ammonium-water clusters, which in turn reduces the abundance of ammonium-organic clusters  $\text{NH}_4^+(\text{VOC})$  and hence the overall instrument sensitivity to oxygenated VOCs (OVOCs). Generally, a stronger humidity dependence is observed for components with lower sensitivities at dry conditions. Higher molecular weight molecules have weaker humidity dependence.

#### 4 Collision-induced dissociation techniques for constraining sensitivity of the $\text{NH}_4^+$ -CIMS

When the instrument operates in the  $\text{NH}_4^+$  mode, organic molecules are detected almost entirely as ammonium-organic clusters. However, kinetic rate constants of ligand-switching reactions (3) from ammonium-water ions to an organic molecule have only been measured for very few analyte molecules. In addition, enhanced reaction time in the reaction chamber relative to conventional PTR-MS instruments increases probability of reverse ligand-switching reactions. Therefore, effective rate constants for both forward and backward reactions (3) are required for analytical estimation of the compound sensitivities. To avoid these complications, we constrain the instrument sensitivities to the detected compounds through an empirically-based collision-induced dissociation (CID) technique similar to the one used by Lopez-Hilfiker et al. (2016) for constraining sensitivity of iodide adduct CIMS. This is accomplished by varying the voltage between the ionization region and vacuum region of the mass-spectrometer (Fig. 1), which increases the electric field, while measuring intensities of detected peaks in the mass spectrum.

The increase of the collisional kinetic energy of the ammonium-organic clusters and air molecules leads to collision-induced dissociation of the clusters. For each analyte ion we determine the voltage value ( $V_{50}$ ) at which the peak intensity drops by 50% relative to the intensity at the operational voltage value and calculate the ion kinetic energy corresponding to this voltage ( $\text{KE}_{50}$ ). Therefore, we can experimentally determine the electric field strength necessary to break each ammonium-organic cluster, which defines the stability of these clusters and hence the sensitivity of our instrument to analyte molecules.



The value of  $E/N$  ( $E$  is the electric field strength and  $N$  is the sample gas number density) is a suitable metric to characterize the motion of ions in the reaction chamber and kinematics of a chemical ionization reaction (Blake et al., 2006). The electric field strength  $E$  in a particular region of the reaction chamber depends on the voltage  $V$  applied in that region and the effective distance between electrodes  $d$

$$5 \quad E = \frac{V}{d} \quad (4)$$

Drift velocity of ions in the reaction chamber  $v_d$  is determined by the electric field strength  $E$  and the ion mobility  $\mu$ :

$$v_d = \mu E \quad (5)$$

The ion mobility depends on reaction pressure and temperature:

$$\mu = \mu_0 \frac{1013 \text{ mbar}}{p_r} \frac{T_r}{273 \text{ K}} \quad (6)$$

10 where  $\mu_0$  is the reduced mobility, which is estimated for each ion using its mass (Ehn et al., 2011),  $p_r$  is pressure in the reaction chamber (in mbar),  $T_r$  is temperature in the reaction chamber (in K). Further, we calculate mean kinetic energy of drifting ions  $\text{KE}_{\text{ion}}$  in the laboratory frame (Lindinger et al., 1998):

$$\text{KE}_{\text{ion}} = \frac{3}{2} k_B T + \frac{M_{\text{buffer}} v_d^2}{2} + \frac{M_{\text{ion}} v_d^2}{2} \quad (7)$$

15 where  $k_B$  is the Boltzmann constant,  $M_{\text{buffer}}$  and  $M_{\text{ion}}$  are the masses of the buffer molecule in the air and reagent ion, respectively. Finally, the kinetic energy of analyte ions in the centre of mass for ion-molecule collisions is given by (McFarland et al., 1973):

$$\text{KE}_{\text{cm}} = \frac{M_{\text{buffer}}}{M_{\text{buffer}} + M_{\text{ion}}} \left( \text{KE}_{\text{ion}} - \frac{3}{2} k_B T \right) + \frac{3}{2} k_B T \quad (8)$$

20 For each ammonium-organic cluster we measure  $V_{50}$  and from this calculate the corresponding kinetic energy at which half of the ions have dissociated ( $\text{KE}_{\text{cm}50}$ ) using formulas (4)-(8). We show a set of declustering scans for eight organic molecules with different functional groups in Fig. 3. Intensities of all clusters follow similar sigmoidal shapes when the voltage is increased. Some clusters (i.e., small alcohols and heterocyclic compounds) are less stable and are dissociated at lower voltages while other clusters (i.e., large ketones) show higher stability. These scans can be obtained within four minutes by steadily increasing the voltage between the ionization region and vacuum region of the mass-spectrometer (Fig. 1).

