System for $\delta^{13}$C-$\text{CO}_2$ and $x\text{CO}_2$ analysis of discrete gas samples by cavity ring-down spectroscopy

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Abstract. A method was devised for analysing small discrete gas samples (50 ml syringe) by cavity ring-down spectroscopy (CRDS). Measurements were accomplished by inletting 50 ml syringed samples into an isotopic-$\text{CO}_2$ CRDS analyser (Picarro G2131-i) between baseline readings of a standard reference air, which produced sharp peaks in the CRDS data feed. A custom software script was developed to manage the measurement process and aggregate sample data in real-time. The method was successfully tested with $\text{CO}_2$ mole fractions ($x\text{CO}_2$) ranging from <0.1 to >20000 ppm and $\delta^{13}$C-$\text{CO}_2$ values from $-100$ up to $+30000$ ‰ vs VPDB. Throughput was typically 10 samples h$^{-1}$, with $\leq 3$ h$^{-1}$ possible under ideal conditions. The measurement failure rate in routine use was ca. 1 %. Calibration to correct for memory effects was performed with gravimetric gas standards ranging from 0.05 to 2109 ppm $x\text{CO}_2$ and $\delta^{13}$C-$\text{CO}_2$ levels varying from -27.3 to +21740 ‰. Repeatability tests demonstrated that method precision for 50 ml samples was ca. 0.05 ‰ in $x\text{CO}_2$ and 0.15 ‰ in $\delta^{13}$C-$\text{CO}_2$ for $\text{CO}_2$ compositions from 300 to 2000 ppm with natural abundance $^{13}$C. Long-term method consistency was tested over a 9-month period, with results showing no systematic measurement drift over time. Standardised analysis of discrete gas samples expands the scope of applications for isotopic-$\text{CO}_2$ CRDS and enhances its potential for replacing conventional isotope ratio measurement techniques. Our method involves minimal set-up costs and can be readily implemented in Picarro G2131-i and G2201-i analysers or tailored for use with other CRDS instruments and trace-gases.
1 Introduction

Cavity Ring-Down Spectroscopy (CRDS) is a high-sensitivity laser absorption technology becoming increasingly common for trace-gas analysis (Wang et al., 2008). As well as returning high-resolution mole fraction measurements (Crosson, 2008), CRDS is used for stable isotope analysis of CO₂, CH₄, H₂O, and N₂O (Crosson et al., 2002; Dahnke et al., 2001; Kerstel et al., 2006; Sigrist et al., 2008). Commercial deployment of CRDS has created novel analytical possibilities with greater stability, precision, instrument portability, and a lower cost-basis compared with many traditional spectroscopic, chromatographic, and mass spectrometric techniques (Berryman et al., 2011; Hancock and Orr-Ewing, 2010; Mürtz and Hering, 2010; Picarro, 2009).

Crosson et al. (2002) provide a description of the working principles for making isotopic measurements by CRDS. Commonly used in atmospheric research, isotopic CRDS gas analysers are normally on-line instruments whereby sample gas is continuously pumped through an optical cavity. While such continuous measurement systems are useful for monitoring applications, technical adaption is necessary for routine handling of small discrete gas samples. Commercial add-on modules are available for this purpose (McAlexander et al., 2010; Picarro, 2013), but these are unable to match the rapidity of conventional methods like gas chromatography (GC) and isotope ratio mass spectrometry (IRMS).

CRDS analysis with discrete sample throughput and handling comparable to IRMS could significantly improve a variety of empirical research. For example, simultaneous high-precision isotope ratio and mole fraction measurements from isotopic- CO₂ CRDS will reduce empirical workload and increase accuracy of CO₂ flux partitioning calculations in soil and plant respiration experiments (Midwood and Millard, 2011; Snell et al., 2014). However, realising these benefits requires regular batch analysis of discrete samples – existing arrangements that couple CRDS instruments directly to soil headspace chambers are generally constrained to measuring just one experiment at a time (Albanito et al., 2012; Bai et al., 2011; Midwood et al., 2008).

Berryman et al. (2011) describe a syringe sample delivery system for isotope ratio CRDS that allows small air samples (20 to 30 ml) to be analysed. In their method, the optical cavity of the CRDS analyser is flushed and completely evacuated prior to direct sample injection to ensure consistency and prevent sample-to-sample contamination. Although an important technical innovation with handling and cost advantages over IRMS, the setup is limited by slow sample turnover rates (3 h⁻¹).

In this paper, we present a new method for measuring discrete syringed gas samples (50 ml) by CRDS. Like Berryman et al. (2011), this method was conceived for isotopic-CO₂ CRDS to provide δ¹³C-CO₂ and CO₂ mole fraction (xCO₂) analysis in
soil respiration studies, but remains general enough to be used in other contexts and adjusted for other gas species. Instead of evacuating the cavity prior to sample introduction, our process intersperses samples against background measurements of a fixed reference air and post-corrects for bias in the measurements. This results in considerably faster throughput for typical atmospheric samples (up to 13 h⁻¹) than the method of Berryman et al. (2011). Additionally, with precision and discrete sample measurement rates comparable to automated continuous-flow IRMS, this method further advances CRDS as an attractive alternative for trace-gas isotope analysis.

