

## **Author response to anonymous referee #2 on “A new method for atmospheric detection of the CH<sub>3</sub>O<sub>2</sub> radical” by L. Onel et al.**

Note: The changes in the manuscript addressing the comments of the referee #2 are highlighted in yellow below. The authors refer to the line numbers in the manuscript before revision mentioned in the comments.

The authors would like to thank anonymous referee #2 for their valuable comments to this manuscript.

The first two questions (1/ and 2/) address the quenching of the CH<sub>3</sub>O fluorescence by water vapour and methane, respectively:

*1/ The sensitivity of conventional FAGE instruments is known to be dependent on the ambient water concentration due to the quenching of excited OH radicals by water molecules. This matrix effect is taken into account through the calibration of the OH sensitivity at different water-vapor concentrations. Can excited CH<sub>3</sub>O radicals also be quenched by water vapor? If so, what is the implication for ambient measurements of CH<sub>3</sub>O<sub>2</sub>?*

*2/ For calibration purposes, CH<sub>3</sub>O<sub>2</sub> is generated using the water-photolysis approach by adding an excess of methane in the photolysis cell. Could the authors comment on the potential quenching of excited CH<sub>3</sub>O by methane during calibration experiments?*

No measurement of the rate coefficients of the fluorescence quenching by the traces gases has been performed in this work. However, a very good agreement was obtained between the flow tube calibrations for CH<sub>3</sub>O<sub>2</sub> with two different concentrations of water vapour in the flow tube:  $7.5 \times 10^{16}$  molecule cm<sup>-3</sup> or  $3 \times 10^{17}$  molecule cm<sup>-3</sup> (corresponding to  $2.6 \times 10^{14}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{15}$  molecule cm<sup>-3</sup>, respectively in the FAGE detection cell) as shown by Figure 6 in Sect. 2.3.2.1. The result presented in Figure 6 shows that the CH<sub>3</sub>O fluorescence quenching rate by water is minor for the above [H<sub>2</sub>O].

Methane was also present in the FAGE chamber in concentrations of several times  $10^{14}$  molecule cm<sup>-3</sup>. Calculations using the CH<sub>3</sub>O fluorescence quenching rate coefficient of CH<sub>4</sub> reported by Wantuck et al. (1987),  $1.05 \times 10^{-10}$  s<sup>-1</sup>, and a pressure in the FAGE detection cell of 2.65 Torr show only minor decreases in the fluorescence quantum yield, by few percent, when [CH<sub>4</sub>] is increased from zero to the experimental values. Assuming a quenching rate coefficient of H<sub>2</sub>O equal to that of CH<sub>4</sub>, similar small decreases in the fluorescence quantum yield were computed when [H<sub>2</sub>O] was increased from zero to the concentration values used in the

flow tube calibration ( $0.3 - 1.0 \times 10^{15}$  molecule  $\text{cm}^{-3}$ ). Therefore, the effects of methane and water on the FAGE sensitivity for  $\text{CH}_3\text{O}_2$  are minimal.

A paragraph which discusses the  $\text{CH}_3\text{O(A)}$  quenching rates of water and methane at the concentrations used in the flow tube calibration of the FAGE instrument for  $\text{CH}_3\text{O}_2$  has been added at the end of the section 3.1.1:

“The calibrations using the flow-tube (“wand”) method have been performed under water vapour concentrations similar to the ambient  $[\text{H}_2\text{O}_{\text{vapour}}]$  but few orders of magnitude higher than those present in the HIRAC chamber experiments. In contrast with  $[\text{H}_2\text{O}_{\text{vapour}}]$  the methane concentrations used in the “wand” method were similar to  $[\text{CH}_4]$  present in HIRAC but higher than  $[\text{CH}_4]$  in the atmosphere. However, as detailed in this paragraph, the effects of methane and water on our sensitivity are minimal. Estimations using the reported fluorescence quenching rate coefficient of  $\text{CH}_3\text{O(A)}$  by  $\text{CH}_4$ ,  $k_{\text{quench.CH}_4} = 1.05 \times 10^{-10} \text{ s}^{-1}$ , (Wantuck et al., 1987) and the concentrations of  $\text{CH}_4$  in the LIF detection cell for the calibrations using the flow-tube ( $1.7 \times 10^{14}$  molecule  $\text{cm}^{-3}$  and  $3.4 \times 10^{14}$  molecule  $\text{cm}^{-3}$ , corresponding to  $5.0 \times 10^{16}$  molecule  $\text{cm}^{-3}$  and  $1.0 \times 10^{17}$  molecule  $\text{cm}^{-3}$ , respectively in the flow tube) resulted in only ~ 1–2% lower fluorescence quantum yield compared to the value determined in the absence of  $\text{CH}_4$ . No literature value has been found for the fluorescence rate coefficient of  $\text{CH}_3\text{O(A)}$  fluorescence by  $\text{H}_2\text{O}$  vapour. However, even if it assumed to be as large as the above reported value for  $\text{CH}_4$  ( $k_{\text{quench.CH}_4}$ ), only a few percent decrease in the fluorescence quantum yield is computed (compared with a water concentration of zero) for the levels of  $\text{H}_2\text{O}$  vapour which are present at the  $\text{CH}_3\text{O}_2$  FAGE detection axis when using the flow tube calibration method. These levels (1–2% v/v) are similar to a typical water vapour concentration in the atmosphere. A very good agreement has been obtained between the calibration factors for  $\text{CH}_3\text{O}_2$  detection with two different concentrations of water vapour in the flow tube:  $7.5 \times 10^{16}$  molecule  $\text{cm}^{-3}$  or  $3.0 \times 10^{17}$  molecule  $\text{cm}^{-3}$  (corresponding to  $2.6 \times 10^{14}$  molecule  $\text{cm}^{-3}$  and  $1.0 \times 10^{15}$  molecule  $\text{cm}^{-3}$ , respectively in the FAGE cell) as shown in Figure 6 in Sect. 2.3.2.1. This very good agreement for  $\text{H}_2\text{O}$  vapour and the above calculations for  $\text{CH}_4$  support the use of the flow tube method for the FAGE calibration of the  $\text{CH}_3\text{O}_2$  concentrations.”

## Minor comments

**P4 L4:** “Here we report he first ...” should read “Here we report the first ...”

The suggested correction has been made.

**P4 L13:** Please report the sampling flow rate of the FAGE apparatus

Now the sampling flow rate is given at the beginning of section 2.1 (page 4):

“The gas was sampled with a flow rate of 3.2 slm through a 1 mm diameter pinhole...”

**P5 L4-5:** Since the detection of the CH<sub>3</sub>O fluorescence is red-shifted from the excitation, why is the counting window delayed by 100 ns from the laser pulse? This time gating approach is usually used for the detection of on-resonant fluorescence.

The off-resonance CH<sub>3</sub>O fluorescence occurs between ~ 300 – 400 nm and, hence a relatively broad bandpass filter, with an average transmission > 80% between 320–430 nm, was used for the fluorescence collection. However, it appears that red-shifted scattered laser light (the excitation wavelength was ~ 298 nm) produced in the FAGE chamber also passed through the interference filter, increasing the background. In order to avoid the majority of these background counts, the gate unit was opened 100 ns after the probe light pulse. As the optimum gate-width found for the CH<sub>3</sub>O fluorescence was 2 μs (*vide infra*), no significant loss of CH<sub>3</sub>O signal was encountered by the 100 ns delay in the fluorescence detection. Future improvements to the instrument will improve changing the cell material or coating to reduce this scattered light background.

The second paragraph on page 5 was changed as follows:

“The relatively broad bandpass filter used for the collection of the CH<sub>3</sub>O fluorescence (average transmission > 80% between 320–430 nm) allowed some red-shifted scattered light (presumably from the walls of the chamber) generated by the probe laser to be transmitted and hence detected by the MCP-PMT. In order to ameliorate this and reduce the background signal, the gate unit was opened 100 ns after the laser pulse to detect fluorescence integrated over a gate-width of 2 μs. The optimum gate-width of 2 μs (values in the range 1-3 μs were compared) is consistent with the CH<sub>3</sub>O fluorescence lifetimes, calculated to be in the range of 0.9 – 1.5 μs, using the reported radiative lifetimes for CH<sub>3</sub>O of 1.5 μs (Inoue et al., 1979), 2.2 μs (Ebata et al., 1982) and (4 ± 2) μs (Wendt and Hunziker, 1979) and using the fluorescence quenching rate coefficients of N<sub>2</sub> and O<sub>2</sub> (Wantuck et al., 1987) to calculate the rate of quenching at the pressure in the FAGE detection cell ((2.65 ± 0.05) Torr). As the fluorescence lifetime of CH<sub>3</sub>O(A) in the detection cell is 0.9–1.5 μs, delaying the counting of the fluorescence by 100 ns makes very little difference (~ 10%) in the fraction of fluorescence collected.”

*P5 L12-15: The authors mention that the wavelength is tuned on/off resonance with the CH<sub>3</sub>O transition line. In FAGE instruments, OH is continuously generated in a reference cell to be able to precisely tune the laser wavelength on and off resonance. How is it performed for CH<sub>3</sub>O on this instrument? Is CH<sub>3</sub>O continuously generated in a reference cell? If so, how is it done?*

The signals were large enough that during conditions where CH<sub>3</sub>O<sub>2</sub> concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser-wavelength was stable over a long period once the laser wavelength had been tuned to the CH<sub>3</sub>O transition. Hence, the online wavelength position for CH<sub>3</sub>O fluorescence detection was found without using a reference cell. The laser excitation scans shown in Figures 2 and 3 were performed using the flow tube method described in the sections 2.3.1 and 2.3.2.1 to generate either CH<sub>3</sub>O (by the CH<sub>3</sub>OH photolysis at 185 nm) or CH<sub>3</sub>O<sub>2</sub> (by the H<sub>2</sub>O photolysis at 185 nm to generate OH followed by the reaction of the produced OH with CH<sub>4</sub> in the presence of O<sub>2</sub>).

In the HIRAC experiments the concentration of CH<sub>3</sub>O<sub>2</sub> radicals generated in the chamber in a steady-state with the UV lamps turned on at the beginning of each experiment using the Cl<sub>2</sub>/CH<sub>4</sub>/air system was used to tune the laser at the correct excitation wavelength by performing similar scans to the laser scans shown in Figure 3.

In all measurements the offline wavelength position was fixed to the value obtained by adding 2.5 nm to  $\lambda(\text{online})$  as described in the third paragraph on page 5. For field measurements in the future, when the concentrations of CH<sub>3</sub>O<sub>2</sub> (and hence CH<sub>3</sub>O after conversion) will be both lower and more variable over short timescales, a reference cell will be necessary. We are in the process of developing a reference cell.

The third paragraph on page 5 was changed to clarify how the laser is tuned to the correct CH<sub>3</sub>O excitation wavelength:

“...Figure 2 shows the laser excitation spectrum centred at ~298 nm in the  $\nu_3$  vibronic band recorded using an increment of  $\Delta\lambda = 10^{-3}$  nm. The spectrum agrees well with previous work (Inoue et al., 1980; Kappert and Temps, 1989; Shannon et al., 2013). Figure 3 shows typical laser excitation scans performed over a narrower range of wavelengths in order to locate  $\lambda(\text{online})$ . The LIF spectra were obtained by using the CH<sub>3</sub>O or CH<sub>3</sub>O<sub>2</sub> radicals generated in a flow tube described in Sect. 2.3.1, with the flow tube output impinged close to the FAGE sampling inlet. The radicals were generated using the 184.9 nm light output of a Hg Pen-Ray lamp by either the photolysis of methanol in nitrogen to generate CH<sub>3</sub>O or the photolysis of water vapour in synthetic air (to generate OH) in the presence of methane to form CH<sub>3</sub>O<sub>2</sub>. The CH<sub>3</sub>O radicals were directly detected, while the CH<sub>3</sub>O<sub>2</sub> radicals were first converted to CH<sub>3</sub>O species by added NO prior to the fluorescence

detection cell (Fig. 1). Similar laser scans to the scans shown in Fig. 3 were recorded by using the  $\text{CH}_3\text{O}_2$  radicals produced in a steady-state concentration in HIRAC using photolytic mixtures of  $\text{Cl}_2/\text{CH}_4/\text{air}$  as described in Sect. 2.3.2.2. There were no unexpected features in the laser excitation scans for  $\text{CH}_3\text{O}$  recorded when FAGE sampled  $\text{CH}_3\text{O}_2$  radicals from HIRAC, consistent with no interference being anticipated in the FAGE measurements of  $\text{CH}_3\text{O}$  as there were no other species in HIRAC absorbing at 298 nm and fluorescing at the wavelengths transmitted by the bandpass filter (average transmission > 80 % over 320 – 430 nm).

In this work the FAGE signals were large enough that during conditions where  $\text{CH}_3\text{O}_2$  concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser wavelength was stable over a long period once  $\lambda$  had been tuned to the  $\text{CH}_3\text{O}$  transition. Hence,  $\lambda(\text{online})$  was found without using a reference cell. We are in the process of developing a reference cell for field measurements in the future, when the concentrations of  $\text{CH}_3\text{O}_2$  (and hence  $\text{CH}_3\text{O}$  after conversion) will be both lower and more variable over short timescales.”

In addition, the future construction of the reference cell is mentioned in the paragraph of section 3.1.1 where all the future instrument improvements are listed:

“The further optimizations of sensitivity and the planned construction of a reference cell to find the online wavelength position could potentially enable  $\text{CH}_3\text{O}_2$  measurements to be made in urban environments where  $\text{CH}_3\text{O}_2$  concentrations are estimated to be considerably lower, for example a few  $10^7$  molecule  $\text{cm}^{-3}$  based on modeling results (Whalley et al., to be submitted).”

*P7 L15: The authors indicate a  $\text{CH}_3\text{O}_2$ -to- $\text{CH}_3\text{O}$  conversion efficiency of 40% at the optimum NO concentration. However, since  $\text{CH}_3\text{O}$  can also be lost through its reaction with NO (and potentially through its reaction with  $\text{O}_2$  as well), isn't the 40% representative of a lower limit of the conversion?*

The 40% value represents the optimum  $\text{CH}_3\text{O}_2$  to  $\text{CH}_3\text{O}$  conversion efficiency as  $\text{CH}_3\text{O}$  is rapidly formed (by the  $\text{CH}_3\text{O}_2 + \text{NO}$  reaction) and removed in the system (by the  $\text{CH}_3\text{O}$  reactions with NO and  $\text{O}_2$ ), as discussed in section 2.2 (page 7). The text in section 2.2 explains that this result was obtained by comparison of the FAGE signal vs. [NO] generated by numerical simulations using a chemistry system formed by the above reactions with experimental data. Therefore, no text change has been made as the value of 40% was obtained from a simulation at the relevant conditions.

*P10 L12-13: It is indicated that the photon flux was varied between 0-1.5E14 photon/cm2/s. However, the lower bound reported for the radical generation is 1.5E10 molecule/cm3, which cannot correspond to a photon flux set at zero. Please clarify.*

The lower limit of the photon flux was corrected:

“The concentration of CH<sub>3</sub>O<sub>2</sub> was varied by changing the photon flux in the range of 0.5–1.5 × 10<sup>14</sup> photon cm<sup>-2</sup> s<sup>-1</sup> to generate [CH<sub>3</sub>O<sub>2</sub>] = 1.5–4.5 × 10<sup>10</sup> molecule cm<sup>-3</sup>.”