25 Fig. 4 shows the relationship between the calculated kinetic energy  $\text{KE}_{\text{cm}50}$  and measured sensitivity for 16 calibrated compounds. We observe a linear relationship ( $R^2 = 0.61$ ) between calculated  $\text{KE}_{\text{cm}50}$  and measured sensitivity for calibrated VOCs. This linear relationship is observed for molecules with  $\text{KE}_{\text{cm}50}$  in the range between 0.10 and 0.19 eV (region B in Fig. 4). Molecules characterized by collisional kinetic energies  $\text{KE}_{\text{cm}50}$  smaller than that of the ammonium-water cluster (0.09 eV, region A in Fig. 4) will show no significant reaction rate since ligand switching reactions between  
 30 such molecules and  $\text{NH}_4^+(\text{H}_2\text{O})$  are endothermic. On the other hand, the ligand switching reaction rate cannot exceed the kinetic limit for ion-molecule collisions, and therefore there is also an upper limit of observed sensitivities. We calculate this limit by using experimentally-determined pressure and reaction time in the reaction chamber (based on the sensitivities



observed in  $\text{H}_3\text{O}^+$  mode) and kinetic limit of ion-molecule reaction rate of  $3 \cdot 10^{-9} \text{ cm}^3 \text{ s}^{-1}$  (Viggiano et al., 1989). In our case, the instrument sensitivity cannot exceed  $70,000 \text{ dcps ppb}^{-1}$ , which is in agreement with the highest sensitivity measured for calibrated compounds. Therefore, we assume that all components with  $\text{KE}_{\text{cm}50}$  greater than  $0.19 \text{ eV}$  (region C in Fig. 4) will be detected at this “kinetic sensitivity”.

## 5 Application to secondary organic aerosols

To demonstrate the application of the procedure described above, we performed a series of laboratory chamber experiments. A complex mixture of organic compounds in both gas and particle phases was generated by the oxidation of 3-methylcatechol ( $\text{C}_7\text{H}_8\text{O}_2$ ), a second-generation oxidation product of toluene and other anthropogenic aromatics, by hydroxyl (OH) radicals in an environmental chamber. Details of the chamber operations are given by Hunter et al. (2014), so we include only a brief description here. Photochemical oxidation occurred in a  $7.5 \text{ m}^3$  temperature-controlled Teflon chamber by OH radicals generated through the photolysis of nitrous acid (HONO). In the experiment described here,  $65 \text{ ppbv}$  of 3-methylcatechol (Sigma-Aldrich, 98% purity) was injected in the chamber and further oxidized in the presence of ammonium nitrate seed aerosol at  $20^\circ\text{C}$  and low humidity (3% RH). Secondary organic aerosol particles produced in this experiment were detected using an Aerodyne Aerosol Mass Spectrometer (AMS, DeCarlo et al., 2006) and the described CIMS instrument operating in both the  $\text{H}_3\text{O}^+$  and  $\text{NH}_4^+$  modes, equipped with the thermal desorption unit described above. In this experiment, we identified 202 peaks in the  $\text{NH}_4^+$  mode mass spectra and grouped them based on the calculated  $\text{KE}_{\text{cm}50}$  as shown in Fig. 5. Among those 202  $\text{OC} \cdot \text{NH}_4^+$  peaks, 125 analyte formulas were also detected as  $\text{OC} \cdot \text{H}^+$  in the  $\text{H}_3\text{O}^+$  mode. We plot the relationship between the detected signals in both modes of our instrument in Fig. 6. We use the declustering technique described above to calculate volume mixing ratios of organic molecules detected as ammonium-organic clusters in the  $\text{NH}_4^+$  mode. In the  $\text{H}_3\text{O}^+$  mode, we apply the calibrated acetone sensitivity to calculate volume mixing ratios of OVOCs. Volume mixing ratios of organic compounds detected by both modes are in excellent agreement with a slope of 0.94 as shown in Fig. 6 ( $R^2=0.78$ ). In addition to 125 peaks measured by both modes, there are peaks that are detected solely by either the  $\text{H}_3\text{O}^+$  or  $\text{NH}_4^+$  modes. In Fig. 7, we plot 34 identified  $\text{C}_x\text{H}_y\text{O}_z \cdot \text{H}^+$  peaks detected by the  $\text{H}_3\text{O}^+$  mode and 17 identified  $\text{C}_x\text{H}_y\text{O}_z \cdot \text{NH}_4^+$  peaks detected by the  $\text{NH}_4^+$  mode on the carbon number-oxidation state diagram. Two modes cover different areas on this diagram: while the  $\text{NH}_4^+$ -CIMS is able to detect larger and more functionalized molecules, PTR-MS is better at detection of smaller organic compounds (some of them can be formed as a result of fragmentation during ionization). Hence, the two modes complement each other and allow for the detection and quantification a broader range of oxidized organic molecules.