2 Materials and methods

2.1 Analyser and sampling system

The CRDS instrument adapted for discrete sample measurement was a Picarro G2131-i isotopic CO₂ gas analyser (Picarro Inc., Santa Clara, CA, USA). Detailed description of the operation and spectroscopy of the G2131-i and predecessor units can be found in Dickinson et al. (2017), Hoffnagle (2015), Rella (2010a, 2010b, 2010c), and Wahl et al. (2006). In brief, sample air is circulated through a high-reflectivity optical cavity (35 cm³) at an inlet flow rate of ca. 25 ml min⁻¹ (NTP). Internal controls maintain the cavity at 318.15 ± 0.02 K and 18.67 ± 0.02 kPa. Spectroscopic ring-down time constants are measured across spectral bands of ¹³C¹⁶O₂ and ¹²C¹⁶O₂ to determine optical absorption peak heights, which are computed into ¹³C/¹²C isotope ratio and CO₂ mole fraction data (Hoffnagle, 2015). Spectral lines of ¹³C¹H₄ and ¹H₂¹⁶O are also measured for correcting direct and indirect spectral interferences from H₂O and CH₄ on the CO₂ bands. The normal measurement range for the G2131-i is set at 380 to 2000 ppm x CO₂ and natural abundance to +5000 ‰ in δ¹³C-CO₂ (Picarro, 2011).

All measurements made by the G2131-i are continually recorded at a rate of ca. 0.8 Hz; specific data must be extracted from log files for further treatment. Although discrete sample measurement is thus possible without special provision – by inleting the G2131-i with 200 to 300 ml of sample from a gasbag or chamber and retrieving the relevant data (Picarro, 2012) – such procedure is time inefficient and prone to errors from operator inconsistency. Furthermore, in many research settings it is impractical or impossible to gather such large samples (e.g. headspace chamber analyses). By instead applying a controlled procedure for inleting smaller volumes and software to automatically process the raw data in real-time, a more feasible method of discrete sample measurement was created.

A schematic of our measurement set-up is shown in Fig. 1. The system was simple in construction and concept: hermetic sample collection and delivery was achieved by high-quality gas-tight syringe with push-button valve and Luer lock fitting (50 ml, SGE Analytical Sci., Australia). A low permeability multi-layer foil gasbag (27 L Plastigas, Linde AG, Germany) functioned as a reservoir for a standard reference air, which was analysed between individual samples so as to give a ‘baseline’ for accurate data delineation. The large, non-pressurised volume of the reservoir meant pressure induced mixing
and back-flow risks were excluded, and allowed prolonged operation before refilling (>15 h). Gas-proof fluorinated-ethylene-propylene (FEP) tubing (Rotilabo, Carl Roth GmbH, Germany), Luer lock fittings, and Luer lock 3-way valves completed the set-up. All permanent tube fittings and joints were adhered with Locite 406/770 (Henkel AG, Germany) to ensure robustness and prevent leakage. The FEP tubing between the syringe sample inlet point and the CRDS inlet port (Fig. 1) was minimised (1/4” OD × 44 cm, connected to the 1/2” CRDS inlet port with reducing ferrule) to decrease mixing and lag time between sample delivery and measurement.

2.2 Sample measurement

The G2131-i and discrete sample measurement system were installed in an environmentally controlled laboratory (20 °C) to ensure stable operation. Syringed sample measurement was performed as follows: After instrument start-up and commencement of normal function, reference air measurement was initiated to establish stable baselines of xCO2 and δ13C-CO2. When a sample was ready for analysis, the syringe was connected to the sample inlet point (Fig. 1), and the 3-way valve manually actuated to stop the flow of reference air and supply the sample directly to the analyser. Upon opening the syringe valve the gas sample was drawn into the G2131-i, causing steady, unassisted collapse of the syringe plunger. Sample evacuation was completed in ca. 2.5 min, after which the sample inlet point valve was immediately reset and reference air intake resumed. Once CO2 and δ13C-CO2 readings had returned to initial baseline levels (thereby safeguarding against sample-to-sample carryover), the process was repeated for the next sample. In this way, reference air readings were punctuated by syringe samples to create ‘peaks’ in the raw data output with a sample-to-sample time of ca. 5 min (Fig. 2).

The gas aliquot size for all measurements was nominally 50 ml NTP. (Analysis of smaller amounts may be possible but 50 ml was assessed as a minimum for reliable operation. Samples larger than 50 ml would be easily handled, although adjustment of peak truncation parameters and re-calibration may be necessary for accurate performance – see below and Sect. 2.3.)

To achieve unambiguous sample peak identification, distinction in CO2 was required between reference air and sample. In practice this meant a relative change of ca. 2 % in xCO2 or ca. 5 % in δ13C-CO2. However, very large differences resulted in slower sample turnover (see Sect. 3.1). Best throughput was obtained using reference air that was similar to samples in xCO2 but contrasting in δ13C-CO2 (e.g. 15 % difference). In this work, dry standard air with 496 ppm xCO2 and -36.1‰ δ13C-CO2 was used as the reference for all formal measurements (NA1, Table 1).