*P12 L9-14: The detection limits are calculated for a BKG signal of approximately 100 ct/s, which is reported as a typical value for this instrument. What are the contributions of the scattered visible and laser lights? How is the BKG signal expected to change when the solar irradiation changes during field measurements? How will it affect the detection limit during daytime?*

The contributions to be background are roughly 50% laser scattered light within the detection cell and 50% visible light which enters the pinhole. For field measurements, there will be a contribution from solar scattered light which will scale with sunlight intensity. As for measurements of OH, the detection limit depends on the standard deviation of the background signal, and for more intense solar radiation, this will increase, increasing the detection limit. The visible scattered light is recorded on its own (together with dark counts) in a separate photon collection integration gate which is delayed a long time after the laser pulse, and is subtracted from the counts from the integration gate containing the fluorescence (after scaling for any differences in the two gate widths).

The second paragraph on page 12 was modified:

“...BKG is the background signal and had a typical value of ~100 counts s<sup>-1</sup>, which represents ~50 counts s<sup>-1</sup> laser scattered light within the detection cell and ~50 counts s<sup>-1</sup> scattered visible light which enters the pinhole from the room with a negligible contribution (1 count s<sup>-1</sup> on average) of the detector dark counts,  $t$  is the time per data point,  $m$  represents the number of online data points and  $n$  is the number of offline data points.”

**P13 L22:** caption Fig. 7. “ $cm^{-1}$ ” should read “ $cm^{-3}$ ”

“ $cm^{-1}$ ” was changed into “ $cm^{-3}$ ”

**P15 L21 & L22:** Two different uncertainties are given for the on-line signal: 12% and 6%. Which one is correct?

The paragraph on page 15 discusses the different components of the total uncertainty: 12% uncertainty represents the  $2\sigma$  error in the fluorescence signal due to the uncertainty in the online wavelength position, while the 6% uncertainty is the  $2\sigma$  error of the laser power measured with the power meter. A minor change was made in the last sentence of section 3.2.1:

“...of 12 % in the online FAGE signal and 6 % uncertainty in the laser power measured by the laser power meter and used to normalize the data. The uncertainty associated with the online signal, 12 % at  $2\sigma$  level, was calculated as the average deviation of the signal value due to the error limits of  $\pm 5 \times 10^{-4}$  nm in the online wavelength position (see the typical laser excitation scans shown in Fig. 3).”

**P16 L39:** The authors indicate that the oxygen concentration was lowered in some experiments performed on the HIRAC chamber. Could the lower oxygen concentration lead to a different sensitivity towards  $CH_3O$  due to changes in quenching rates of excited  $CH_3O$ ?

Line 39 of page 16 describes the methoxy radical measurement in HIRAC (section 3.3) which is shown in Figure 8. The concentration of  $CH_3O$  in these HIRAC experiments was obtained by using the calibration factor for methoxy radicals, which in turn was determined using the photolysis of methanol in  $N_2$  method described in section 2.3.1. In the HIRAC experiment shown in Figure 8  $O_2$  was only present in trace amounts ( $[O_2]_{HIRAC} = 5.4 \times 10^{15}$  molecule  $cm^{-3}$ , which corresponds to  $1.8 \times 10^{13}$  molecule  $cm^{-3}$   $O_2$  in the fluorescence detection cell) as described in section 3.3. This  $[O_2]$  is too small to produce a faster quenching rate of the  $CH_3O$  LIF signal in the chamber experiment compared to the quenching rate when using pure  $N_2$ , as estimated using the quenching rate coefficient of  $O_2$  reported by Wantuck et al. (1986),  $2.5 \times 10^{-11}$   $cm^3$  molecule $^{-1}$  s $^{-1}$ . The following sentence was added in the first paragraph of section 3.3 for clarification:

“The concentration of CH<sub>3</sub>O during the experiment was computed by using the FAGE calibration factor for methoxy radicals generated from the photolysis of methanol in N<sub>2</sub>,  $C_{\text{CH}_3\text{O}} = (5.1 \pm 2.2) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> (Sect. 3.1.1). The temporal profile of the CH<sub>3</sub>O is shown in Fig. 8...”

### **Author response to anonymous referee #3 on “A new method for atmospheric detection of the CH<sub>3</sub>O<sub>2</sub> radical” by L. Onel et al.**

Note: The changes in the manuscript addressing the comments of the referee #3 are highlighted in yellow below. The authors refer to the line numbers in the manuscript before revision mentioned in the comments.

The authors would like to thank anonymous referee #3 for their valuable comments to this manuscript.

*Page 4 line 30 and Page 6 Figures 2 and 3: As with detection of OH by the LIF FAGE technique, the authors must tune the laser on and off of the CH<sub>3</sub>O transition to determine the net signal due to CH<sub>3</sub>O fluorescence and the background signal due to laser scatter and other broadband fluorescence. OH LIF-FAGE instruments use a reference cell that generates high concentration of OH radicals to ensure that the laser is tuned to the correct frequency. It is unclear how the authors know that the laser is tuned to the correct CH<sub>3</sub>O excitation wavelength. Do they use a spectrometer to measure the wavelength, or do they have a reference cell that generates CH<sub>3</sub>O radicals?*

The signals were large enough that during conditions where CH<sub>3</sub>O<sub>2</sub> concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser-wavelength was stable over a long period once the laser wavelength had been tuned to the CH<sub>3</sub>O transition. Hence, the online wavelength position for CH<sub>3</sub>O fluorescence detection was found without using a reference cell. The laser excitation scans shown in Figures 2 and 3 were performed using the flow tube method described in the sections 2.3.1 and 2.3.2.1 to generate either CH<sub>3</sub>O (by the CH<sub>3</sub>OH photolysis at 185 nm) or CH<sub>3</sub>O<sub>2</sub> (by the H<sub>2</sub>O photolysis at 185 nm to generate OH followed by the reaction of the produced OH with CH<sub>4</sub> in the presence of O<sub>2</sub>).

In the HIRAC experiments the concentration of CH<sub>3</sub>O<sub>2</sub> radicals generated in the chamber in a steady-state with the UV lamps turned on at the beginning of each experiment using the Cl<sub>2</sub>/CH<sub>4</sub>/air system was used to

tune the laser at the correct excitation wavelength by performing similar scans to the laser scans shown in Figure 3.

In all measurements the offline wavelength position was fixed to the value obtained by adding 2.5 nm to  $\lambda(\text{online})$  as described at page 5, line 14. For field measurements in the future, when the concentrations of  $\text{CH}_3\text{O}_2$  (and hence  $\text{CH}_3\text{O}$  after conversion) will be both lower and more variable over short timescales, a reference cell will be necessary. We are in the process of developing a reference cell.

The third paragraph on page 5 (lines 10-24) was changed for clarification:

“The signals were large enough that during conditions where  $\text{CH}_3\text{O}_2$  concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser-wavelength was stable over a long period once the laser wavelength had been tuned to the  $\text{CH}_3\text{O}$  transition. Hence, the online wavelength position for  $\text{CH}_3\text{O}$  fluorescence detection was found without using a reference cell. Figure 2 shows the laser excitation spectrum centred at  $\sim 298$  nm in the  $\nu_3$  vibronic band recorded using an increment of  $\Delta\lambda = 10^{-3}$  nm. The spectrum agrees well with previous work (Inoue et al., 1980; Kappert and Temps, 1989; Shannon et al., 2013). Figure 3 shows typical laser excitation scans performed over a narrower range of wavelengths in order to locate  $\lambda(\text{online})$ . The LIF spectra were obtained by using the  $\text{CH}_3\text{O}$  or  $\text{CH}_3\text{O}_2$  radicals generated in a flow tube described in Sect. 2.3.1, with the flow tube output impinging close to the FAGE sampling inlet. The radicals were generated using the 184.9 nm light output of a Hg Pen-Ray lamp by either the photolysis of methanol in nitrogen to generate  $\text{CH}_3\text{O}$  or the photolysis of water vapour in synthetic air (to generate OH) in the presence of methane to form  $\text{CH}_3\text{O}_2$ . The  $\text{CH}_3\text{O}$  radicals were directly detected, while the  $\text{CH}_3\text{O}_2$  radicals were first converted to  $\text{CH}_3\text{O}$  species by added NO prior to the fluorescence detection cell (Fig. 1). Similar laser scans to the scans shown in Fig. 3 were recorded by using the  $\text{CH}_3\text{O}_2$  radicals produced in a steady-state concentration in HIRAC using photolytic mixtures of  $\text{Cl}_2/\text{CH}_4/\text{air}$  as described in Sect. 2.3.2.2. There were no unexpected features in the laser scans recorded when FAGE sampled  $\text{CH}_3\text{O}_2$  radicals from HIRAC, consistent with no interference being anticipated in the FAGE measurements of  $\text{CH}_3\text{O}$  as there were no other species in HIRAC absorbing at 298 nm and fluorescing at the wavelengths transmitted by the bandpass filter (average transmission  $> 80\%$  over 320 – 430 nm).

In this work the FAGE signals were large enough that during conditions where  $\text{CH}_3\text{O}_2$  concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser wavelength was stable over a long period once  $\lambda$  had been tuned to the  $\text{CH}_3\text{O}$  transition. Hence,  $\lambda(\text{online})$  was found without using a reference cell. We are in the process of developing a reference cell for field measurements in the future, when the concentrations of  $\text{CH}_3\text{O}_2$  (and hence  $\text{CH}_3\text{O}$  after conversion) will be both lower and more variable over short timescales.”

**Page 8 line 25:** Equation 2 assumes that the concentration of methanol is proportional to the concentration of water vapor and that any loss of methanol in their bubbler system is equal to any loss of water in their flow tube. Can the authors justify this assumption?

Equation 2 assumes that the concentration of methanol vapour in the photolysis flow tube is equal to the concentration of water vapour in the flow tube obtained when the bubbler contained water instead of methanol. The flow tube calibration using the water vapour photolysis represents the conventional FAGE calibration method for OH and HO<sub>2</sub> and previous investigations have shown that the water vapour loss in the system formed by the bubbler and the flow tube is negligible. Even less wall losses can be expected in the case of methanol, which has a significantly higher vapour pressure than water.

The following sentence was added after equation 2 (page 8, line 29):

“Equation 2 assumes that there were no losses of water vapour and methanol vapour by condensation in the tubing connecting the bubbler to the flow tube. This is as expected based on the small difference in temperature between the bubbler (*vide supra*) and the connecting tubing (typically held at ~ 20 °C) and as the gas going through the bubbler was diluted with the gas by-passing the bubbler.”

**Page 12, line 25:** (i) The authors claim that reducing the pressure in their FAGE detection cell could increase the sensitivity of the instrument. Is this due to reduced quenching of the CH<sub>3</sub>O fluorescence by air? (ii) Have the authors measured the impact trace gases on the fluorescence efficiency, such as water vapor?

(i) A potential improvement of the instrument sensitivity for CH<sub>3</sub>O<sub>2</sub> by using a pressure in the detection cell lower than the present limit of 2.65 Torr is expected because of the experimental observation of an increase in the fluorescence signal when the pressure in the detection cell is reduced from 10.00–2.65 Torr. As the pressure is reduced there is a reduction in the CH<sub>3</sub>O number density (which would decrease the LIF signal) and also a decrease in the quenching rate of the CH<sub>3</sub>O fluorescence by air, and hence an increase in the fluorescence quantum yield (which would increase the LIF signal). These two effects are opposing, but at low pressures do not cancel, leading to the observed increase in signal with lower pressures. It is therefore expected that as the pressure is reduced further below 2.65 Torr that the signal would continue to increase. Another reason could be that the characteristics of the jet expansion and/or the ensuing flow to the LIF detection region change with pressure, leading to a more favourable transmission of radicals to the detection region, but it is difficult to test this experimentally. For clarification the text (page 12, lines 24 – 27) was modified as follows:

“The present investigations into the change of sensitivity with pressure in the range from 2.65–10.00 Torr found that 2.65 Torr is the optimum value in this pressure interval. The result suggests that, by reducing the pressure in the above range of values, the decrease in fluorescence due to the reduction in the CH<sub>3</sub>O number density was overcome by the increase in the fluorescence quantum yield due to a lower fluorescence quenching rate. Another reason could be that the characteristics of the jet expansion and/or the ensuing flow to the LIF detection region change with pressure, leading to a more favourable transmission of radicals to the detection region, but it is difficult to test this experimentally. Hence an additional improvement in the sensitivity might be obtained by using a lower detection cell pressure than the current value of 2.65 Torr using a more powerful pump.”

(ii) No measurement of the rate coefficients of the fluorescence quenching by the traces gases has been performed in this work. However, a very good agreement was obtained between the flow tube calibrations for CH<sub>3</sub>O<sub>2</sub> with two different concentrations of water vapour in the flow tube:  $7.5 \times 10^{16}$  molecule cm<sup>-3</sup> or  $3 \times 10^{17}$  molecule cm<sup>-3</sup> (corresponding to  $2.6 \times 10^{14}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{15}$  molecule cm<sup>-3</sup>, respectively in the FAGE detection cell) as shown by Figure 6 in Sect. 2.3.2.1. The result presented in Figure 6 shows that the CH<sub>3</sub>O fluorescence quenching rate by water is minor for the above [H<sub>2</sub>O].

Methane was also present in the FAGE chamber in concentrations of several times  $10^{14}$  molecule cm<sup>-3</sup>. Calculations using the CH<sub>3</sub>O fluorescence quenching rate coefficient of CH<sub>4</sub> reported by Wantuck et al. (1987),  $1.05 \times 10^{-10}$  s<sup>-1</sup>, and a pressure in the FAGE detection cell of 2.65 Torr show only minor decreases in the fluorescence quantum yield, by few percent, when [CH<sub>4</sub>] is increased from zero to the experimental values. Assuming a quenching rate coefficient of H<sub>2</sub>O equal to that of CH<sub>4</sub>, similar small decreases in the fluorescence quantum yield were computed when [H<sub>2</sub>O] was increased from zero to the concentration values used in the flow tube calibration ( $0.3 - 1.0 \times 10^{15}$  molecule cm<sup>-3</sup>).

A paragraph which discusses the CH<sub>3</sub>O(A) quenching rates of water and methane at the concentrations used in the flow tube calibration of the FAGE instrument for CH<sub>3</sub>O<sub>2</sub> has been added at the end of the section 3.1.1:

“The calibrations using the flow-tube (“wand”) method have been performed under water vapour concentrations similar to the ambient [H<sub>2</sub>O<sub>vapour</sub>] but few orders of magnitude higher than those present in the HIRAC chamber experiments. In contrast with [H<sub>2</sub>O<sub>vapour</sub>] the methane concentrations used in the “wand” method were similar to [CH<sub>4</sub>] present in HIRAC but higher than [CH<sub>4</sub>] in the atmosphere. However, as detailed in this paragraph, the effects of methane and water on our sensitivity are minimal. Estimations using the reported fluorescence quenching rate coefficient of CH<sub>3</sub>O(A) by CH<sub>4</sub>,  $k_{\text{quench.CH}_4} = 1.05 \times 10^{-10}$  s<sup>-1</sup>, (Wantuck et al., 1987) and the concentrations of CH<sub>4</sub> in the LIF detection cell for the calibrations using the flow-tube ( $1.7 \times 10^{14}$  molecule cm<sup>-3</sup> and  $3.4 \times 10^{14}$  molecule cm<sup>-3</sup>, corresponding to  $5.0 \times 10^{16}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{17}$  molecule cm<sup>-3</sup>, respectively in the flow tube) resulted in only ~ 1–2% lower fluorescence

quantum yield compared to the value determined in the absence of CH<sub>4</sub>. No literature value has been found for the fluorescence rate coefficient of CH<sub>3</sub>O(A) fluorescence by H<sub>2</sub>O vapour. However, even if it assumed to be as large as the above reported value for CH<sub>4</sub> ( $k_{\text{quench,CH}_4}$ ), only a few percent decrease in the fluorescence quantum yield is computed (compared with a water concentration of zero) for the levels of H<sub>2</sub>O vapour which are present at the CH<sub>3</sub>O<sub>2</sub> FAGE detection axis when using the flow tube calibration method. These levels (1–2 % v/v) are similar to a typical water vapour concentration in the atmosphere. A very good agreement has been obtained between the calibration factors for CH<sub>3</sub>O<sub>2</sub> detection with two different concentrations of water vapour in the flow tube:  $7.5 \times 10^{16}$  molecule cm<sup>-3</sup> or  $3.0 \times 10^{17}$  molecule cm<sup>-3</sup> (corresponding to  $2.6 \times 10^{14}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{15}$  molecule cm<sup>-3</sup>, respectively in the FAGE cell) as shown in Figure 6 in Sect. 2.3.2.1. This very good agreement for H<sub>2</sub>O vapour and the above calculations for CH<sub>4</sub> support the use of the flow tube method for the FAGE calibration of the CH<sub>3</sub>O<sub>2</sub> concentrations.”