Fig. S3 shows a comparison between the total mass loading of all organic components measured by the AMS with the sum of masses of all organic compounds measured by our instrument in both  $\text{H}_3\text{O}^+$  and  $\text{NH}_4^+$  modes. The sum of signals of all components detected in the  $\text{NH}_4^+$  mode account for 65% of the total aerosol organic mass measured by AMS as shown in Fig. 8. This discrepancy can be explained by a combination of the following factors: 1) uncertainties in the sensitivities



obtained using the presented technique and in the AMS measurements; 2) thermal fragmentation of organic molecules in the thermal desorption unit which leads to lower observed masses in the mass spectrum; 3) low  $\text{NH}_4^+$ -CIMS sensitivity to certain compounds of organic aerosols if ligand switching reactions between these molecules and ammonium-water clusters are endothermic (e.g., small organic acids). Although the  $\text{NH}_4^+$ -CIMS does not detect all organic compounds to explain the total organic mass measured by AMS, it gives valuable insight into the composition of SOA as shown in Fig. 8.

## 6 Conclusion

In this study, a new CIMS instrument is described based on the recently introduced PTR3. The instrument is equipped with three corona discharge ion sources and can be operated in both  $\text{NH}_4^+$  and  $\text{H}_3\text{O}^+$  modes as  $\text{NH}_4^+$ -CIMS and PTR-MS, respectively, while switching between the two modes can be done within two minutes. Compared to the  $\text{H}_3\text{O}^+$  mode, the  $\text{NH}_4^+$  mode is able to detect more functionalized and larger organic molecules. In the  $\text{NH}_4^+$  mode, the instrument has sensitivities in the range of 80-65,000 dcps ppbv<sup>-1</sup> and detection limits in the range of 1.5-60 pptv for a 1 second integration time ( $2\sigma$ ). We present a procedure based on collision-induced dissociation that allows us to estimate the stability of detected ammonium-organic clusters and therefore to constrain the sensitivities of hundreds of compounds detected by the  $\text{NH}_4^+$  mode of the new instrument without their direct calibration within several minutes.

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*Author contributions.* MB and AZ designed and built the CIMS instrument. AZ and MB developed the methodology with contributions from ARC and FNK. AZ, ARC and MB performed the laboratory experiments. AZ and MB provided data and analysis for the CIMS instrument. CYL and JCR provided data and analysis for the AMS instrument. AZ prepared the manuscript with contributions from all co-authors.

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*Competing interests.* Authors declare no competing interests.

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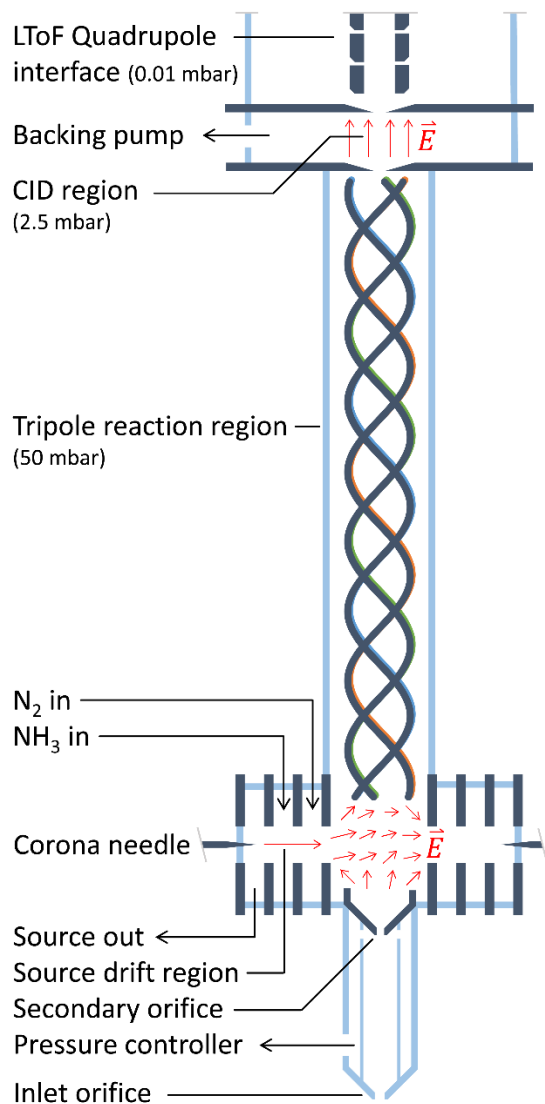