While sample measurement was performed manually (i.e. syringe connection and disconnection, valve operation etc.), to ensure method consistency we composed a custom computer software script to manage the process in real-time (script available in the Supplement). Running through the built-in ‘Coordinator’ software program of the G2131-i, our script prompted the user for correct timing of sample introduction, detected and extracted sample peak data, monitored reference air values, filtered problem measures, and recorded measurement results. The software script isolated individual samples
from the CRDS data-stream by using specific events and timings in the measurement process as cues (e.g. a basic peak recognition algorithm; Fig. 3a). Prior to introduction of a sample, a reference air baseline was recorded for 30 s and averaged. Sample detection (trigger) then occurred when \( x\text{CO}_2 \) or \( \delta^{13}\text{C-CO}_2 \) values deviated from the baseline beyond a fixed threshold (default: 0.5 % of \( x\text{CO}_2 \) or 2 % in \( \delta^{13}\text{C-CO}_2 \)). The sample end (detrigger) was detected when measures returned halfway to baseline values (Fig. 3b). By truncating the sample peak data +80 s from the trigger and -29 s from the detrigger, ca. 30 s of representative measurement data was obtained for each sample (Fig. 3a). Upon completion of a sample measurement, the script computed means and standard deviations (SDs) of all data elements reported by the G2131-i (i.e. \( x\text{H}_2\text{O} \) and \( x\text{CH}_4 \) values together with \( x\text{CO}_2 \) and \( \delta^{13}\text{C-CO}_2 \)). These statistics were compiled along with corresponding baseline measures, timestamped, assigned sample descriptors, and then outputted into a concise results file (see example in the Supplement). After each detrigger event the software monitored CRDS readings for return to initial reference air baseline values before directing the operator to proceed with the next sample.

In addition to the G2131-i analyser, our method was successfully trialled on a sister CRDS instrument (Picarro G2201-i). The G2201-i differs from the G2131-i only in additionally measuring \( \delta^{10}\text{CH}_4 \) to give \( \delta^{13}\text{C-CH}_4 \) data (Picarro, 2015). To assist method adoption, we supply software scripts customised for each instrument (Supplement). The scripts include provision for user-adjustment of peak identification and truncation parameters to suit individual set-ups. A short video recording of the system and measurement demonstration is also available (https://youtu.be/jqVFUO-EuCk).

### 2.3 Measurement calibration

As discussed in studies by Gkinis et al. (2011) and Stowasser et al. (2012), stepwise changes to the inlet gas composition (as occur with discrete samples) do not give rise to correspondingly abrupt jumps in CRDS measurements, and instead result in sigmoid-shaped steps in the data (Fig. 3b). These smoothed transitions are the combined result of (i) the rate of gas replenishment in the optical cavity (Stowasser et al., 2014), (ii) partial mixing (turbulence and diffusion) of gas compositions downstream of the sample inlet (Gkinis et al., 2011), and (iii) molecular sorption and desorption on internal surfaces of the cavity and inlet tubes (Friedrichs et al., 2010). Although ‘response times’ of CRDS instruments typically range 1 to 3 min (Picarro, 2011; Sumner et al., 2011), the actual time required for an optical cavity to completely transition to a new gas composition can be substantially longer. In testing the G2131-i, we observed remnants of previous gases persisting with asymptotical decline for as long as 40 min following very large shifts in \( \text{CO}_2 \) composition (e.g. \( \Delta x\text{CO}_2 \geq 10000 \text{ ppm} \) or \( |\Delta \delta^{13}\text{C-CO}_2| > 5000 \% \)). While the error caused by the residual gases may sometimes be relatively trivial, all measurements that occur prior to the cavity attaining equilibrium will experience these ‘memory effects’.

In the case of our 50 ml syringe samples, memory effects were clearly present, as evidenced by the asymptotic curvature in the data peaks (Fig. 2). This meant that reported measures of syringe samples were biased towards reference air compared to ‘true’ values that would be determined from measurements of indeterminately large sample volumes and monitoring for
asymptotic closure. Other researchers have mitigated memory effects by evacuating the optical cavity before sample introduction (Berryman et al., 2011), or through several replicate measurements (Gupta et al., 2009; Leffler and Welker, 2013). In this work however, we elected to post-correct for reference air contamination by calibrating our measurement method with bottled gas standards. More specifically, we compared discrete sample measurements of gas standards against measures of the same standards directly inlet to the G2131-i for prolonged periods (>1 h). Importantly, no attempt was made to calibrate the syringe measurements directly against the gravimetric values of the standards – we were only concerned with isolating the bias associated with syringe sampling and not with any inaccuracies internal to the instrument spectroscopy (calibration of which should be undertaken separately; see for instance Dickinson et al., 2017). In this way we prevented convolution of errors that might have occurred if combining multiple layers of corrections into one step.

To this end, seven gravimetric gas standards with wide variation in CO2 composition (0.05 to 2109 ppm \(x\text{CO}_2\), -27.3 to +21740 ‰ \(\delta^{13}\text{CO}_2\)) were used as fixed source calibrants (see Table 1; exact compositions detailed in Dickinson et al., 2017). Direct measurements were performed by inletting the bottled standards to the G2131-i for more than one hour to ensure the absence of memory effects before taking formal measures for 10 min (ca. 460 data points; averages reported in Table 1). Next, 50 ml syringe samples of the standards were taken directly from bottles (syringe was pre-flushed several times to preclude contamination) and measured using our method as outlined (ca. 8 sample of each standard, for 56 measures in total – dataset in the Supplement). Before further analysis, due to the high \(^{13}\text{C}\) abundance in several gas standards, all reported CO2 data were adjusted for accuracy by the formulae in Dickinson et al. (2017), thereby eliminating unaddressed interferences and calculation abnormalities in the internal spectroscopy of the G2131-i.