*Page 12, line 26: How does the OH sensitivity of the HIRAC FAGE compare to the field instrument? Assuming the CH<sub>3</sub>O sensitivity scales with the differences in the OH sensitivity, can the authors be more specific regarding the potential improvement in the LOD if this technique were to be used in the field instrument?*

The HIRAC FAGE sensitivity for OH is about two times lower than the ground-based field instrument sensitivity for OH:  $C_{\text{OH (HIRAC)}} = 8 \times 10^{-8}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>,

$C_{\text{OH (field)}} = 1.5 \times 10^{-7}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>.

As the distance from the inlet pinhole to the laser axis in the CH<sub>3</sub>O<sub>2</sub> fluorescence cell (Figure 1, 580 mm) is considerably longer than the corresponding distance in the ground-based field fluorescence cell for OH and HO<sub>2</sub> detection (88 mm), improvements in the CH<sub>3</sub>O<sub>2</sub> sensitivity are expected for the field FAGE instrument. The decrease in the pinhole-to-laser axis from 580 mm to 88 mm would result in a reduced loss of the CH<sub>3</sub>O<sub>2</sub> radicals on the instrument internal walls and would provide a greater population in the laser probed rotational level as the gas is still cooler than ambient following the pinhole expansion. However, the increase in the CH<sub>3</sub>O sensitivity cannot be quantified simply using the difference in the OH sensitivity between the HIRAC instrument and the field instrument. A larger increase in sensitivity between the field instrument and HIRAC would be expected for OH than for CH<sub>3</sub>O<sub>2</sub> based on expected heterogeneous losses, as the wall loss of OH is larger than the wall loss of CH<sub>3</sub>O<sub>2</sub>. However, how much the decrease in temperature at the laser axis owing to a smaller nozzle-to-laser axis distance improves the FAGE sensitivity for CH<sub>3</sub>O compared to the sensitivity for OH needs further investigation.

We think that no modification of the text is necessary as it cannot be assumed that improvements in the CH<sub>3</sub>O<sub>2</sub> sensitivity will scale with the difference in the OH sensitivity between the HIRAC instrument and the ground-field instrument.

**Page 16, line 19:** *The authors suggest that based on their flow tube calibrations that the rate constant for the CH<sub>3</sub>O<sub>2</sub> + CH<sub>3</sub>O<sub>2</sub> reaction may be 25% too high, perhaps due to a 25% overestimation of the CH<sub>3</sub>O<sub>2</sub> absorption cross section. What is the uncertainty associated with the recommended rate constant? Does the rate constant derived using their flow tube calibration factor agree to within the combined uncertainty of the calibration and the rate constant?*

The associated uncertainty with the IUPAC recommended value of the rate coefficient for the CH<sub>3</sub>O<sub>2</sub> self-reaction,  $k_{\text{CH}_3\text{O}_2}$ , is ~ 12% (1 $\sigma$ ). Our measured value, based on the flow tube calibration factor is ~25% lower than the IUPAC recommendation, with an overall error of ~20% (1 $\sigma$ ). Therefore, the obtained  $k_{\text{CH}_3\text{O}_2}$  have overlapping error limits with the IUPAC preferred value at the 1 $\sigma$  level.

The overall uncertainties of the two calibration methods of FAGE are discussed in detail in the manuscript. Even though the kinetic method agrees well with the flow tube method, it should be noted that the use of a lower value of  $k$  than  $k_{\text{CH}_3\text{O}_2}$ (IUPAC) would improve the level of agreement. Therefore, the text has not been changed.

**Page 17, Figure 8:** *The authors measure the concentration of CH<sub>3</sub>O in nitrogen to reduce the loss of CH<sub>3</sub>O from the CH<sub>3</sub>O<sub>2</sub> + O<sub>2</sub> reaction. However, it appears that they use the calibration factor determined in air to estimate the CH<sub>3</sub>O concentrations in this experiment. Does the calibration factor change in N<sub>2</sub> compared to air due to different fluorescence quenching rates?*

The concentration of CH<sub>3</sub>O in the HIRAC experiment shown in Figure 8 was obtained by using the calibration factor for methoxy radicals, which in turn was determined using the photolysis of methanol in N<sub>2</sub> method described in section 2.3.1. In this HIRAC experiment O<sub>2</sub> was only present in trace amounts ( $[\text{O}_2]_{\text{HIRAC}} = 5.4 \times 10^{15}$  molecule cm<sup>-3</sup>, which corresponds to  $1.8 \times 10^{13}$  molecule cm<sup>-3</sup> O<sub>2</sub> in the fluorescence detection cell) as described in section 3.3. This [O<sub>2</sub>] is too small to produce a faster quenching rate of the CH<sub>3</sub>O LIF signal in the chamber experiment compared to the quenching rate when using pure N<sub>2</sub>, as estimated using the quenching rate coefficient of O<sub>2</sub> reported by Wantuck et al. (1986),  $2.5 \times 10^{-11}$  cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup>.

The following sentence was added in the first paragraph of section 3.3 for clarification:

“The concentration of CH<sub>3</sub>O during the experiment was computed by using the FAGE calibration factor for methoxy radicals generated from the photolysis of methanol in N<sub>2</sub>,  $C_{\text{CH}_3\text{O}} = (5.1 \pm 2.2) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> (Sect. 3.1.1). The temporal profile of the CH<sub>3</sub>O is shown in Fig. 8...”

# A new method for atmospheric detection of the CH<sub>3</sub>O<sub>2</sub> radical

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**Abstract.** A new method for measurement of the methyl peroxy (CH<sub>3</sub>O<sub>2</sub>) radical has been developed using the conversion of CH<sub>3</sub>O<sub>2</sub> into CH<sub>3</sub>O by excess NO with subsequent detection of CH<sub>3</sub>O by fluorescence assay by gas expansion (FAGE) with laser excitation at *ca.* 298 nm. The method can also directly detect CH<sub>3</sub>O, when no nitric oxide is added. Laboratory calibrations were performed to characterise the FAGE instrument sensitivity using the conventional radical source employed in OH calibration with conversion of a known concentration of OH into CH<sub>3</sub>O<sub>2</sub> via reaction with CH<sub>4</sub>/O<sub>2</sub>. Detection limits of  $3.8 \times 10^8$  molecule cm<sup>-3</sup> and  $3.0 \times 10^8$  molecule cm<sup>-3</sup> were determined for CH<sub>3</sub>O<sub>2</sub> and CH<sub>3</sub>O, respectively for a signal-to-noise ratio of 2 and 5 min averaging time. Averaging over 1 hour reduces the detection limit for CH<sub>3</sub>O<sub>2</sub> to  $1.1 \times 10^8$  molecule cm<sup>-3</sup> comparable to atmospheric concentrations. The kinetics of the second-order decay of CH<sub>3</sub>O<sub>2</sub> via its self-reaction were observed in HIRAC (Highly Instrumented Reactor for Atmospheric Chemistry) at 295 K and 1 bar and used as an alternative method of calibration to obtain a calibration constant with overlapping error limits at the 1σ level with the result of the conventional method of calibration. The overall uncertainties of the two methods of calibrations are similar: 15 % for the kinetic method and 17 % for the conventional method and are discussed in detail. The capability to quantitatively measure CH<sub>3</sub>O in chamber experiments is demonstrated via observation in HIRAC of CH<sub>3</sub>O formed as a product of the CH<sub>3</sub>O<sub>2</sub> self-reaction.

## 1 Introduction

Methyl peroxy (CH<sub>3</sub>O<sub>2</sub>) radicals are critical intermediates in the atmospheric oxidation (Orlando and Tyndall, 2012) and combustion of hydrocarbons (Zador et al., 2011). In the remote atmosphere CH<sub>3</sub>O<sub>2</sub> is mainly formed by the reaction of methane with the OH radical via abstraction of an H atom (R1), followed by the reaction of the produced CH<sub>3</sub> radical with O<sub>2</sub> (R2).



Methyl radicals can also be formed from more complex species, e.g. the reaction of acetyl peroxy radicals with HO<sub>2</sub> in low NO<sub>x</sub> environments or the reaction of acetyl peroxy radicals with NO in anthropogenically influenced environments. CH<sub>3</sub>O<sub>2</sub> is predicted to be the most abundant peroxy radical in the atmosphere, yet there are no specific measurements of its concentration. Daytime concentrations estimated using a box model utilizing the MCM (Master Chemical Mechanism) version 3.3.1 (Saunders et al., 2003; Jenkin et al., 2015) are  $\sim 6 \times 10^8$  molecule cm<sup>-3</sup> in the tropical Atlantic ocean in summer (Whalley et al., 2010),  $\sim 2 \times 10^8$  molecule cm<sup>-3</sup> in a tropical rainforest (Whalley et al., 2011), and lower in polluted environments, for example  $\sim 5 \times 10^7$  molecule cm<sup>-3</sup> in London in summertime (Whalley et al., to be submitted).

The reaction of CH<sub>3</sub>O<sub>2</sub> with NO (R3) usually dominates the chemistry of CH<sub>3</sub>O<sub>2</sub>, particularly in environments influenced by anthropogenic NO<sub>x</sub> emissions, resulting in NO<sub>2</sub> production and hence ozone production:



The subsequent reaction of CH<sub>3</sub>O with O<sub>2</sub> (R4) produces HO<sub>2</sub>, which in turn oxidises another NO to NO<sub>2</sub> (R5) with further production of O<sub>3</sub> and propagation of the HO<sub>x</sub> radical chain:



However, under low NO<sub>x</sub> levels (e.g. remote forested environments and the marine boundary layer) the self-reaction of CH<sub>3</sub>O<sub>2</sub> (R6) and the reactions of CH<sub>3</sub>O<sub>2</sub> with HO<sub>2</sub> and other organic peroxy (RO<sub>2</sub>) species are important radical removal/termination reactions. The CH<sub>3</sub>O<sub>2</sub> self-reaction occurs through two channels, (R6.a) and (R6.b) (Tyndall et al., 1998):



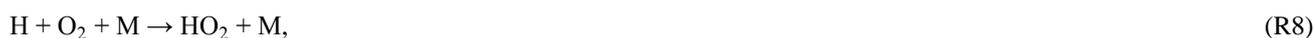
Despite the importance of the reaction (R6), there are uncertainties of about a factor of two in the value of its rate coefficient at room temperature,  $k_6$ , which ranges from  $(2.7\text{--}5.2) \times 10^{-13} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  (Atkinson et al., 2006); the preferred IUPAC value is  $k_6 = 3.5 \times 10^{-13} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  (Atkinson et al., 2006). The previous kinetic studies used time-resolved UV-absorption spectroscopy to detect CH<sub>3</sub>O<sub>2</sub> radical, typically at 250 nm, (Sander and Watson, 1980, 1981;McAdam et al., 1987;Kurylo and Wallington, 1987;Jenkin et al., 1988;Simon et al., 1990;Lightfoot et al., 1990). UV-absorption spectroscopy is a relatively insensitive technique and hence the detection limits of CH<sub>3</sub>O<sub>2</sub> were quite high, for example approximately  $4 \times 10^{12} \text{ molecule cm}^{-3}$  (Sander and Watson, 1980, 1981). In addition, due to the broad, featureless spectra of RO<sub>2</sub> species, which often overlap, UV-absorption is a relatively unselective technique for the study of the kinetics of individual RO<sub>2</sub>. Therefore, there is a clear need for the determination of  $k_6$  using a more selective method, which will be addressed in subsequent studies.

At present, CH<sub>3</sub>O<sub>2</sub> is not specifically measured in the atmosphere by any direct or indirect method. Time-resolved continuous-wave cavity ringdown spectroscopy (CRDS), using the  $\nu_{12}$  transition of the  $A \leftarrow X$  band at  $\sim 1.3 \mu\text{m}$  has been used to detect CH<sub>3</sub>O<sub>2</sub> directly in a photoreactor (Farago et al., 2013;Bossolasco et al., 2014). However, the detection limit is not sufficiently sensitive to enable tropospheric detection. Typically, the sum of HO<sub>2</sub> and all organic RO<sub>2</sub> has been measured in the atmosphere, making no distinction between HO<sub>2</sub> and different RO<sub>2</sub> species, although more recently the sum of RO<sub>2</sub> has been quantified separately to HO<sub>2</sub>. One of the methods uses Chemical Ionisation Mass Spectrometry to determine the sum  $[\text{HO}_2] + \sum_i [\text{RO}_{2,i}]$  or separately  $[\text{HO}_2]$ , depending on the control of the flows of the NO and SO<sub>2</sub> reagents (Hanke et al., 2002;Edwards et al., 2003). The sum  $[\text{HO}_2] + \sum_i [\text{RO}_{2,i}]$  has also been determined for many years by the Peroxy Radical Chemical Amplifier (PERCA) method, which uses NO and CO to generate NO<sub>2</sub> amplified by a chain reaction, and subsequently measured by a variety of methods, for example luminol fluorescence, laser-induced fluorescence (LIF) or cavity absorption methods (Cantrell and Stedman, 1982;Cantrell et al., 1984;Miyazaki et al., 2010;Hernandez et al., 2001;Green et al., 2006;Chen et al., 2016). A modification of PERCA, using a denuder to remove HO<sub>2</sub> has been used to estimate the sum of RO<sub>2</sub> (Miyazaki et al., 2010). RO<sub>x</sub>LIF is a more recent method, which uses OH LIF detection at low pressure, known as FAGE (fluorescence assay by gas expansion) (Fuchs et al., 2008;Whalley et al., 2013). The RO<sub>x</sub>LIF method measures either  $[\text{HO}_x] = [\text{OH}] + [\text{HO}_2]$  by converting HO<sub>x</sub> into HO<sub>2</sub> through addition of CO, or  $[\text{RO}_x] = [\text{HO}_x] + \sum_i ([\text{RO}_{2,i}] + [\text{RO}_1])$  by titrating RO<sub>x</sub> to HO<sub>2</sub> by added NO and CO. After the conversion into HO<sub>2</sub>, HO<sub>2</sub> is converted into OH in the FAGE chamber and detected by LIF. The sum  $\sum_i [\text{RO}_{2,i}]$  and the concentration of the initial HO<sub>2</sub> can be determined from the separate measurements of HO<sub>x</sub>, RO<sub>x</sub> and OH. The limit of detection of the RO<sub>x</sub>LIF method is  $\sim 0.1 \text{ pptv}$  ( $2.5 \times 10^6 \text{ molecule cm}^{-3}$ ) (Fuchs et al., 2008;Whalley et al., 2013). Recently, the interference from certain types of RO<sub>2</sub> radicals in the FAGE detection of HO<sub>2</sub> was deliberately exploited to enable a partial RO<sub>2</sub> speciation (Whalley et al., 2013). The method was used in the Clean Air for

London campaign (ClearfLo) to distinguish between the sum of alkene, aromatic and long-chain alkane-derived RO<sub>2</sub> radicals and the sum of short-chain alkane-derived RO<sub>2</sub> radicals (Whalley et al., 2013).