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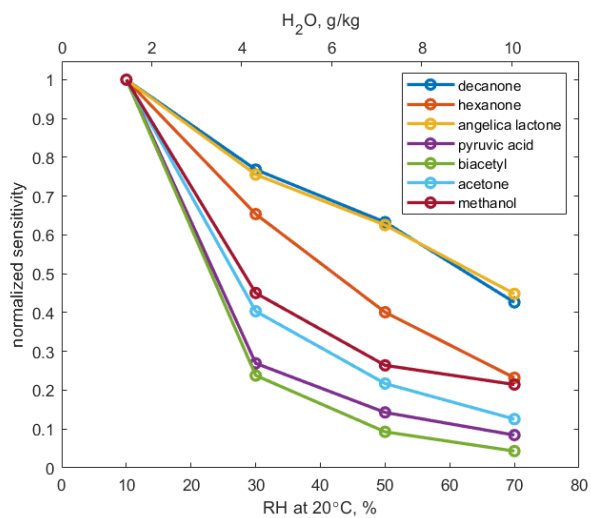


Species	Ion formula	m/z [Th]	Sensitivity		$2\sigma$ -LOD [pptv] (1s)	$V_{50}$ [V]	$KE_{cm\ 50}$ [eV]
			[cps/ppb]	[dcps/ppb]			
methanol	$CH_3ONH_4^+$	50.06	59	83	62	27.8	0.091
acetonitrile	$C_2H_3NNH_4^+$	59.0604	9700	12600	6	34.5	0.120
acetone	$C_3H_6ONH_4^+$	76.0757	21400	24600	1.83	36.4	0.129
acetic acid	$C_2H_4O_2NH_4^+$	78.055	1890	2140	66	31.4	0.105
isopropanol	$C_3H_8ONH_4^+$	78.0913	1100	1240	15	36.5	0.131
MVK	$C_4H_6ONH_4^+$	88.0757	27900	29700	13	36.9	0.131
MEK	$C_4H_8ONH_4^+$	90.0913	39300	41400	5	37.8	0.136
hydroxyacetone	$C_3H_6O_2NH_4^+$	92.0706	17600	18300	9	35.8	0.126
furanone	$C_4H_4O_2NH_4^+$	102.055	64000	63400	38	40.3	0.149
biacetyl	$C_4H_6O_2NH_4^+$	104.0706	3490	3420	23	36.6	0.130
pyruvic acid	$C_3H_4O_3NH_4^+$	106.0499	1650	1600	35	34.8	0.122
angelica lactone	$C_5H_6O_2NH_4^+$	116.0706	65500	60800	0.57	39.6	0.145
hexanone	$C_6H_{12}ONH_4^+$	118.1226	59000	54300	5	41.5	0.155
benzaldehyde	$C_7H_6ONH_4^+$	124.0757	43200	38800	1.35	36.7	0.130
heptanol	$C_7H_{16}ONH_4^+$	134.1539	12150	10500	1.52	39.5	0.144
decanone	$C_{10}H_{20}ONH_4^+$	174.1852	89400	67800	1.66	47.1	0.189