The relationship between syringe and bottle measurements was established by recognising that the data peaks generated by syringe samples could be approximated by generalised logistic curves (Fig. 3b; also Gkinis et al., 2011). From this, together with a constant aliquot size for all syringe measures, we were able to predict a simple linear scaling of syringe values:

\[
syringe = base + (bottle - base)/K
\]

(1)

where syringe refers to the measurement value obtained for a syringe sample of a gas standard, base to the baseline measurement of reference air prior to sample introduction, bottle to the direct bottle measurement of the same standard, and \(K\) is a dimensionless empirical constant.

While all \(\text{CO}_2\) data elements reported by the G2131-i exhibited reasonably similar sample peak geometry, the empirical constants for \(^{12}\text{CO}_2\) and \(^{13}\text{CO}_2\) were expected to differ due to (de) sorption and diffusion induced isotope fractionation during sample filling of the optical cavity. Further, theoretical gas mixing considerations entailed Eq. (1) would not consistently hold for \(^{13}\text{C}/^{12}\text{C}\) isotope ratio data (\(R_{\text{CO}_2}\)) where a simultaneous change in total \(\text{CO}_2\) also occurred. Consequently, only \(x^{12}\text{CO}_2\) and \(x^{13}\text{CO}_2\) data were explicitly calibrated, with \(R_{\text{CO}_2}\) being subsequently recalculated. (Moreover, only the dry mole
fraction data of $^{12}$CO$_2$ and $^{13}$CO$_2$ were used due to the high likelihood of different transition equalisation rates for CO$_2$ and H$_2$O. For explanation of dry and wet mole fraction data see: Hoffnagle, 2015; Rella, 2010a; Rella et al., 2013.) Accordingly, the following correction formulae were derived from Eq. (1):

\[
x^{12}\text{CO}_2(\text{corrected}) = x^{12}\text{CO}_2(\text{base}) + [x^{12}\text{CO}_2(\text{syringe}) - x^{12}\text{CO}_2(\text{base})] \cdot K_{C12}
\]

(2)

\[
x^{13}\text{CO}_2(\text{corrected}) = x^{13}\text{CO}_2(\text{base}) + [x^{13}\text{CO}_2(\text{syringe}) - x^{13}\text{CO}_2(\text{base})] \cdot K_{C13}
\]

(3)

Total-xCO$_2$, R$_{C}$, and $\delta^{13}$C-CO$_2$ data were then determined from the resulting corrected values of $x^{12}$CO$_2$ and $x^{13}$CO$_2$:

\[
x_{CO_2} = x^{12}\text{CO}_2(\text{corrected}) + x^{13}\text{CO}_2(\text{corrected})
\]

(4)

\[
R_{CO_2} = \frac{x^{12}\text{CO}_2(\text{corrected})}{x^{13}\text{CO}_2(\text{corrected})}
\]

(5)

\[
\delta^{13}\text{C}-\text{CO}_2 = \left(\frac{K_{C13}}{K_{C12}}\right) - 1 \cdot 1000 \%
\]

(6)

The correction constants, $K_{C12}$ and $K_{C13}$, were found through weighted least squares analysis (WLS) of Eqs. (2) and (3) with syringe and bottle measurements of gas standards as input data (i.e. reverse regression of Eq. 1; bottle measures substituting for the left-hand-sides of Eqs. 2 and 3). To increase statistical power, $R_{CO_2}$, and total-xCO$_2$ data from bottle measurements were also incorporated into the analysis with Eqs. (4) and (5), thereby forming an extended optimisation problem (n = 216). In a similar vein to the WLS approach used by both Dickinson et al. (2017) and Stowasser et al. (2014) for calibrating CRDS measures, residuals weights were taken as the reciprocals of the individual summed variances resulting from the SDs of each syringe sample and bottle measurement (see Supplement and Table 1). The WLS solution was determined in R (version 3.2.1; R Core Team, 2015) by general purpose optimisation using the L-BFGS-B algorithm (Zhu et al., 1997) to yield the best-fit correction constants for all available CO$_2$ mole fraction and $^{13}$C/$^{12}$C isotope ratio data.

2.4 Precision and consistency tests

The gas standards used for compensating memory effects in syringe sample measurements covered a wide span of CO$_2$ mole fractions and very high $\delta^{13}$C-CO$_2$ values. While this was necessary for ensuring calibration accuracy and applicability, because several of the standards contained CO$_2$ compositions beyond the normal operating range of the G2131-i, those data were inappropriate for drawing conclusions about measurement precision.

Precision of method was therefore evaluated by replicate measurements of a systematic set of CO$_2$ mixtures that better conformed to G2131-i specifications. Using gas standards as blending sources (Table 1; Dickinson et al., 2017), 20 unique mixes with varied CO$_2$ mole fractions (ca. 300, 600, 1000, 1500, 2000 ppm) and $\delta^{13}$C-CO$_2$ values (ca. -30, +800, +1750, +2700, +3600 %) were prepared into multi-layer foil gasbags (1000 ml Supel Inert, SigmaAldrich Corp., St. Louis, MO, USA). (The set of mixtures formed an orthogonal array of cross combinations of $x$CO$_2$ and $\delta^{13}$C-CO$_2$: cf. Fig. S1 in the
Supplement. Each mix was sampled and measured with the syringe method three times in succession, and results analyzed for inter- and intra-measurement variability.