As methoxy (CH<sub>3</sub>O) radicals can be generated by techniques such as pulsed laser photolysis and microwave discharge and detected with high sensitivity by LIF (Shannon et al., 2013;Chai et al., 2014;Albaladejo et al., 2002;Biggs et al., 1993;Biggs et al., 1997), the method has been used in kinetic studies of a range of CH<sub>3</sub>O reactions. These studies used the electronic excitation of the methoxy radical from the ground state to the first electronically excited state ( $A^2A_1 \leftarrow X^2E$ ). The  $A \leftarrow X$  excitation spectrum covers the range ~ 275–317 nm and leads to fluorescence from several vibronic bands in the near UV, and has been reported in a series of experimental and theoretical studies (Inoue et al., 1980;Kappert and Temps, 1989;Powers et al., 1997;Nagesh et al., 2014).

This paper reports the development of a new method for the selective and sensitive detection of CH<sub>3</sub>O<sub>2</sub> radicals using FAGE by titrating CH<sub>3</sub>O<sub>2</sub> to CH<sub>3</sub>O by reaction with added NO (R3) and then detecting the resultant CH<sub>3</sub>O by off-resonant LIF with laser excitation at *ca.* 298 nm. The method is similar to the standard method used for the detection of HO<sub>2</sub> radicals by FAGE through conversion of HO<sub>2</sub> to OH by reaction with added NO followed by OH on-resonance LIF at about 308 nm (Heard and Pilling, 2003). As LIF is not an absolute detection method, FAGE instruments require calibration, with the 184.9 nm photolysis of water vapour in air using a mercury (Hg) Pen-Ray lamp being a common method employed for generating known concentrations of OH and HO<sub>2</sub> (Heard and Pilling, 2003):



where M = N<sub>2</sub>, O<sub>2</sub> and the photodissociation quantum yield of OH and H is unity. In this study the photolysis of water vapour is performed in the presence of excess methane to produce CH<sub>3</sub>O<sub>2</sub>:



An alternative CH<sub>3</sub>O<sub>2</sub> calibration is also presented, consisting of the analysis of the kinetics of the CH<sub>3</sub>O<sub>2</sub> decay by self-reaction monitored by FAGE and compared with the water photolysis method. The studies are performed within HIRAC (Highly Instrumented Reactor for Atmospheric Chemistry) which is a 2.25 m<sup>3</sup>, custom-built, stainless steel chamber simulating the ambient conditions (Glowacki et al., 2007). HIRAC has been used in alternative calibrations of FAGE for OH and HO<sub>2</sub> using the temporal evolution of appropriate species, in validation and development of new atmospheric measurement techniques as well as in kinetic and mechanistic studies of atmospheric relevant reactions (Malkin et al., 2010;Winiberg et al., 2015;Winiberg et al., 2016).

Direct LIF detection of CH<sub>3</sub>O radicals, which is also a key intermediate in the oxidation of methane and other VOCs in the troposphere and formed by reactions such as (R3) and (R6.b), is also reported here. However, in the atmosphere CH<sub>3</sub>O is exclusively consumed by reaction with O<sub>2</sub> (R4) generating formaldehyde and recycling HO<sub>2</sub>, resulting in a very short lifetime and consequently very low concentration (~10<sup>2</sup>-10<sup>3</sup> molecule cm<sup>-3</sup>). For this reason no measurements in the atmosphere have previously been attempted. The photolysis of CH<sub>3</sub>OH at 184.9 nm is used to estimate the FAGE sensitivity for CH<sub>3</sub>O. The dominant photolysis channel of methanol between 165 and 200 nm generates CH<sub>3</sub>O radicals (Wen et al., 1994;Kassab et al., 1983;Marston et al., 1993):

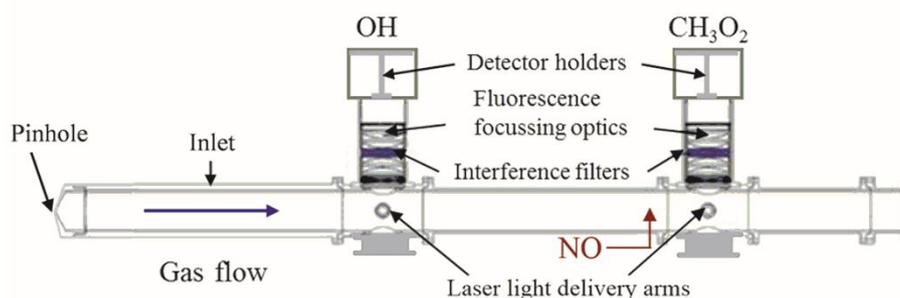


A photodissociation quantum yield of  $\text{CH}_3\text{O}$  of  $0.86 \pm 0.10$  has been found at 193.3 nm (Satyapal et al., 1989) in qualitative agreement with analysis of the end-products of the methanol photodissociation at 184.9 nm (Porter and Noyes, 1959; Bunker et al., 1984). Here we report **the first** measurements of  $\text{CH}_3\text{O}$  concentrations in an atmospheric simulation chamber. Methoxy radicals are generated by the  $\text{CH}_3\text{O}_2$  self-reaction carried out within HIRAC at 295 K and 1000 mbar of  $\text{N}_2$  containing  $\text{O}_2$  in trace amounts to reduce the rate of removal of  $\text{CH}_3\text{O}$  by reaction with  $\text{O}_2$ . This work enhances the capability of HIRAC to measure short-lived radical species by the addition of both  $\text{CH}_3\text{O}_2$  and  $\text{CH}_3\text{O}$  detection, and we discuss the potential of the method for detection of  $\text{CH}_3\text{O}_2$  in the atmosphere itself.

## 2 Experimental

### 2.1 The FAGE instrument

Details on the HIRAC-based FAGE instrument for the detection of OH and  $\text{HO}_2$  has been presented previously (Winiberg et al., 2015). Figure 1 shows a schematic cross-section of the instrument inlet and the two fluorescence detection cells. The gas was sampled **with a flow rate of 3.2 slm** through a 1 mm diameter pinhole and passed down a 50 mm diameter flow tube of 280 mm length first into the OH detection axis and, after a further 300 mm, into the  $\text{CH}_3\text{O}_2$  detection axis. The pressure in the detection cells was maintained at  $(2.65 \pm 0.05)$  Torr by using a high capacity rotary-backed roots blower pumping system (Leybold, trivac D40B and RuVac WAU251).  $\text{CH}_3\text{O}_2$  radicals were titrated to  $\text{CH}_3\text{O}$  by adding high purity NO (BOC, N2.5 Nitric Oxide) with a typical 2.5 sccm flow rate (further details in Section 2.2) ~25 mm before the second detection axis into the centre of the flow. The resultant  $\text{CH}_3\text{O}$  radicals were measured by LIF.



**Figure 1.** Vertical cross-section of the FAGE fluorescence cells. The first (left) fluorescence cell was used to detect OH fluorescence through a  $(308.8 \pm 5.0)$  nm bandpass filter (transmission  $> 50\%$ ) and the second cell to detect  $\text{CH}_3\text{O}_2$  after titration with added NO to form  $\text{CH}_3\text{O}$  using a bandpass filter between 320–430 nm with an average transmission  $> 80\%$ .

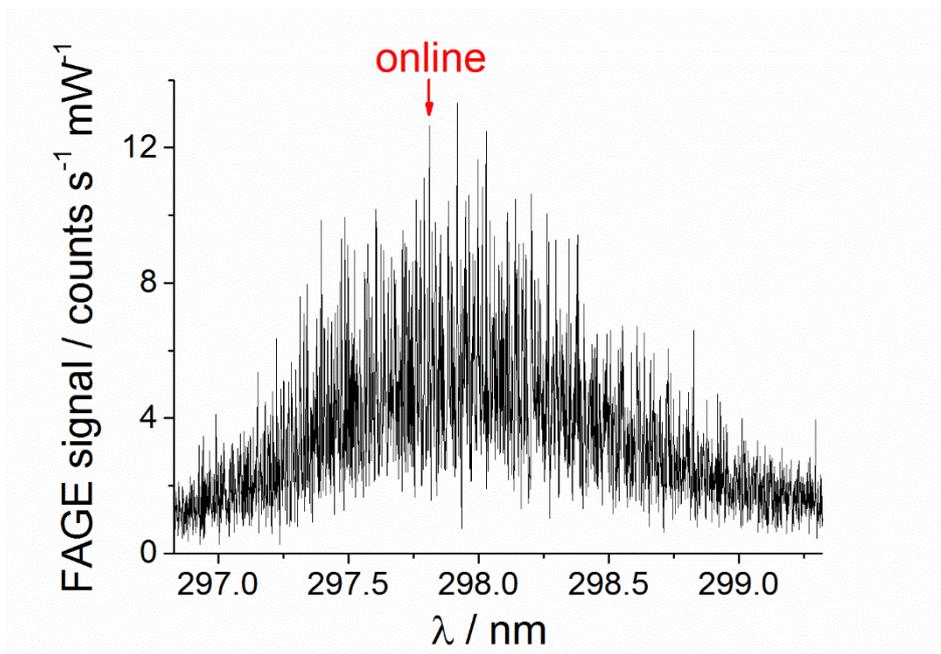
Probe laser light was generated by a Nd:YAG (JDSU Q201-HD) pumped dye laser (SIRAH Credo-Dye-N) using a DCM dye (Sirah) in ethanol and operating at 5 kHz pulse repetition frequency, with a pulse width at half maximum of 25 ns, typical pulse energy of  $120 \mu\text{J pulse}^{-1}$  and a linewidth of  $0.08 \text{ cm}^{-1}$  at 595 nm. The frequency doubled light at either ~308 nm (OH detection) or ~298 nm ( $\text{CH}_3\text{O}$  detection), was focused into fibre optic cables to be delivered to the two detection cells. OH and  $\text{CH}_3\text{O}$  radicals were separately detected by LIF spectroscopy by exciting at 307.99 nm using the  $Q_1(2)$  rotational line of the  $A^2\Sigma^+ (\nu' = 0) \leftarrow X^2\Pi_i (\nu'' = 0)$  OH transition in the first detection axis to monitor on-resonant fluorescence ( $308.8 \pm 5.0$  nm) and excitation at 297.79 nm in the  $A^2A_1 (\nu'_3 = 3) \leftarrow X^2E (\nu''_3 = 0)$   $\text{CH}_3\text{O}$  transition in the second detection axis to monitor red-shifted off-resonant LIF (320–430 nm). Here  $\nu_3$  refers to the C–O stretching vibrational mode of  $\text{CH}_3\text{O}$  which demonstrates a progression in the LIF spectrum (Inoue et al., 1980; Kappert and Temps, 1989; Powers et al., 1997; Nagesh et al., 2014). The fluorescence in the two cells was collected orthogonal to the gas flow by two microchannel plate photomultiplier tubes (MCP-PMT) (Photek PMT325/Q/BI/G)

equipped with a 50 ns gate unit (Photek GM10-50) for gated photon-counting, and the signal was amplified using a pre-amplifier (Photek PA200-10). Further details on the OH detection and calibration in HIRAC have been reported previously (Winiberg et al., 2015).

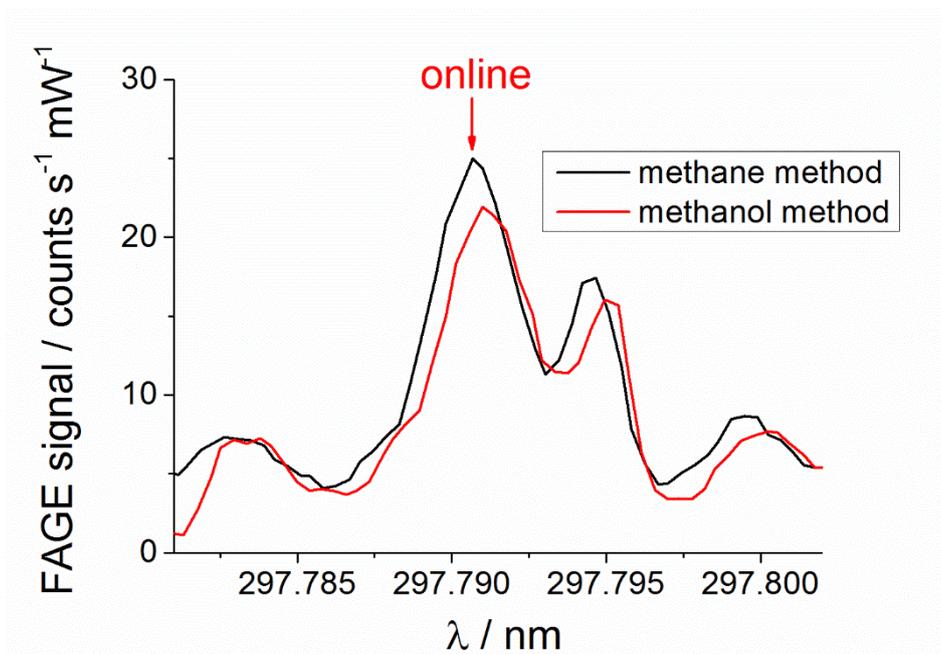
The laser and photon-counting timing for CH<sub>3</sub>O detection was controlled by a delay pulse generator (9520 Quantum Composers). The relatively broad bandpass filter used for the collection of the CH<sub>3</sub>O fluorescence (average transmission > 80% between 320–430 nm) allowed some red-shifted scattered light (presumably from the walls of the chamber) generated by the probe laser to be transmitted and hence detected by the MCP-PMT. In order to ameliorate this and reduce the background signal, the gate unit was opened 100 ns after the laser pulse to detect fluorescence integrated over a gate-width of 2 μs. The optimum gate-width of 2 μs (values in the range 1-3 μs were compared) is consistent with the CH<sub>3</sub>O fluorescence lifetimes, calculated to be in the range of 0.9 – 1.5 μs, using the reported radiative lifetimes for CH<sub>3</sub>O of 1.5 μs (Inoue et al., 1979), 2.2 μs (Ebata et al., 1982) and (4 ± 2) μs (Wendt and Hunziker, 1979) and using the fluorescence quenching rate coefficients of N<sub>2</sub> and O<sub>2</sub> (Wantuck et al., 1987) to calculate the rate of quenching at the pressure in the FAGE detection cell ((2.65 ± 0.05) Torr). As the fluorescence lifetime of CH<sub>3</sub>O(A) in the detection cell was 0.9–1.5 μs, delaying the counting of the fluorescence by 100 ns makes very little difference (~10%) in the fraction of fluorescence collected.

All LIF signals reported here were normalized to the probe laser power as measured with a laser power meter (Maestro, Gentec-EO) before the start of each LIF measurement. Fluctuations in the relative laser power were monitored via a photodiode (UDT-555UV, Laser Components) during the measurements and were accounted for in the signal normalization. The LIF spectrum was corrected for the laser-scattered background by subtracting the normalized offline signal recorded over 60 s at the end of each LIF measurement using an offline wavelength  $\lambda(\text{offline} = 300.29 \text{ nm}) = \lambda(\text{online} = 297.79 \text{ nm}) + 2.5 \text{ nm}$ , well away from any CH<sub>3</sub>O absorption. The signals were large enough that during conditions where CH<sub>3</sub>O<sub>2</sub> concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser-wavelength was stable over a long period once the laser wavelength had been tuned to the CH<sub>3</sub>O transition. Hence, the online wavelength position for CH<sub>3</sub>O fluorescence detection was found without using a reference cell. Figure 2 shows the laser excitation spectrum centred at ~298 nm in the  $\nu_3$  vibronic band recorded using an increment of  $\Delta\lambda = 10^{-3}$  nm. The spectrum agrees well with previous work (Inoue et al., 1980; Kappert and Temps, 1989; Shannon et al., 2013). Figure 3 shows typical laser excitation scans performed over a narrower range of wavelengths in order to locate  $\lambda(\text{online})$ . The LIF spectra were obtained by using the CH<sub>3</sub>O or CH<sub>3</sub>O<sub>2</sub> radicals generated in a flow tube described in Sect. 2.3.1, with the flow tube output impinged close to the FAGE sampling inlet. The radicals were generated using the 184.9 nm light output of a Hg Pen-Ray lamp by either the photolysis of methanol in nitrogen to generate CH<sub>3</sub>O or the photolysis of water vapour in synthetic air (to generate OH) in the presence of methane to form CH<sub>3</sub>O<sub>2</sub>. The CH<sub>3</sub>O radicals were directly detected, while the CH<sub>3</sub>O<sub>2</sub> radicals were first converted to CH<sub>3</sub>O species by added NO prior to the fluorescence detection cell (Fig. 1). Similar laser scans to the scans shown in Fig. 3 were recorded by using the CH<sub>3</sub>O<sub>2</sub> radicals produced in a steady-state concentration in HIRAC using photolytic mixtures of Cl<sub>2</sub>/CH<sub>4</sub>/air as described in Sect. 2.3.2.2. There were no unexpected features in the laser scans recorded when FAGE sampled CH<sub>3</sub>O<sub>2</sub> radicals from HIRAC, consistent with no interference being anticipated in the FAGE measurements of CH<sub>3</sub>O as there were no other species in HIRAC absorbing at 298 nm and fluorescing at the wavelengths transmitted by the bandpass filter (average transmission > 80 % over 320 – 430 nm).