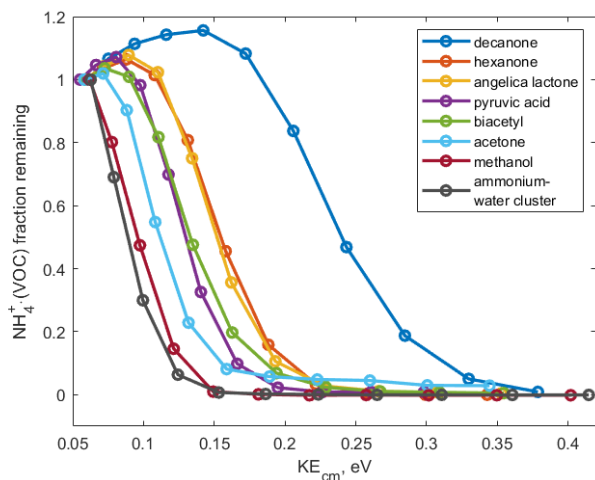
**Table 1: Sensitivities and detection limits of  $NH_4^+$ -CIMS for various VOC species; voltage ( $V_{50}$ ) and corresponding kinetic energy ( $KE_{cm\ 50}$ ) at which half of the ions have dissociated.**



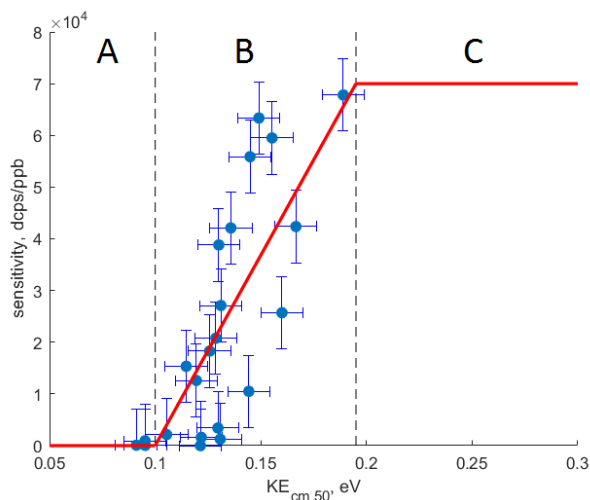
**Figure 1: Schematic drawing of the NH<sub>4</sub><sup>+</sup>-CIMS.**



**Figure 2: Humidity dependence curves for the normalized signals relative to the dryer conditions.**



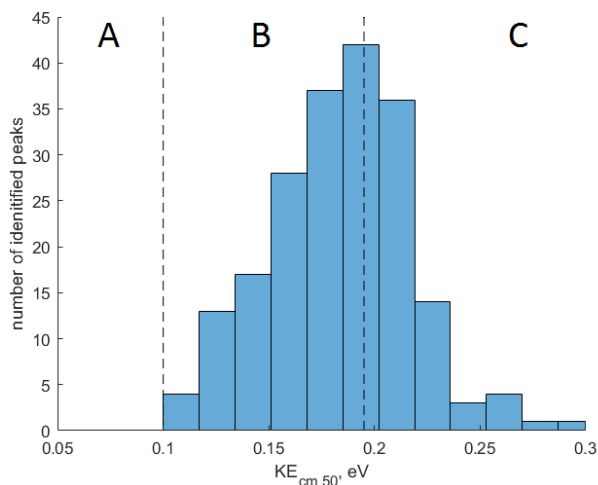
**Figure 3:** Declustering scans of ammonium-organic clusters  $\text{NH}_4^+(\text{VOC})$  for calibrated components and  $\text{NH}_4^+(\text{H}_2\text{O})$  reagent ions.



**Figure 4: The relationship between calculated kinetic energy of the ammonium-organic clusters  $KE_{cm\ 50}$  and measured sensitivity for calibrated compounds. Molecules characterized by  $KE_{cm\ 50}$  smaller than 0.10 eV (region A) cannot be detected by  $NH_4^+$ -CIMS; for molecules characterized by  $KE_{cm\ 50}$  between 0.10 and 0.19 eV (region B) a linear relationship between  $KE_{cm\ 50}$  and measured sensitivity is observed; molecules characterized by  $KE_{cm\ 50}$  greater than 0.19 eV (region C) are detected at the “kinetic sensitivity”.**