Long-term consistency and reliability of our syringe method was assessed by periodic analysis of a standard air (NA2, Table 1) during the course of 9 months of routine instrument use. More than 200 measurements were conducted and results examined for precision and drift.

3 Results and discussion

3.1 System operation

Though somewhat labor intensive and requiring continual operator attention, the syringe sample measurement process was uncomplicated, reliable, and economical. Sample handling and CRDS operation was non-specialist in comparison to conventional IRMS. The method was flexible to CO₂ composition, successfully handling samples <0.1 to >20000 ppm xCO₂ and -100 to +30000 ‰ δ¹³C-CO₂. The only significant methodological constraint observed was a reduction in sample turnover rate for compositions greater than either 3000 ppm xCO₂ or +4000 ‰ δ¹³C-CO₂. This was because post-sample reference air measures took longer to return to pre-sample baselines due to memory effects, thereby extending the inter-sample period. Keeping CO₂ levels within G2131-i specifications resulted in a throughput of ca. 10 samples h⁻¹. Best measurement rates of 12 to 13 samples h⁻¹ occurred when sample CO₂ compositions neighboured the reference air (e.g. within ca. 100 ppm xCO₂ and ca. 20 ‰ δ¹³C-CO₂ of reference). These throughput rates are at least a 2-fold improvement over both the method of Berryman et al. (2011) and specialty peripheral devices (Picarro, 2013).

Following initial development, the syringe method was incorporated into our general laboratory practices and during the first year of implementation more than 10000 samples were measured. Despite intense instrument usage, we noticed no changes or adverse impacts on G2131-i function, although increased external pressure variations caused by frequent syringe evacuations may conceivably reduce mechanical lifetimes of optical cavity pressure control valves. Failures occurred in ca. 1 % of measurements, principally due to operator mistakes, but occasionally because of leakage in sample inlet valve, syringe fault, or complications from the peak identification algorithm for samples very similar to the reference air (see Sect. 2.2). Very rarely, minor instabilities in reference air readings caused false peak detections and baseline return problems, but such instances were usually identified by the software script and internally resolved.

Durability of the gas-tight syringes used for sample delivery was excellent, although regular monitoring and maintenance was important to ensure smooth sample evacuation during the measurement process. Excessive plunger friction led to significant ‘jumpiness’ in syringe collapse, which manifested as small pressure fluctuations to the optical cavity and increased measurement noise (evidenced by larger reported SDs). Careful cleaning and exact silicone lubrication was carried...
out every few hundred samples to ensure uniform plunger operation and prolongation of syringe life. Syringe push-button and sample inlet point valves also required periodic attention and were replaced as necessary to pre-empt leaks and breakages.

3.2 Correction of memory effects

From rearranging Eq. (1), the discrepancy between syringe and bottle measurements (syringe bias) was predicted to be proportional to the difference of the syringe value and reference air baseline (sample peak height):

\[(\text{syringe} - \text{bottle}) = (\text{syringe} - \text{base}) \cdot (1 - K)\]  

(7)

Comparing the actual syringe sample and bottle measurements of gas standards, we observed systematic memory effect bias that was indeed consistent with this postulated relationship (Fig. 4). WLS across all CO₂ data yielded estimates of 1.00341 for \(K_{C12}\) and 1.0040 for \(K_{C13}\), with a coefficient of determination (\(r^2\)) of 0.84 (weighted residuals) for the complete correction model. Standard errors for \(K_{C12}\) and \(K_{C13}\) estimates were respectively 0.00017 and 0.00014 (see confidence intervals in Fig. 4). The Pearson’s correlation coefficient between \(K_{C12}\) and \(K_{C13}\) estimates was 0.26. The observed divergence in correction constants for \(^{12}\text{CO}_2\) and \(^{13}\text{CO}_2\) was statistically significant (\(t\)-test: \(P < 0.001\)) with a larger memory effect present in \(^{13}\text{CO}_2\) measurements. This result corroborates the expectation of isotope fractionation occurring during gas equalisation in the CRDS optical cavity, putatively due to surface (de)sorption and diffusion phenomena.

Having determined \(K_{C12}\) and \(K_{C13}\), syringe CO₂ measurements can be adjusted for bias with Eqs. (2)–(6). Accuracy of these corrections is very good: The standard errors on \(K_{C12}\) and \(K_{C13}\) add uncertainty to \(x_{\text{CO}_2}\) and \(\delta^{13}\text{C-CO}_2\) data of less than 0.02% of the difference between the sample and baseline values. For typical atmospheric samples, this additional source of error is entirely negligible compared to the uncertainty deriving from measurement precision and gas standard analytical accuracy.

While the correction coefficients (\(K_{C12}\) and \(K_{C13}\)) found in this work are unique to our sampling equipment and G2131-i analyser, the equivalent calibration may be easily performed on replica setups. We provide a generic spreadsheet to post-correct syringe sample CO₂ data for any values of \(K_{C12}\) and \(K_{C13}\), and a template for simultaneously applying the syringe correction with the spectroscopic calibration strategy of Dickinson et al. (2017) for \(^{13}\text{C}\)-enriched samples (Supplement).