In this work the FAGE signals were large enough that during conditions where CH<sub>3</sub>O<sub>2</sub> concentrations were constant (e.g. in calibrations or during HIRAC experiments where steady-state concentrations were generated) it was established that the laser wavelength was stable over a long period once  $\lambda$  had been tuned to the CH<sub>3</sub>O transition. Hence,  $\lambda(\text{online})$  was found without using a reference cell. We are in the process of developing a reference cell for field measurements in the future, when the concentrations of CH<sub>3</sub>O<sub>2</sub> (and hence CH<sub>3</sub>O after conversion) will be both lower and more variable over short timescales.



**Figure 2.** Laser excitation spectrum of the  $A^2A_1 (\nu'_3 = 3) \leftarrow X^2E (\nu''_3 = 0)$  transition of the methoxy radical.  $\text{CH}_3\text{O}$  radicals were obtained by photolysis of methanol in  $\text{N}_2$  at 184.9 nm. Fluorescence cell pressure =  $(2.65 \pm 0.05)$  Torr; wavelength increment  $\Delta\lambda = 10^{-3}$  nm, with each point corresponding to 5000 laser shots. The red arrow indicates the wavelength  $\lambda(\text{online}) \sim 297.79$  nm used for the time-resolved kinetic studies of  $\text{CH}_3\text{O}$ .



**Figure 3.** Typical laser excitation scans of  $\text{CH}_3\text{O}$  performed over a much smaller range of wavelengths. Methoxy radicals were generated using  $\text{OH}/\text{CH}_4$  (black line) to produce  $5.5 \times 10^{10}$  molecule  $\text{cm}^{-3}$   $\text{CH}_3\text{O}_2$ , subsequently titrated to  $\text{CH}_3\text{O}$  by adding  $\text{NO}$ , and the photolysis of methanol (red line) to generate  $4.9 \times 10^{10}$  molecule  $\text{cm}^{-3}$   $\text{CH}_3\text{O}$  directly. See main text for the description of the methods and calibration. The signal was normalised for the laser power ( $(10.3 \pm 0.3)$  mW in methane method and  $(8.7 \pm 0.2)$

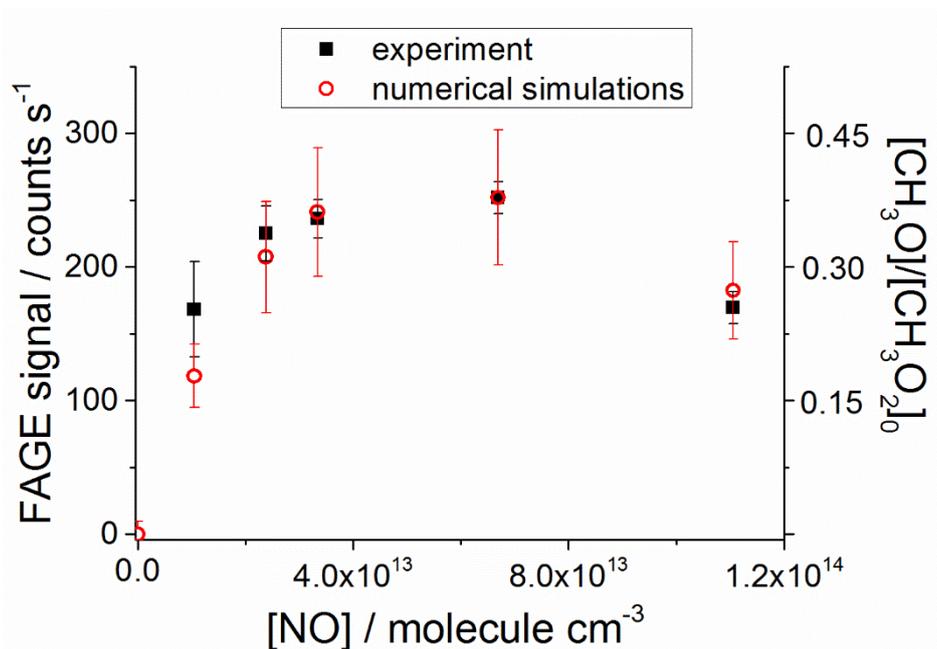
mW in methanol method). Fluorescence cell pressure =  $(2.65 \pm 0.05)$  Torr; wavelength increment  $\Delta\lambda = 10^{-3}$  nm, with each point corresponding to 5000 laser shots. The red arrow indicates the wavelength  $\lambda(\text{online}) \sim 297.79$  nm used for the time-resolved kinetic studies of  $\text{CH}_3\text{O}$ .

## 2.2 Optimisation of the NO concentration for methyl peroxy radical detection

As NO was added  $\sim 25$  mm prior the methoxy detection axis (Fig. 1), some of the methoxy radicals formed by Reaction (R3) reacted further with NO before the fluorescence detection:



where  $\text{M} = \text{N}_2, \text{O}_2$ . In addition to the above reactions,  $\text{CH}_3\text{O}$  reacts with  $\text{O}_2$  by Reaction (R4). Figure 4 shows the dependence of the LIF signal on the concentration of NO obtained experimentally and by numerical simulations using Reactions (R3)–(R4) and (R10)–(R11) and outlined in the Supplementary Information. A maximum signal was obtained with added  $[\text{NO}] = 6.7 \times 10^{13}$  molecule  $\text{cm}^{-3}$  for a reaction time of 3 ms, estimated from the linear flow velocity within the FAGE reactor. Figure 4 shows that the functional dependence with added  $[\text{NO}]$  of the experimental  $\text{CH}_3\text{O}$  signal and the simulated  $[\text{CH}_3\text{O}]/[\text{CH}_3\text{O}_2]_0$  ratio display the same shape (within overlapping error limits) with the numerical simulations showing that  $[\text{CH}_3\text{O}]/[\text{CH}_3\text{O}_2]_0$  at the detection axis was  $\sim 0.4$  (i.e. 40 % conversion to  $\text{CH}_3\text{O}$ ).



**Figure 4.** FAGE signal (left axis) and the ratio  $[\text{CH}_3\text{O}]/[\text{CH}_3\text{O}_2]_0$  (right axis) as a function of the concentration of NO for a reaction time of 3 ms. Black squares are experimental  $\text{CH}_3\text{O}$  signals (errors are  $1\sigma$ ) and red circles are the ratio  $[\text{CH}_3\text{O}]/[\text{CH}_3\text{O}_2]_0$  generated by numerical simulations (percentage uncertainties are 20 %) using the chemistry system outlined in the main text and described in further detail in the Supplementary Information.

## 2.3 FAGE calibrations

CH<sub>3</sub>O and CH<sub>3</sub>O<sub>2</sub> calibrations were carried out using the conventional radical source employed in fieldwork OH and HO<sub>2</sub> calibrations (Heard and Pilling, 2003) that produces radicals in a flow tube impinging just outside the FAGE inlet pinhole (Winiberg et al., 2015) and is described in Sect. 2.3.1. Two methods of calibration have been used for CH<sub>3</sub>O<sub>2</sub>: the flow tube method and the kinetics of the self-reaction of CH<sub>3</sub>O<sub>2</sub> carried out in HIRAC.

### 2.3.1 Calibration for methoxy radicals

In the CH<sub>3</sub>O calibration experiments nitrogen (BOC, > 99.998 %) was used as carrier gas. Part of the N<sub>2</sub> flow was passed through a methanol (Sigma Aldrich, ≥ 99.9 %) bubbler while the other portion bypassed the bubbler. The gas containing methanol vapour was then passed through a square cross-section flow tube of dimensions 13 × 13 (internal) × 300 mm length with a flow rate of 40 slm (ensuring turbulent flow conditions), controlled by an electronic flow controller (Brooks, 0–100 slm air). The collimated light of a Hg Pen-Ray lamp (LOT–Oriol Hg–Ar) was directed across the flow tube (close to the downstream end) to photolyse methanol vapour. The flow tube output was impinged close to the FAGE inlet to sample CH<sub>3</sub>O radicals at atmospheric pressure through a 1 mm diameter pinhole (Fig. 1).

The concentration of CH<sub>3</sub>O radicals was calculated using Eq. (1):

$$[\text{CH}_3\text{O}] = [\text{CH}_3\text{OH}] \sigma_{\text{CH}_3\text{OH}, 184.9 \text{ nm}} \Phi_{\text{CH}_3\text{O}, 184.9 \text{ nm}} F_{184.9 \text{ nm}} \Delta t, \quad (1)$$

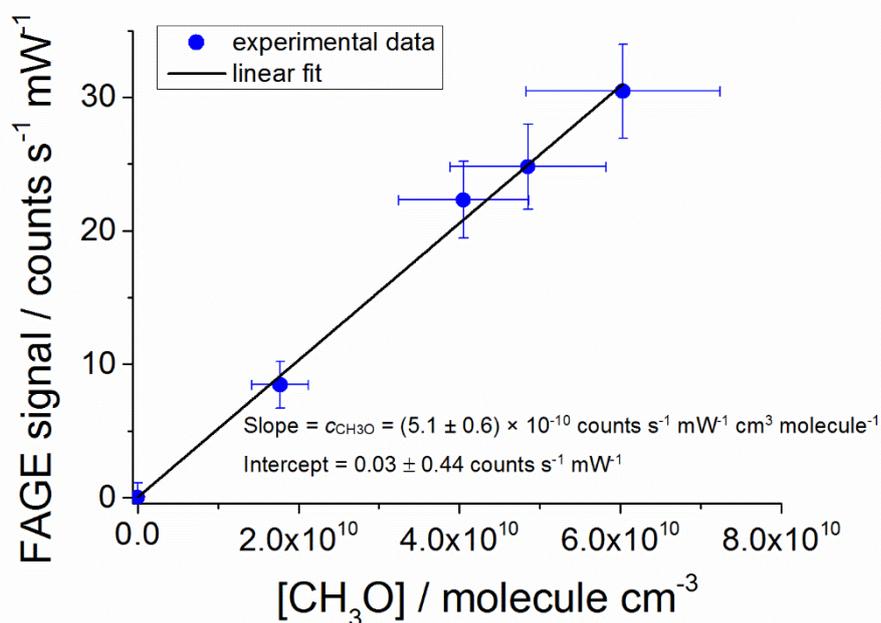
where  $\sigma_{\text{CH}_3\text{OH}, 184.9 \text{ nm}}$  is the absorption cross section of methanol at 184.9 nm,  $(6.35 \pm 0.28) \times 10^{-19} \text{ cm}^2 \text{ molecule}^{-1}$ , obtained by averaging reported values (Dillon et al., 2005; Jimenez et al., 2003; Nee et al., 1985),  $F_{184.9 \text{ nm}}$  is the photon flux of 184.9 nm light and  $\Delta t$  is the irradiation time of the gas. Although it is known, based on end-product analysis, that the scission of O–H bond is a major photolysis channel of methanol at 184.9 nm (Buenker et al., 1984; Porter and Noyes, 1959), the photodissociation quantum yield of CH<sub>3</sub>O at 184.9 nm,  $\Phi_{\text{CH}_3\text{O}, 184.9 \text{ nm}}$ , has not been yet reported. Here it is assumed that  $\Phi_{\text{CH}_3\text{O}, 184.9 \text{ nm}}$  is equal to the photodissociation quantum yield at 193.3 nm,  $\Phi_{\text{CH}_3\text{O}, 193.3 \text{ nm}} = 0.86 \pm 0.10$ , which has been reported (Satyapal et al., 1989). In order to determine the methanol vapour concentration in the flow tube, [CH<sub>3</sub>OH], separate experiments were carried out with the same calibration system to bubble deionised water instead of methanol with the same flow rate. The water vapour concentration, [H<sub>2</sub>O], was measured using a dew-point hygrometer (CR4, Buck Research Instrument) prior to the flow tube. Then [CH<sub>3</sub>OH] was calculated using the averaged [H<sub>2</sub>O] and the vapour pressures  $p_{\text{CH}_3\text{OH}}$  and  $p_{\text{H}_2\text{O}}$  at the temperatures measured for CH<sub>3</sub>OH (13 °C) and H<sub>2</sub>O (15 °C) in the bubbler:

$$[\text{CH}_3\text{OH}] = [\text{H}_2\text{O}] \frac{p_{\text{CH}_3\text{OH}}}{p_{\text{H}_2\text{O}}} \quad (2)$$

Equation 2 assumes that there were no losses of water vapour and methanol vapour by condensation in the tubing connecting the bubbler to the flow tube. This is as expected based on the small difference in temperature between the bubbler (*vide supra*) and the connecting tubing (typically held at ~ 20 °C) and as the gas going through the bubbler was diluted with the gas by-passing the bubbler.

N<sub>2</sub>O photolysis at 184.9 nm to generate NO (via reaction of the photoproduct (O<sup>1</sup>D) with N<sub>2</sub>O giving a known yield of NO), which was subsequently measured using a commercial analyser, was used as a chemical actinometer to obtain the product  $F_{184.9 \text{ nm}} \times \Delta t$  (Winiberg et al., 2015) and hence calculate [CH<sub>3</sub>O] via Eq. (1). The photolysis time,  $\Delta t$ , was estimated to be 8.3 ms, using the volumetric flow rate and the geometric parameters of the flow tube (assuming plug flow) and was in turn used to determine  $F_{184.9 \text{ nm}}$ . Although it is the product  $F_{184.9 \text{ nm}} \times \Delta t$  which is used to calculate [CH<sub>3</sub>O], any change in the volumetric flow rate between the

calibration and actinometry experiments will change  $\Delta t$ , and hence the product was corrected for any changes in volumetric flow rate. A range of  $[\text{CH}_3\text{O}]$  at constant  $[\text{CH}_3\text{OH}]$  was produced by changing the electrical current through the Hg lamp between 0 and 20 mA, and hence  $F_{184.9 \text{ nm}}$ , to generate the calibration plot presented in Fig. 5.



**Figure 5.** FAGE calibration for  $\text{CH}_3\text{O}$  at atmospheric pressure and 293 K; laser power  $P = (12.9 \pm 0.3) \text{ mW}$  and pressure in the detection cell of  $(2.65 \pm 0.05) \text{ Torr}$ . The FAGE signal, including the measurement with the Hg lamp turned off ( $[\text{CH}_3\text{O}] = 0$ ), was obtained after subtraction of the offline signal,  $(12.3 \pm 0.9) \text{ counts s}^{-1} \text{ mW}^{-1}$ . Averaging time per point = 120 s. The error limits in  $[\text{CH}_3\text{O}]$  and the FAGE signal for the  $x$  and  $y$  axes respectively are representative of the  $1\sigma$  overall uncertainty, which contains the total systematic and statistical errors (see text for details of these). The error limits shown in the legend are the standard errors in the slope and intercept of the fit to the experimental data.