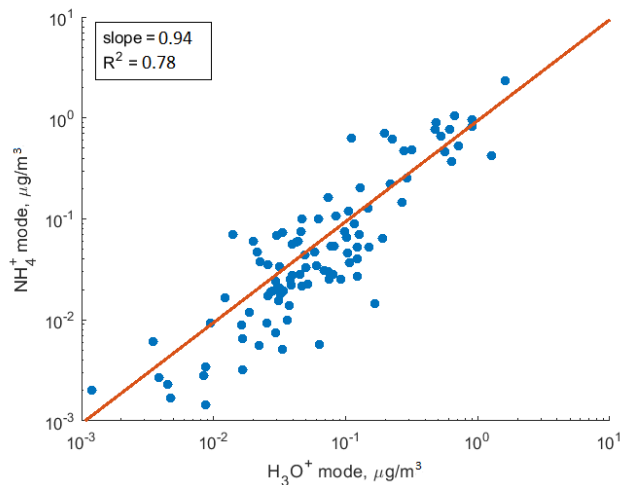
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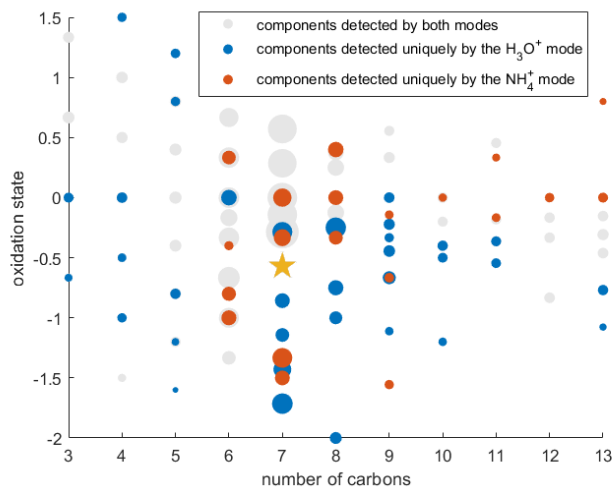


**Figure 5: Application of the collision-induced dissociation techniques for measurement of SOA composition produced during photooxidation of 3-methylcatechol in a laboratory experiment. 202 peaks are detected in  $\text{NH}_4^+$  mode and binned based on their  $\text{KE}_{\text{cm } 50}$ . Molecules with  $\text{KE}_{\text{cm } 50}$  smaller than 0.10 eV cannot be detected by  $\text{NH}_4^+$ -CIMS (region A); sensitivities of molecules characterized by  $\text{KE}_{\text{cm } 50}$  between 0.10 and 0.19 eV (region B) can be calculated using the linear fit presented in Fig 4; molecules with  $\text{KE}_{\text{cm } 50}$  greater than 0.19 eV are detected at the “kinetic sensitivity” (region C).**

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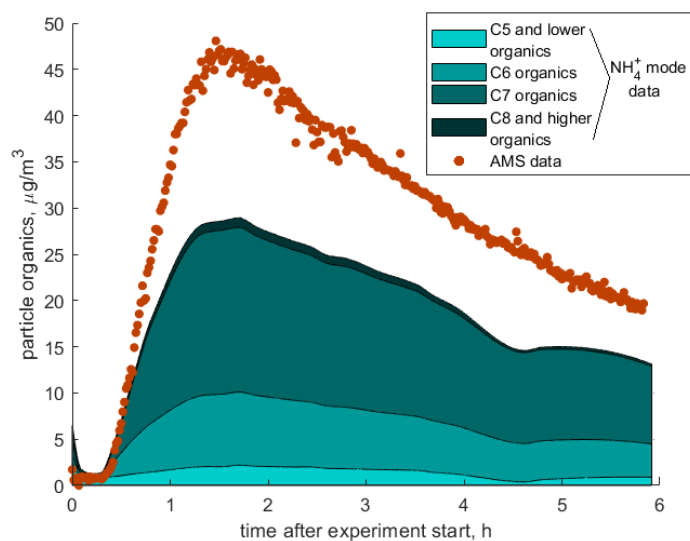


**Figure 6:** Comparison of volume mixing ratios of SOA components detected by the CIMS instrument in both  $\text{H}_3\text{O}^+$  and  $\text{NH}_4^+$  modes in the photooxidation experiment of 3-methylcatechol.



**Figure 7: Identified SOA components detected in both  $\text{H}_3\text{O}^+$  and  $\text{NH}_4^+$  modes (125 peaks), uniquely in the  $\text{H}_3\text{O}^+$  mode (34 peaks), and uniquely in  $\text{NH}_4^+$  mode (17 peaks) plotted on the  $n_{\text{C}}-\overline{\text{OS}}_{\text{C}}$  diagram. The gold star corresponds to the precursor of the photooxidation experiment, 3-methylcatechol. The size of the dots is proportional to the logarithm of the volume mixing ratio of each compound produced at the end of the experiment.**

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**Figure 8:** SOA produced during photooxidation of 3-methylcatechol in a laboratory experiment. The total organic aerosol mass is measured by AMS. OVOC detected by  $\text{NH}_4$ -CIMS are binned in four groups.