Although our work only addresses memory effect bias in CO₂ data, we are confident the same strategy (Eq. 1) is straightforwardly applicable to other gas species (and isotopes) that can be similarly analysed by syringed samples and CRDS (e.g. CH₄, H₂O, N₂O).

3.3 Measurement precision and consistency

Precision of CRDS data can be evaluated in several ways: The SD of a moving average is a common approach in continuous analyses of a dynamic source (e.g. the ambient atmosphere; Zellweger et al., 2016) while measures of homogenous gas...
sources can be assessed by the SD of replicated samples (e.g. Wang et al., 2013) or by the SD of aggregated data in a single long-duration measurement (e.g. ≥10 min; as in Sect. 2.3 for bottle measurements, also Pang et al., 2016; and Stowasser et al., 2014). For our case of 50 ml syringe samples, replicate tests provided a detailed account of precision throughout the normal operational CO₂ range of the G2131-i, quantified in terms of both internal variation in individual sample analyses (i.e. intra-sample SD of the ca. 30 s of CRDS data comprising each measure, see Sect. 2.2) and the statistical dispersion of repeated samples (i.e. inter-sample SD).

Figure 6 shows δ and variance from isotope ratio data were (i.e. precision in δCO₂ and δ¹³CO₂ measurements were mutually independent). Relative SDs for both isotopologues remained near constant at ≤0.05 % across the tested ranges however (Fig. 5c, d). Notably, the majority of intra-sample SDs for both x¹³CO₂ and x¹²CO₂ data were found to be in general agreement with counterpart inter-sample SDs (see trendlines in Fig. 5). This means that the SDs reported by our software script for δ¹³CO₂ and δ¹²CO₂ mole fractions in individual syringe sample measures will reasonably approximate the expected precision for replicated measurements of those samples.

On the other hand, inter- and intra-sample SDs in ¹³C/¹²C isotope ratio data were dependent on the δ¹³CO₂ and δ¹²CO₂ mole fraction, increasing with higher δ¹³CO₂ and lower xCO₂ (see Fig. S1a, b in the Supplement). The relative SDs of isotope ratio measurements were unaffected by δ¹³CO₂ and δ¹²CO₂ but steadily decreased with increasing xCO₂ – declining from between 0.07 and 0.04 % at 300 ppm xCO₂ to between 0.03 and 0.015 % at 2000 ppm (Fig. S1d). One exception was at natural abundance isotope ratios (δ¹³CO₂ < -30 ‰) where inter-sample relative SDs of RCO₂ were steady at ca. 0.015 % (i.e. 0.15 ‰) across the tested xCO₂, range (Fig. S1b). Notably, opposing CO₂ mole fraction data, intra-sample SDs of isotope ratio data were almost always greater than corresponding inter-sample SDs, which largely reflects the summation of variance from the δ¹³CO₂ and δ¹²CO₂ spectral measurements used to generate the ¹³C/¹²C ratios. Nevertheless, as with δ¹²CO₂ and δ¹³CO₂, the SD reported for δ¹³CO₂ in individual syringe sample measures may be used as a conservative proxy of δ¹³CO₂ replicate precision.

Consistency of the syringed sample method was established by long-term repeated analysis of a standard air (NA2, Table 1). Figure 6 shows δ¹³CO₂ and δ¹²CO₂ data from 200 measurements covering a 9-month period (dataset available in the Supplement). Calibrating each measure for memory effects inherent to the syringe sample method (i.e. by Eqs. 2–6) but without correcting for instrument errors, these measures averaged 1024.18 ppm in δ¹³CO₂ and -27.35 ‰ in δ¹²CO₂ with respective SDs of 0.50 ppm and 0.33 ‰. The latter SD is larger than the inter-sample SD found in replicate measure testing, (0.15 ‰, see above), possibly indicating the presence of instrument drift in the data in addition to random errors of repeated syringe sampling. However, because these data were only adjusted for systematic memory effects, they represent a simultaneous time-series test of instrument accuracy and methodological noise. While the separate components of error
In sum, despite the short CRDS analysis period for a syringe sample (ca. 30 s), and limited number of replicates in performance testing, achieved measurement precision was excellent. With our system and G2131-i analyser, replicate sample SDs of ≤0.05 % may be expected for $^{13}$CO$_2$ and $^{13}$CO$_2$ mole fraction data, while typical resolution in $^{13}$C-CO$_2$ measurements will be ca. 0.15 % at natural $^{13}$C abundance. Moreover, to a first approximation, similar precisions can be obtained from intra-sample SDs of single syringe sample measures. Our results are an improvement upon the 0.3 % ($\delta$CO$_2$) and 0.3 % ($^{13}$C-CO$_2$) precision attained by the system of Berryman et al. (2011), although this is likely due to the enhanced spectroscopic sensitivity of the G2131-i compared to the older G1101-i analyser used in their study. Additionally, our method delivers precision in $^{13}$C-CO$_2$ comparable to the Picarro SSIM2 discrete sample peripheral device (0.11 ‰; Picarro, 2013) and traditional continuous-flow IRMS (typically 0.1 ‰), which, by contrast, are single-purpose instruments that do not also report accurate CO$_2$ mole fraction measurements. And finally, although finer measurement resolution is possible with CRDS (e.g. by analysing larger gas aliquots or with continuous sampling; Pang et al., 2016), the uncertainties deriving from the precision of our discrete sample measures will be, in many cases, no worse than the typical tolerances on gravimetric gas standards used for instrument calibrations (e.g. Brewer et al., 2014; Dickinson et al., 2017). In such contexts, applying our method for isotopic and mole fraction analyses of trace-gases should generally not result in significantly poorer absolute accuracy compared to other sampling techniques (i.e. uncertainties on gas standards, rather than measurement precision, may limit overall accuracy).