## 2.3.2 Calibration for methyl peroxy radicals

### 2.3.2.1 Flow tube method

Methyl peroxy radicals were generated by water photolysis at 184.9 nm (Reaction (R7)) to give OH followed by the reaction with excess methane in air (BOC, synthetic BTCA 178) – Reactions (R1)–(R2) to give  $\text{CH}_3\text{O}_2$ . The calibrations were performed using the set-up described above. Methane (BOC, CP grade, 99.5 %) was flowed at 82.5 sccm to convert OH into  $\text{CH}_3$ , which subsequently reacted rapidly with  $\text{O}_2$  to form  $\text{CH}_3\text{O}_2$ . Figure S1 (Supplementary Information) shows an example of the OH signal with and without  $\text{CH}_4$ . The signal in the presence of  $\text{CH}_4$  was  $(0.04 \pm 0.04)$  of the signal in the absence of  $\text{CH}_4$  showing that  $(0.96 \pm 0.04)$  of OH was converted into  $\text{CH}_3\text{O}_2$ . The result is in agreement with the estimation of the fraction of OH titrated to  $\text{CH}_3\text{O}_2$ , 0.97, using a rate coefficient of  $6.4 \times 10^{-15} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  for the OH +  $\text{CH}_4$  reaction (Atkinson et al., 2006) and an average residence time of OH in the calibration flow tube of 11 ms determined using the volumetric flow rate and the geometric parameters of the flow tube and position of the Hg pen lamp.

The concentration of  $\text{CH}_3\text{O}_2$  was determined using Eq. (3):

$$[\text{CH}_3\text{O}_2] = 0.96 [\text{OH}] = 0.96 [\text{H}_2\text{O}] \sigma_{\text{H}_2\text{O}, 184.9 \text{ nm}} \Phi_{\text{H}_2\text{O}, 184.9 \text{ nm}} F_{184.9 \text{ nm}} \Delta t \quad (3)$$

where  $\sigma_{\text{H}_2\text{O}, 184.9 \text{ nm}}$  is the absorption cross section of water vapour at 184.9 nm,  $(7.22 \pm 0.22) \times 10^{-20} \text{ cm}^2 \text{ molecule}^{-1}$  (Cantrell et al., 1997; Creasey et al., 2000) and  $\Phi_{\text{H}_2\text{O}, 184.9 \text{ nm}}$  is the photodissociation quantum yield of OH, which is equal to unity. The values of  $F_{184.9 \text{ nm}}$  and  $\Delta t$  were determined as described in the Sect. 2.3.1. No loss of  $\text{CH}_3\text{O}_2$  by reaction with the  $\text{HO}_2$  radicals generated by the reaction of H atoms with  $\text{O}_2$  (R8) was encountered over the residence time of the radicals in the calibration flow tube ( $\sim 11 \text{ ms}$ ) as  $\text{CH}_3\text{O}_2$  reacts with  $\text{HO}_2$  on a ten second timescale as determined using a reaction rate coefficient of  $5.2 \times 10^{-12} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  (Atkinson et al., 2006) and the radical concentrations in the flow tube. The  $\text{CH}_3\text{O}_2$  radicals sampled through the FAGE pinhole expansion to a pressure of 2.65 Torr reached the detection region in about 85 ms while the calculated  $\text{CH}_3\text{O}_2 + \text{HO}_2$  reaction half-life at this reduced pressure in the FAGE inlet was thousands of seconds and any change in the  $\text{CH}_3\text{O}_2$  concentration is expected to be negligible.

Figure 6 shows results obtained from three separate calibration experiments. In the first two experiments air was humidified by passing a fraction of the air flow (40 slm total flow rate) through a deionised water bubbler. The hygrometer measured  $7.5 \times 10^{16} \text{ molecule cm}^{-3}$  of water vapour prior to the calibration flow tube and the concentration of methane in the flow tube was  $5 \times 10^{16} \text{ molecule cm}^{-3}$ . In the second experiment, a series of FAGE measurements were performed using a photon flux of  $\sim 1.6 \times 10^{14} \text{ photon cm}^{-2} \text{ s}^{-1}$  to generate  $\sim 4.5 \times 10^9 \text{ molecule cm}^{-3} \text{ CH}_3\text{O}_2$ . In the third experiment  $[\text{CH}_4] = 10^{17} \text{ molecule cm}^{-3}$  and all the air flow (now at 20 slm) was passed through the water bubbler to obtain  $3 \times 10^{17} \text{ molecule cm}^{-3} \text{ H}_2\text{O}$  vapour. The concentration of  $\text{CH}_3\text{O}_2$  was varied by changing the photon flux in the range of  $0.5\text{--}1.5 \times 10^{14} \text{ photon cm}^{-2} \text{ s}^{-1}$  to generate  $[\text{CH}_3\text{O}_2] = 1.5\text{--}4.5 \times 10^{10} \text{ molecule cm}^{-3}$ .

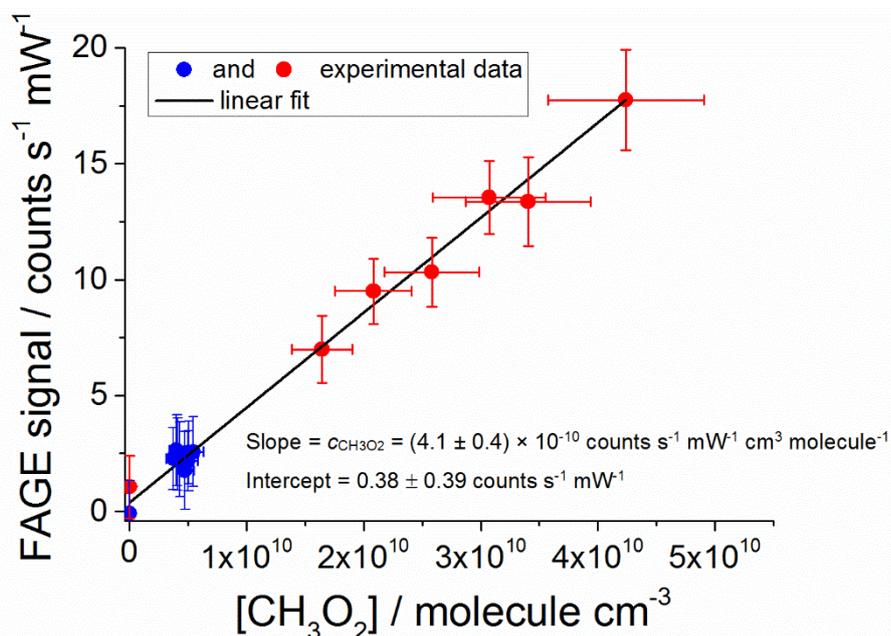


Figure 6. FAGE calibration for  $\text{CH}_3\text{O}_2$  at atmospheric pressure and 293 K. The data were obtained by three separate experiments: two of them generating  $[\text{CH}_3\text{O}_2] \cong 4.5 \times 10^9 \text{ molecule cm}^{-3}$  in the calibration flow tube (blue circles); laser power  $P = (9.5 \pm 0.3) \text{ mW}$  and  $(11.6 \pm 0.4) \text{ mW}$ , respectively and one experiment using  $[\text{CH}_3\text{O}_2]$  in the range of  $1.5\text{--}4.5 \times 10^{10} \text{ molecule cm}^{-3}$  (red circles);  $P = (9.2 \pm 0.2) \text{ mW}$ . The pressure in the FAGE detection cell was maintained  $(2.65 \pm 0.05) \text{ Torr}$  in all experiments. Averaging time per point = 120 s. The error limits in  $[\text{CH}_3\text{O}_2]$  and the FAGE signal for the  $x$  and  $y$  axes respectively are representative for the  $1\sigma$  overall uncertainty, which contains the total systematic and statistical errors. The error limits shown in the legend are the standard errors in the slope and intercept of the fit to the experimental data.

### 2.3.2.2 CH<sub>3</sub>O<sub>2</sub> second-order decay method

The principle behind this calibration method is that the second-order decay of CH<sub>3</sub>O<sub>2</sub> is dependent upon its initial concentration, and hence its quantification offers an alternative way to calibrate the signal. The experiments were performed in the HIRAC chamber at 295 K and 1 bar of synthetic air obtained by mixing high purity oxygen (BOC, > 99.999 %) and nitrogen (BOC, > 99.998 %) in the ratio of O<sub>2</sub>:N<sub>2</sub> = 1:4. Methane (BOC, CP grade, 2–3 × 10<sup>17</sup> molecule cm<sup>-3</sup>) and molecular chlorine (Sigma Aldrich, ≥ 99.5 %, 0.3–2.1 × 10<sup>14</sup> molecule cm<sup>-3</sup>) were delivered to the chamber. Eight UV black lamps (Phillips, TL-D 36W/BLB, λ = 350–400 nm) housed in quartz tubes mounted radially inside the reactive volume were used to photolyse Cl<sub>2</sub> to generate Cl atoms and initiate the chemistry:



Numerical simulations using the chemical system described in Table S3 in the Supplementary Information showed that [Cl]<sub>0</sub> = 1–6 × 10<sup>6</sup> molecule cm<sup>-3</sup> (varied by changing the initial [Cl<sub>2</sub>]). The high excess of methane (2–3 × 10<sup>17</sup> molecule cm<sup>-3</sup>) relative to [Cl]<sub>0</sub> ensured that the reactions of the Cl atoms with the self-reaction products formaldehyde and methanol were negligible. In each HIRAC experiment the lamps were alternatively turned on for 2–3 min and then off over 1–2 min to generate a series of typically 3–4 CH<sub>3</sub>O<sub>2</sub> kinetic decays.

In order to detect CH<sub>3</sub>O<sub>2</sub> the FAGE instrument was coupled to HIRAC through a custom-made ISO-K160 flange to sample the gas with a flow rate of ~ 3 slm. For most measurements, the 1 mm pinhole of the 280 mm long FAGE inlet was sampling ~230 mm from the chamber wall as in the OH measurements reported previously (Winiberg et al., 2015). Additional investigations into any CH<sub>3</sub>O<sub>2</sub> gradient across the ~ 600 mm radius of HIRAC were conducted using measurements of CH<sub>3</sub>O<sub>2</sub> formed by the CH<sub>4</sub> reaction with O(<sup>1</sup>D) generated by the photolysis of O<sub>3</sub> at 254 nm followed by the reaction of the produced CH<sub>3</sub> radical with O<sub>2</sub> at 295 K and 1 bar of synthetic air. An extended FAGE inlet (length 520 mm) was used to sample along 500 mm across the chamber starting with the inlet pinhole flush at the wall. A constant concentration of CH<sub>3</sub>O<sub>2</sub> was found (within the 10 % overall error of the measurement) for all the sampled distance 0 – 500 mm from the wall (note that 0 mm here refers to the FAGE inlet being at an equivalent position to the wall away from the mounting flange). The absence of a CH<sub>3</sub>O<sub>2</sub> gradient across the chamber provides evidence of the efficacy of the mixing in HIRAC and shows that the wall-loss of CH<sub>3</sub>O<sub>2</sub> is negligible and hence that a shorter inlet, and hence distance from inlet to CH<sub>3</sub>O<sub>2</sub> detection axis could be used in future CH<sub>3</sub>O<sub>2</sub> FAGE measurements within HIRAC, improving further the sensitivity.

### 2.4 Methoxy radical measurements within HIRAC

The experiment was carried out in HIRAC at 295 K and 1 bar of N<sub>2</sub> (BOC, > 99.998 %), but without any NO added to the FAGE cell (the cell furthest from the pinhole as shown in Fig. 1) so that [CH<sub>3</sub>O] is measured directly. Initial concentrations in HIRAC were: [CH<sub>4</sub>]<sub>0</sub> = 4.50 × 10<sup>17</sup> molecule cm<sup>-3</sup> and [Cl<sub>2</sub>]<sub>0</sub> = 5.57 × 10<sup>15</sup> molecule cm<sup>-3</sup>. After adding the reagents into the chamber the lamps (*vide supra*) were turned on to generate CH<sub>3</sub>O by Reaction (R6.b).

### 3 Results

#### 3.1 Sensitivity and detection limits for CH<sub>3</sub>O<sub>2</sub> and CH<sub>3</sub>O radicals obtained from calibrations

##### 3.1.1 Flow tube method

The FAGE sensitivity for CH<sub>3</sub>O<sub>2</sub> ( $C_{\text{CH}_3\text{O}_2}$ ) and CH<sub>3</sub>O ( $C_{\text{CH}_3\text{O}}$ ), is the slope of the linear regressions in Fig. 5 and Fig. 6, which were  $C_{\text{CH}_3\text{O}_2} = (4.1 \pm 1.4) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> and  $C_{\text{CH}_3\text{O}} = (5.1 \pm 2.2) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>. The error limits, 34 % for  $C_{\text{CH}_3\text{O}_2}$  and 43 % for  $C_{\text{CH}_3\text{O}}$ , are overall  $2\sigma$  uncertainties calculated using the sum in quadrature of the systematic uncertainties, 33 % for CH<sub>3</sub>O<sub>2</sub> and 42 % for CH<sub>3</sub>O (details in Section 3.2.1), and the statistical errors from the calibration plots, ~ 8 %. The higher errors in  $C_{\text{CH}_3\text{O}}$  compared to  $C_{\text{CH}_3\text{O}_2}$  are due to the uncertainty in the methanol concentration, which is not determined directly (*vide supra*),  $1\sigma \cong 7\%$  and the error in the yield of CH<sub>3</sub>O. The value of the CH<sub>3</sub>O photolysis yield from CH<sub>3</sub>OH reported at 193 nm was used ( $0.86 \pm 0.10$ ), which has an uncertainty of 11.63 % at the  $1\sigma$  level (Satyapal et al., 1989).

From the sensitivity factor,  $C$ , the limit of detection ( $LOD$ ) was calculated using Eq. (4) and assuming Poisson statistics appropriate for single photon counting:

$$LOD(\text{CH}_3\text{O}_2) = \frac{S/N}{C_{\text{CH}_3\text{O}_2} P} \sqrt{\frac{BKG}{t} \left( \frac{1}{m} + \frac{1}{n} \right)}, \quad LOD(\text{CH}_3\text{O}) = \frac{S/N}{C_{\text{CH}_3\text{O}} P} \sqrt{\frac{BKG}{t} \left( \frac{1}{m} + \frac{1}{n} \right)}, \quad (4)$$

where  $S/N$  is the signal-to-noise ratio,  $P$  is the laser power,  $BKG$  is the background signal and had a typical value of ~100 counts s<sup>-1</sup>, which represents ~50 counts s<sup>-1</sup> laser scattered light within the detection cell and ~50 counts s<sup>-1</sup> scattered visible light which enters the pinhole from the room with a negligible contribution (1 count s<sup>-1</sup> on average) of the detector dark counts,  $t$  is the time per data point,  $m$  represents the number of online data points and  $n$  is the number of offline data points. For a typical 5 min averaged signal, i.e.  $m = n = 150$ ,  $S/N = 2$ ,  $P = 15$  mW and  $t = 1$  s, and using the values of  $C$  from the calibration,  $LOD(\text{CH}_3\text{O}_2) = 3.8 \times 10^8$  molecule cm<sup>-3</sup> and  $LOD(\text{CH}_3\text{O}) = 3.0 \times 10^8$  molecule cm<sup>-3</sup>. An increase of the averaging time to 1 hour, i.e.  $m = n = 1800$  data points, results in a decrease of the detection limits to  $LOD(\text{CH}_3\text{O}_2) = 1.1 \times 10^8$  molecule cm<sup>-3</sup> and  $LOD(\text{CH}_3\text{O}) = 8.7 \times 10^7$  molecule cm<sup>-3</sup>.

Although CH<sub>3</sub>O<sub>2</sub> has not been measured specifically in the atmosphere, there have been several calculations of its concentration using numerical models. In general, the concentration of CH<sub>3</sub>O<sub>2</sub> is a function both of the loadings of volatile organic compounds (VOCs) and the levels of NO<sub>x</sub>. For the clean, remote environments at Cape Verde in the tropical Atlantic ocean and in the Borneo rainforest [CH<sub>3</sub>O<sub>2</sub>] is calculated to peak around  $6 \times 10^8$  molecule cm<sup>-3</sup> and about  $2 \times 10^8$  molecule cm<sup>-3</sup>, respectively at noon using the modeling studies reported by Whalley et al. (Whalley et al., 2010; Whalley et al., 2011). Therefore, it should be possible using the FAGE conversion method to CH<sub>3</sub>O and for an averaging time of 1 hour (*vide supra*) to achieve a measurement of atmospheric levels of CH<sub>3</sub>O<sub>2</sub> in such clean environments, and shorter averaging times in some cases. Further optimizations of FAGE sensitivity can be achieved by the removal of the fibre optic cables to deliver the probe laser beam directly to the CH<sub>3</sub>O detection cell to increase the laser power and, by increasing the pulse repetition frequency above the current value of 5 kHz (but without significant reduction in the pulse energy). The present investigations into the change of sensitivity with pressure in the range from 2.65–10.00 Torr found that 2.65 Torr is the optimum value in this pressure interval. The result suggests that, by reducing the pressure in the above range of values, the decrease in fluorescence due to the reduction in the CH<sub>3</sub>O number density was overcome by the increase in the fluorescence quantum yield due to a lower fluorescence quenching rate. Another reason could be that the characteristics of the jet expansion and/or the ensuing flow to the LIF detection region change with pressure, leading to a more favourable transmission of radicals to the detection region, but it is difficult to test this experimentally. Hence an additional improvement in the sensitivity might be obtained by using a lower detection cell pressure than the current value of 2.65 Torr using a more powerful pump. It should be also noted that the distance from the inlet pinhole to the laser-axis in the CH<sub>3</sub>O and CH<sub>3</sub>O<sub>2</sub> fluorescence cell

(Figure 1, ~ 580 mm) is considerably longer than the corresponding distance in the ground-based field fluorescence cell for OH and HO<sub>2</sub> detection (88 mm), and improvements in sensitivity would be expected for a shorter pinhole-to-laser excitation distance for CH<sub>3</sub>O<sub>2</sub>. The further optimizations of sensitivity and the planned construction of a reference cell to find the online wavelength position could potentially enable CH<sub>3</sub>O<sub>2</sub> measurements to be made in urban environments where CH<sub>3</sub>O<sub>2</sub> concentrations are estimated to be considerably lower, for example a few 10<sup>7</sup> molecule cm<sup>-3</sup> based on modeling results (Whalley et al., to be submitted).

The calibrations using the flow tube (“wand”) method have been performed under water vapour concentrations similar to the ambient [H<sub>2</sub>O<sub>vapour</sub>] but few orders of magnitude higher than those present in the HIRAC chamber experiments. In contrast with [H<sub>2</sub>O<sub>vapour</sub>], the methane concentrations used in the “wand” method were similar to [CH<sub>4</sub>] present in HIRAC but higher than [CH<sub>4</sub>] in the atmosphere. However, as detailed in this paragraph, the effects of methane and water on our sensitivity are minimal. Estimations using the reported fluorescence quenching rate coefficient of CH<sub>3</sub>O(A) by CH<sub>4</sub>,  $k_{\text{quench,CH}_4} = 1.05 \times 10^{-10} \text{ s}^{-1}$ , (Wantuck et al., 1987) and the concentrations of CH<sub>4</sub> in the LIF detection cell for the calibrations using the flow-tube ( $1.7 \times 10^{14}$  molecule cm<sup>-3</sup> and  $3.4 \times 10^{14}$  molecule cm<sup>-3</sup>, corresponding to  $5.0 \times 10^{16}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{17}$  molecule cm<sup>-3</sup>, respectively in the flow tube) resulted in only ~ 1–2% lower fluorescence quantum yield compared to the value determined in the absence of CH<sub>4</sub>. No literature value has been found for the fluorescence rate coefficient of CH<sub>3</sub>O(A) fluorescence by H<sub>2</sub>O vapour. However, even if it assumed to be as large as the above reported value for CH<sub>4</sub> ( $k_{\text{quench,CH}_4}$ ), only a few percent decrease in the fluorescence quantum yield is computed (compared with a water concentration of zero) for the levels of H<sub>2</sub>O vapour which are present at the CH<sub>3</sub>O<sub>2</sub> FAGE detection axis when using the “wand” calibration method. These levels (1–2% v/v) are similar to a typical water vapour concentration in the atmosphere. A very good agreement has been obtained between the calibration factors for CH<sub>3</sub>O<sub>2</sub> detection with two different concentrations of water vapour in the flow tube:  $7.5 \times 10^{16}$  molecule cm<sup>-3</sup> or  $3.0 \times 10^{17}$  molecule cm<sup>-3</sup> (corresponding to  $2.6 \times 10^{14}$  molecule cm<sup>-3</sup> and  $1.0 \times 10^{15}$  molecule cm<sup>-3</sup>, respectively in the FAGE cell) as shown in Figure 6 in Sect. 2.3.2.1. This very good agreement for H<sub>2</sub>O vapour and the above calculations for CH<sub>4</sub> support the use of the flow tube method for the FAGE calibration of the CH<sub>3</sub>O<sub>2</sub> concentrations.

### 3.1.2 Methyl peroxy calibration using kinetics of the CH<sub>3</sub>O<sub>2</sub> second-order decay

An alternative method of calibration for CH<sub>3</sub>O<sub>2</sub> was to generate CH<sub>3</sub>O<sub>2</sub> radicals in HIRAC to monitor the temporal decay of the CH<sub>3</sub>O<sub>2</sub> FAGE signal once the photolysis lamps were turned off. Figure 7 shows an example of a decay in the CH<sub>3</sub>O<sub>2</sub> signal generated by extinguishing the HIRAC lamps following the production of CH<sub>3</sub>O<sub>2</sub> by the Cl atom initiated oxidation of CH<sub>4</sub> in the presence of O<sub>2</sub> (Reactions (R12) and (R2)). In the absence of other processes, the loss of CH<sub>3</sub>O<sub>2</sub> is described by the integrated second-order rate law equation describing the CH<sub>3</sub>O<sub>2</sub> self-reaction (Reaction (R6)):

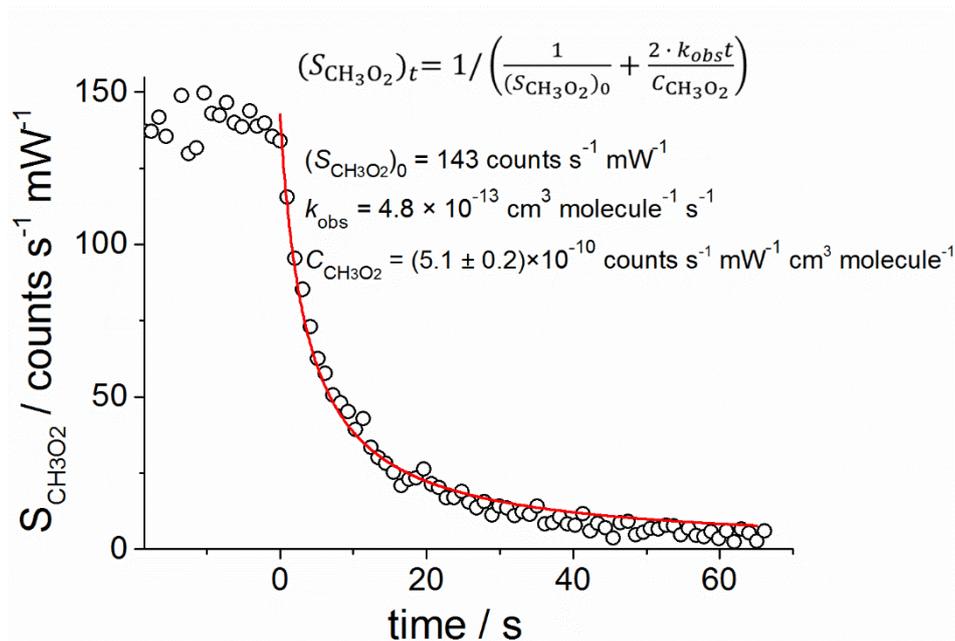
$$\frac{1}{[\text{CH}_3\text{O}_2]_t} = \frac{1}{[\text{CH}_3\text{O}_2]_0} + 2 \cdot k_{\text{obs}} t, \quad (5)$$

where [CH<sub>3</sub>O<sub>2</sub>]<sub>t</sub> is the methyl peroxy concentration at reaction time *t*, [CH<sub>3</sub>O<sub>2</sub>]<sub>0</sub> is the initial concentration when the lights are switched off and  $k_{\text{obs}}$  is the observed rate coefficient (which is not equal to  $k_6$ , see below). Using  $[\text{CH}_3\text{O}_2] = \frac{S_{\text{CH}_3\text{O}_2}}{C_{\text{CH}_3\text{O}_2}}$ , where  $S_{\text{CH}_3\text{O}_2}$  is the signal measured by FAGE and  $C_{\text{CH}_3\text{O}_2}$  is the instrument sensitivity, Eq. (6) is obtained for the temporal profile of the methyl peroxy signal:

$$\frac{1}{(S_{\text{CH}_3\text{O}_2})_t} = \frac{1}{(S_{\text{CH}_3\text{O}_2})_0} + \frac{2 \cdot k_{\text{obs}} t}{C_{\text{CH}_3\text{O}_2}} \quad \text{or} \quad (S_{\text{CH}_3\text{O}_2})_t = 1 / \left( \frac{1}{(S_{\text{CH}_3\text{O}_2})_0} + \frac{2 \cdot k_{\text{obs}} t}{C_{\text{CH}_3\text{O}_2}} \right), \quad (6)$$

In Eq. (6)  $(S_{\text{CH}_3\text{O}_2})_t$  and  $(S_{\text{CH}_3\text{O}_2})_0$  are the signal at time  $t$  and  $t = 0$  respectively.

Eq. (6) was fitted to the experimental decays of  $S_{\text{CH}_3\text{O}_2}$  (see Fig. 7 as an example) fixing  $k_{\text{obs}}$  to the IUPAC recommendation,  $k_{\text{obs}} = (4.8 \pm 1.1) \times 10^{-13} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$ , in order to obtain  $C_{\text{CH}_3\text{O}_2}$ . Eighteen  $\text{CH}_3\text{O}_2$  decays were analysed, which yielded an average value of  $C_{\text{CH}_3\text{O}_2} = (5.6 \pm 1.7) \times 10^{-10} \text{ counts cm}^3 \text{ molecule}^{-1} \text{ s}^{-1} \text{ mW}^{-1}$ . The error limit, 30 %, is the  $2\sigma$  composite error calculated as the sum in quadrature of the total systematic uncertainty, 29 % (see Section 3.2.2), and the average random error of all determinations, with 8 %, taken as two standard errors in the fit of Eq. (6) to the  $\text{CH}_3\text{O}_2$  temporal decays. This value agrees well with  $C_{\text{CH}_3\text{O}_2} = (4.1 \pm 1.4) \times 10^{-10} \text{ counts cm}^3 \text{ molecule}^{-1} \text{ s}^{-1} \text{ mW}^{-1}$  obtained from the flow-tube calibration method (section 3.1.1).



**Figure 7.** Second-order decay of the normalized  $\text{CH}_3\text{O}_2$  signal with 1 second time resolution generated in HIRAC using  $\text{Cl}/\text{CH}_4/\text{O}_2$  and black lamps (see main text for details);  $[\text{CH}_4]_0 = 2.3 \times 10^{17} \text{ molecule cm}^{-3}$  and  $[\text{Cl}_2]_0 = 5.8 \times 10^{13} \text{ molecule cm}^{-3}$  at 295 K and 1 bar mixture of  $\text{N}_2:\text{O}_2 = 4:1$ . At time zero the lamps were turned off. Fitting Eq. (6) to the data yielded  $C_{\text{CH}_3\text{O}_2} = (5.1 \pm 0.2) \times 10^{-10} \text{ counts cm}^3 \text{ molecule}^{-1} \text{ s}^{-1} \text{ mW}^{-1}$  (statistical error at  $1\sigma$  level).

Based on the lack of a measurable  $\text{CH}_3\text{O}_2$  radical gradient across HIRAC (Section 2.3.2.2, *vide supra*) it is assumed that the loss of  $\text{CH}_3\text{O}_2$  to the walls of HIRAC in these experiments was negligible over the timescale of 1–2 min of the temporal decay measurements. Our finding is consistent with previous results showing that the heterogeneous wall-loss rates for  $\text{CH}_3\text{O}_2$  were significantly lower than the corresponding removal rates of  $\text{HO}_2$  (Miyazaki et al., 2010; Mihele et al., 1999; Fuchs et al., 2008). Using a 30 cm long glass tube of 2 cm diameter, Miyazaki et al. measured that the heterogeneous removal efficiency for  $\text{CH}_3\text{O}_2$  was six times lower than for  $\text{HO}_2$ . The  $\text{HO}_2$  wall-loss rate coefficient at room temperature and 1000 mbar in HIRAC was found to be of  $\sim 10^{-2} \text{ s}^{-1}$  (Winiberg et al., 2015). Therefore, it can be expected that the wall-loss rate coefficient of  $\text{CH}_3\text{O}_2$  in HIRAC was  $k_{\text{loss}} \cong 10^{-3} \text{ s}^{-1}$  and so is not considered in the analysis here for  $\text{CH}_3\text{O}_2$  decays which typically last for  $\sim 100 \text{ s}$ . In order to investigate the sensitivity of  $C_{\text{CH}_3\text{O}_2}$  obtained by the kinetic analysis of the  $\text{CH}_3\text{O}_2$  decay to  $k_{\text{loss}}$  higher than  $10^{-3} \text{ s}^{-1}$ , a wall-loss rate coefficient of  $10^{-2} \text{ s}^{-1}$  was included in the analysis of the experimental decays of  $\text{CH}_3\text{O}_2$  to obtain  $C_{\text{CH}_3\text{O}_2}$ , but only an increase in  $C_{\text{CH}_3\text{O}_2}$  of about 6 % on average was seen. A small deviation of the experimental data from the fit was obtained at the end of the measurements

whether or not  $k_{\text{loss}}$  was included in the analysis (Fig. 7). The role of potential secondary chemistry at later times of the reaction will be investigated in future kinetic studies of the  $\text{CH}_3\text{O}_2$  self-reaction.

Using the average sensitivity factor  $C_{\text{CH}_3\text{O}_2} = (5.6 \pm 1.7) \times 10^{-10}$  counts  $\text{cm}^3$  molecule $^{-1}$  s $^{-1}$  mW $^{-1}$  determined by the  $\text{CH}_3\text{O}_2$  decay method in HIRAC, and for a signal to noise ratio  $S/N = 2$ , a laser power  $P = 15$  mW and a time per data point  $t = 1$  s in Eq. (4) results in an improved (compared with the flow tube calibration)  $\text{LOD}(\text{CH}_3\text{O}_2) = 2.8 \times 10^8$  molecule  $\text{cm}^{-3}$  for 5 min averaging time, i.e. 150 online data points ( $m$ ) and 150 offline points ( $n$ ). The corresponding  $\text{LOD}$  for an averaging time of 1 hour, i.e.  $m = n = 1800$  is  $\text{LOD}(\text{CH}_3\text{O}_2) = 7.9 \times 10^7$  molecule  $\text{cm}^{-3}$ .