### 3.4 Potential applications

At present, isotope ratio analysis of fixed trace-gas samples is usually achieved by IRMS interfaced to autosampling GC systems. Such instruments require specialised user training and carry high consumable costs however. Similarly capable CRDS-based techniques can avoid both these limitations and represent an advance in stable isotope analysis. Although not suitable for all sample types (e.g. non-atmospheric compositions of background gases; Friedrichs et al., 2010), adapting the present generation of CRDS gas analysers for rapid discrete sample measurement has promising application in contexts where syringe or flask sampling is frequently performed – especially where accurate gas mole fraction data is also valuable.
such as in ecosystem respiration and emission studies (e.g. Zeeman et al., 2008), analysing dissolved gases in terrestrial waters (Hope et al., 1995; Loose et al., 2009), and certain instances of trapped air in ice-cores (e.g. Sowers et al., 2005).

A specific example where our method has immediate relevance is in measuring CO₂ respiration in soil microcosm headspace studies. To date, applying CRDS gas analysers to such research is mostly achieved through closed-loop recirculation (Christiansen et al., 2015; Ramlow and Cotrufo, 2017) or continuous analysis of open chamber systems (Bai et al., 2011; Jassal et al., 2016). Apart from cost and complexity, these solutions typically restrict the number of experiments that can be concurrently measured by a single instrument. Our system significantly eases this constraint however. For instance, assuming a sample turnover of 10 h⁻¹ and conducting four syringed headspace measures per microcosm over the course of a 10 h workday, it is feasible to use one analyser for measuring daily respiration rates in 25 simultaneous experiments. Further, where CO₂ flux partitioning by isotopic analysis is undertaken, achieving sample measurement precision of ca. 0.05 ‰ in xCO₂ and ca. 0.15 ‰ in δ¹³C-CO₂ means that the resulting uncertainties on efflux partitions will be comparable (if not smaller) to those in studies using infrared gas analysis and IRMS or IRMS alone (compare Joos et al., 2008 and Munksgaard et al., 2013).

The primary drawbacks of employing our method for isotopic-CO₂ measurement of discrete samples compared to an automated GC-IRMS system are (i) the larger sampling size, (ii) a more constrained operational xCO₂ range, and (iii) the necessity of near-continuous operator presence at the instrument. However, implementation of smaller volume CRDS optical cavities (8.5 cm³) could dramatically decrease the required sample amount and allow even shorter measurement times (e.g. Stowasser et al., 2014), while dilution methods and calibration can expand the xCO₂ measurement range of CRDS. Similarly, methodological refinement to integrate automated syringe sampling and valve systems would curtail labour requirements.

4 Conclusions and outlook

Discrete sample analysis of trace-gases by CRDS is possible through basic instrument adaptation. We have set forth a scheme for xCO₂ and δ¹³C-CO₂ determination of 50 ml syringed samples on a Picarro G2131-i isotopic-CO₂ analyser. With software to manage the measurement process and compute results data, our method offers substantially faster analysis of small gas volumes with equal or better precision than comparable set-ups. Memory effects present in syringe sample measurements can be accurately compensated by calibration against large-volume measures of gravimetric gas standards.

Although CRDS is gaining scientific acceptance for isotopic-CO₂ measurement, so far the technology has not seriously challenged IRMS in discrete gas sample analysis, despite lower running and capital costs, simpler operation, less measurement drift, and the added benefit of providing more accurate xCO₂ data concurrently with δ¹³C-CO₂. In achieving
similar precision and sample throughput to IRMS, our syringe sample method helps position CRDS as a tenable competitor for isotopic analysis of discrete samples.

The chief disadvantages of our process compared to IRMS for isotopic-CO$_2$ analysis are a narrower $\delta^{13}$CO$_2$ performance range, higher labour demands, and a comparatively large sample size (50 ml NTP). Method improvements towards automation may greatly ease user workload however, and the development of smaller optical cavities could reduce the sample gas needed for discrete analysis on future CRDS analysers as well as increasing sample throughput rates even further.

This system can be applied with any Picarro G2131-i or G2201-i CRDS analyser, though calibration and tuning of parameters in the software script may be necessary to account for variations in set-up, sample volume (and pressure), and reference air composition. Implementation on other CRDS instruments and conversion for measurements of other trace gases are anticipated with only minor software amendments.

Supplement items

- Fig. S1. Precision in syringe sample $^{13}$C/$^{12}$C isotope ratio data
- Discrete sampling software scripts for Picarro G2131-i and G2201-i analysers
- Example_discrete_sample_data_output.csv
- Measurement_data.xlsx
- Templates for bias correction (2)

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References


McAlexander, W. I., Fellers, R., Owano, T. G., and Baer, D. S.: Carbon isotope analysis of discrete CO$_2$ samples ranging from 300 ppmv to 100% using cavity enhanced laser absorption, European Geophysical Union General Assembly, Vienna, Austria, 2010.


Figure 1: Schematic diagram of the discrete gas sample measurement system coupled to the isotopic-CO₂ CRDS analyser.
Figure 2: Example CRDS data feed for syringe samples. Reference air measurements (ca. 425 ppm $^{12}$CO$_2$ and -37 ‰ $\delta^{13}$C-CO$_2$) are interrupted by successive samples to form consistently identifiable peaks in the data.
Figure 3: (a) Example of raw G2131-i measurement data and breakdown of events during analysis of a 50 ml syringe sample. Blue segments are truncated from the sample peak by our software script while red segments are the extracted measurement data. All timings and thresholds are user-customisable in the software for variation in sample size and equipment. (b) The most reliable sample end time (detrigger) was established as the point when measures returned to half the difference between peak-maximum (or minimum in the case of samples with lower xCO₂ than reference air) and the baseline value. Grey lines are amplitude-normalised tailing segments from 23 test samples varying in xCO₂. The broken green curve denotes a generalised logistic function fit to these test data by non-linear least squares optimisation. Solving the fitted function determined that 29 ± 2 s elapsed between peak-maximum and half-maximum irrespective of sample composition.
Figure 4: Discrepancies between syringe sample and direct bottle measurements (syringe bias) of gas standards as a function of the syringe sample peak height (Eq. 7) for (a) $^{12}$CO$_2$ and (b) $^{13}$CO$_2$. The WLS fitted linear models (see Sect. 2.3) are overlaid for comparison (solid lines; slopes = $1 - K$, Eq. 7), with 95 % confidence intervals (shaded) and 95 % prediction intervals (dashed lines) as determined from the standard error estimates of $K_{C12}$ and $K_{C13}$ (Sect. 3.2).
Figure 5: Precision in syringe sample data for $^{12}$CO$_2$ (left: a, b) and $^{13}$CO$_2$ (right: c, d) as quantified by standard deviations (top: a, c) and relative standard deviations (bottom: b, d) for individual measures (red) and replicate measurements (blue).
Figure 6: Repeated syringe sample measurements in (a) $x^2\text{CO}_2$ and (b) $\delta^{13}\text{C}-\text{CO}_2$ of standard gas NA2 (Table 1) over a 9-month period ($n = 200$). Error bars denote ±1 intra-sample SD of each individual measure. Grand means are the solid black horizontal lines with dotted lines indicating ±1 SD of all measurements. 10-sample moving averages are shown in red. Histograms inset right depict cumulative distributions of syringe measures. Blue dashed lines indicate the direct bottle measurement of NA2 with blue shaded areas covering ±1 SD of the bottle measure.
Figure S1: Precision in syringe sample $^{13}$C/$^{12}$C isotope ratio data ($\delta^{13}$C, $\delta^{12}$C) for individual measures (red) and replicate measurements (blue). (a) Standard deviations and (b) relative standard deviations as a function of total CO$_2$ mole fraction and grouped by $\delta^{13}$C-CO$_2$. (c) Standard deviations and (d) relative standard deviations as a function of $\delta^{13}$C-CO$_2$ and grouped by total xCO$_2$. 
<table>
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<th>Standard ID</th>
<th>$x^{13}$CO$_2$ (ppm)</th>
<th>$x^{18}$CO$_2$ (ppm)</th>
<th>$x$CO$_2$ (ppm)</th>
<th>$R_{CO_2}$ ($\delta^{13}$CO$_2$/12CO$_2$)</th>
<th>$\delta^{13}$C-CO$_2$ (‰)</th>
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<tr>
<td>NA1 (Ref. air)</td>
<td>490.55 (0.13)</td>
<td>5.286 (0.004)</td>
<td>495.84 (0.13)</td>
<td>1.0776 (0.0006)</td>
<td>-36.14 (0.57)</td>
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<td>NA2</td>
<td>1024.26 (0.21)</td>
<td>11.137 (0.004)</td>
<td>1035.39 (0.21)</td>
<td>1.0874 (0.0003)</td>
<td>-27.43 (0.28)</td>
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<td>ZERO</td>
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<td>0.004 (0.004)</td>
<td>0.05 (0.04)</td>
<td>-</td>
<td>-</td>
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<td>HE1</td>
<td>2028.98 (0.47)</td>
<td>25.528 (0.007)</td>
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<td>1.2582 (0.0004)</td>
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<td>HE2</td>
<td>2009.15 (0.53)</td>
<td>100.11 (0.02)</td>
<td>2109.26 (0.53)</td>
<td>4.983 (0.001)</td>
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<td>TT</td>
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<td>30.216 (0.008)</td>
<td>1032.40 (0.22)</td>
<td>5.011 (0.001)</td>
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<td>LE1</td>
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<td>25.249 (0.005)</td>
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<td>LE2</td>
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<td>101.24 (0.01)</td>
<td>499.45 (0.16)</td>
<td>25.42 (0.01)</td>
<td>+21739 (9)</td>
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</tbody>
</table>

$R_{CO_2}$ data are scaled by 10$^2$ for ease of comprehension.

$\delta^{13}$C-CO$_2$ values are reported against VPDB (Werner and Brand, 2001).

Table 1: Bottle measurement data of the standard air used as baseline for syringe sample measures (NA1) and the gas standards used in method calibration (NA2 through LE2). Values are the averages (SDs in parentheses) of 10 min measurements taken for each standard directly inlet to the CRDS analyser (see Sect. 2.3). Data have been post-corrected as per the calibration of Dickinson et al. (2017).