It should be noted that the observed rate coefficient,  $k_{\text{obs}}$ , is larger than the second-order rate coefficient of just the  $\text{CH}_3\text{O}_2$  recombination reaction (R6),  $k_6$ , as the methoxy radicals generated by channel R6.b react rapidly with molecular oxygen present in large excess,  $5 \times 10^{18}$  molecule  $\text{cm}^{-3}$ , to produce  $\text{HO}_2$  (R4) which in turn reacts with  $\text{CH}_3\text{O}_2$  (R13):



As each  $\text{HO}_2$  radical consumes one  $\text{CH}_3\text{O}_2$  species (R13) on the time scale of the reaction (R6),  $k_{\text{obs}}$  is given by (Sander and Watson, 1981; Lightfoot et al., 1990):

$$k_{\text{obs}} = k_6 \cdot (1 + r_{6,\text{b}}) \quad (7)$$

where  $r_{6,\text{b}}$  is the branching ratio for the reaction channel R6.b. According to IUPAC (Atkinson et al., 2006), there is a 23 % uncertainty in  $k_{\text{obs}}$  of the  $\text{CH}_3\text{O}_2$  recombination at 298 K with a recommended value  $k_{\text{obs}} = 4.8 \times 10^{-13}$   $\text{cm}^3$  molecule $^{-1}$  s $^{-1}$ . This value corresponds to  $k_6 = (3.5 \pm 1.0) \times 10^{-13}$   $\text{cm}^3$  molecule $^{-1}$  s $^{-1}$  and  $r_{6,\text{b}} = 0.37 \pm 0.06$  (Atkinson et al., 2006).

In order to check the validity of Eq. (7) in the presence of  $\text{HO}_2$  removal by self-reaction and wall-loss, numerical simulations were performed to generate  $\text{CH}_3\text{O}_2$  decays using a system incorporating the chemistry described by Reactions (R4), (R6), (R13) and (R14) (*vide infra*) and a heterogeneous loss of  $\text{HO}_2$ ,  $k_{\text{loss}(\text{HO}_2)}$  (Supplementary Information). The rate coefficients were sourced from the IUPAC preferred values at 298 K (Table S3 in Supplementary Information) and  $k_{\text{loss}(\text{HO}_2)}$  was varied. The simulated decays of  $[\text{CH}_3\text{O}_2]$  vs. time were analysed using Eq. (5) (see Fig. S3 as an example) and gave an average observed rate coefficient of  $k_{\text{obs}} = 4.7 \times 10^{-13}$   $\text{cm}^3$  molecule $^{-1}$  s $^{-1}$ , which is only 2 % lower than the IUPAC recommendation, for  $k_{\text{loss}(\text{HO}_2)}$  varied between 0.01–0.10 s $^{-1}$  and, hence confirm the applicability of Eq. (7).



## 3.2 Calibration uncertainties

### 3.2.1 Flow tube method

The  $2\sigma$  error associated with  $C_{\text{CH}_3\text{O}_2}$  of 34 % obtained by the flow tube method ( $C_{\text{CH}_3\text{O}_2} = (4.1 \pm 1.4) \times 10^{-10}$  counts  $\text{cm}^3$  molecule $^{-1}$  s $^{-1}$  mW $^{-1}$ ), represents the overall uncertainty calculated using the sum in quadrature of the systematic uncertainty, 33 %, and the

statistical error from the calibration plots,  $\sim 8\%$ . The overall 34 % uncertainty is similar to the previous estimated total uncertainty, 36 %, in the use of the same method for calibration of OH and HO<sub>2</sub> measurements in HIRAC, where no CH<sub>4</sub> is added (Winiberg et al., 2015). The flow tube method is a proven method to generate known amounts of OH and HO<sub>2</sub> by the photolysis of H<sub>2</sub>O at 184.9 nm in order to calibrate field instruments (Heard and Pilling, 2003).

The largest contribution to the total error of the method came from the 28 % total uncertainty in the photon flux of the calibration source,  $F_{184.9\text{nm}}$ . The product  $F_{184.9\text{nm}} \times \Delta t$  is determined using N<sub>2</sub>O actinometry relying on the measurement of [NO] in trace amounts (0.2–1.5 ppbv) using a commercial NO analyser (Thermo Electron Corporation 42C) followed by the data analysis using four rate coefficients each with  $\sim 10\%$  uncertainty (Burkholder et al., 2015). Although it is the product  $F_{184.9\text{nm}} \times \Delta t$  which is directly determined by the actinometric method and used to calculate the concentration of radicals to calibrate FAGE (Eq. (3)), any difference in the volumetric flow rate between the calibration and actinometry experiments will change  $\Delta t$ . Therefore, the uncertainty in  $\Delta t$ , 2 %, needs to be accounted for. The contributions from the rest of the terms in Eq. (3) to the systematic uncertainty in the determination of [CH<sub>3</sub>O<sub>2</sub>] by this method were as follows: 6 % total error in  $\sigma_{\text{H}_2\text{O}, 184.9\text{nm}}$  (Cantrell et al., 1997), 10 % uncertainty in [H<sub>2</sub>O], taken from the instrumental uncertainty of the hygrometer and 4 % error in the yield of CH<sub>3</sub>O<sub>2</sub> produced by the OH conversion into CH<sub>3</sub> followed by the CH<sub>3</sub> + O<sub>2</sub> reaction. The contribution of the uncertainties in the FAGE measurements to the 33 % overall systematic uncertainty in the calibration were estimated to consist of 12 % in the online FAGE signal and 6 % uncertainty in the laser power measured by the laser power meter and used to normalize the data. The uncertainty associated with the online signal, 12 % at 2 $\sigma$  level, was calculated as the average deviation of the signal value due to the error limits of  $\pm 5 \times 10^{-4}$  nm in the online wavelength position (see the typical laser excitation scans shown in Fig. 3).

### 3.2.2 CH<sub>3</sub>O<sub>2</sub> second-order decay calibration

The largest contribution to the calculated overall 2 $\sigma$  uncertainty of 30 % in  $C_{\text{CH}_3\text{O}_2}$  obtained by the CH<sub>3</sub>O<sub>2</sub> second-order decay method ( $C_{\text{CH}_3\text{O}_2} = (5.6 \pm 1.7) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>), derives from the 23 % error in the IUPAC preferred value of the observed rate coefficient for the effective CH<sub>3</sub>O<sub>2</sub> self-reaction,  $k_{\text{obs}} = (4.8 \pm 1.1) \times 10^{-13}$  cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> (Atkinson et al., 2006). It is instructive to examine the origin of the 23 % error. The studies which led to the IUPAC recommendation utilized the UV-absorption of CH<sub>3</sub>O<sub>2</sub>, typically at 250 nm, and the determined quantity was the ratio between the observed rate coefficient and the absorption cross section of CH<sub>3</sub>O<sub>2</sub>,  $k_{\text{obs}}/\sigma_{250\text{nm}}$ . IUPAC and the Jet Propulsion Laboratory (JPL) recommend  $3.9 \times 10^{-18}$  and  $3.8 \times 10^{-18}$  cm<sup>2</sup> molecule<sup>-1</sup>, respectively for  $\sigma_{250\text{nm}}$  (Atkinson et al., 2006; Burkholder et al., 2015). The JPL recommendation (Burkholder et al., 2015) is the cross section obtained by the re-evaluation of the previous reported UV-absorption spectra by Tyndall et al. in 2001 (Tyndall et al., 2001), yielding  $\sigma_{250\text{nm}} = 3.78 \times 10^{-18}$  cm<sup>2</sup> molecule<sup>-1</sup>. Tyndall et al. used a cross section of  $(4.26 \pm 0.52) \times 10^{-18}$  cm<sup>2</sup> molecule<sup>-1</sup> (error at 2 $\sigma$ ) for the maximum at 237.3 nm, obtained by analysing the shape of the absorption spectra between 200–300 nm reported since 1990. The studies before 1990 were not included due to errors in the calibration of the CH<sub>3</sub>O<sub>2</sub> cross section leading to large discrepancies in the reported values. The 2001 evaluation of Tyndall et al. calculated  $k_{\text{obs}} = (4.7 \pm 0.8) \times 10^{-13}$  cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> where the error limits are two standard deviations of the mean. Including an error of 10 % in the cross section of CH<sub>3</sub>O<sub>2</sub>, as suggested by the authors (Tyndall et al., 2001), a 19 % composite uncertainty in  $k_{\text{obs}}$  is obtained. The result is in good agreement with the 23 % uncertainty in the IUPAC recommendation.

The remaining contributions to the uncertainty in the calibration using the CH<sub>3</sub>O<sub>2</sub> second-order decay method are: 6 % error in the laser power, 12 % uncertainty in the online signal determined by how well the laser is able to find the online wavelength position (*vide supra*) and 10 % error in  $(S_{\text{CH}_3\text{O}_2})_0$  in Eq. (6), the value of the CH<sub>3</sub>O<sub>2</sub> signal at the moment when the HIRAC lamps were turned off to generate a second-order decay.

### 3.2.3 Comparison between the FAGE sensitivities for CH<sub>3</sub>O<sub>2</sub> obtained by the two calibration methods

The FAGE sensitivity factor obtained using the flow tube method,  $C_{\text{CH}_3\text{O}_2} = (4.1 \pm 1.4) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>, is 27 % lower but has overlapping error limits with the result found using the CH<sub>3</sub>O<sub>2</sub> second-order decay method,  $C_{\text{CH}_3\text{O}_2} = (5.6 \pm 1.7) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> (uncertainties quoted to  $2\sigma$ ). The calculated overall error in the CH<sub>3</sub>O<sub>2</sub> second-order decay method, 30 %, is similar to the total uncertainty in the flow tube method, 34 %. The flow tube method is known to reliably generate accurate concentrations of radicals and has been used for many years in the calibration of FAGE instruments employed in field measurements of OH and HO<sub>2</sub> (Heard and Pilling, 2003). The flow tube method has also been validated by using alternate methods of calibration, for example using the decay of a hydrocarbon in the HIRAC chamber to obtain [OH] (Winiberg et al., 2015). The method of using a time-resolved kinetic quantity to derive a calibration factor was validated for HO<sub>2</sub> in HIRAC, where  $C_{\text{HO}_2}$  obtained from analysis of the temporal decay of HO<sub>2</sub> agreed with  $C_{\text{HO}_2}$  from the flow tube method (Winiberg et al., 2015). These results suggest that the sensitivity of the FAGE system, represented by the value of  $C$ , is not changed between sampling from the calibration flow tube and sampling from within HIRAC itself.

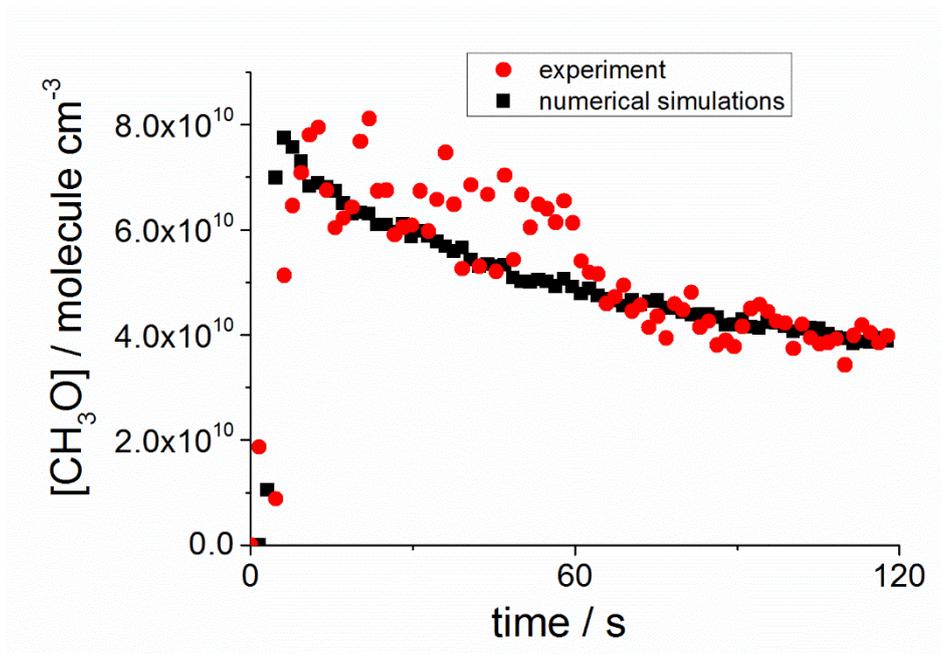
The accuracy of the CH<sub>3</sub>O<sub>2</sub> temporal decay method is largely determined by the accuracy of  $k_{\text{obs}}$  (see section 3.2.2. above). The quantity measured in the previous kinetic studies of CH<sub>3</sub>O<sub>2</sub> + CH<sub>3</sub>O<sub>2</sub> is  $k_{\text{obs}}/\sigma_{250\text{nm}}$  and hence the accuracy of  $k_{\text{obs}}$  is directly affected by any systematic errors in the determination of  $\sigma_{250\text{nm}}$ . In order to make  $C_{\text{CH}_3\text{O}_2}$  derived from the temporal decay and flow tube methods of the same, the value of  $k_{\text{obs}}$  would need to be reduced by ~ 25 %, which in turn requires a ~ 25 % reduction in  $\sigma_{250\text{nm}}$ . It is noted that the UV-absorption spectrum of CH<sub>3</sub>O<sub>2</sub> is relatively broad and hence may prevent a selective detection due to the difficulty to discriminate from the potential presence of other species also absorbing around 250 nm, such as Cl<sub>2</sub> and CH<sub>3</sub>CHO used in concentrations as high as 10<sup>16</sup> molecule cm<sup>-3</sup>, while [CH<sub>3</sub>O<sub>2</sub>] was ~ 10<sup>13</sup> molecule cm<sup>-3</sup> (Dagaut and Kurylo, 1990;Roehl et al., 1996). As the absorption cross sections of Cl<sub>2</sub> and CH<sub>3</sub>CHO at 250 nm lay in the range 10<sup>-21</sup>–10<sup>-22</sup> cm<sup>2</sup> molecule<sup>-1</sup> (Keller-Rudek et al., 2013), the unaccounted for absorption of these species may have led to an overestimation of  $\sigma_{250\text{nm}}(\text{CH}_3\text{O}_2)$ .

As noted in the 2001 review by Tyndall et al. (Tyndall et al., 2001), none of the previous laboratory studies of the CH<sub>3</sub>O<sub>2</sub> recombination measured [CH<sub>3</sub>O<sub>2</sub>] by any method other than UV-spectroscopy. In addition, the traditional time-resolved measurements of CH<sub>3</sub>O<sub>2</sub> used high CH<sub>3</sub>O<sub>2</sub> concentrations (10<sup>13</sup>–10<sup>15</sup> molecule cm<sup>-3</sup>) and, as the self-reaction is fairly slow, Tyndall et al. stated that the results were potentially affected by secondary chemistry (Tyndall et al., 2001). Therefore, there is a need for the use of a complementary technique in the kinetic study of this reaction, for example by LIF as described in this paper, which may offer some advantages to probe CH<sub>3</sub>O<sub>2</sub> selectively in the absence of interferences from other species. In addition, LIF is more sensitive and hence requires significantly lower radical concentrations ( $[\text{CH}_3\text{O}_2]_0 = (1-3) \times 10^{11}$  molecule cm<sup>-3</sup> here) than for the UV-absorption studies which may help to minimize potential secondary chemistry.

### 3.3 Methoxy radical measurement within HIRAC

The typical concentration of [O<sub>2</sub>] = 5 × 10<sup>18</sup> molecule cm<sup>-3</sup> used in the HIRAC experiments described above was lowered in some experiments to decrease the consumption of CH<sub>3</sub>O by O<sub>2</sub> via Reaction (R4). In this manner, a concentration of methoxy radicals was obtained above the FAGE limit of detection in HIRAC to enable a direct measurement over few minutes. The chamber was filled with high purity nitrogen (> 99.998 %), but the ~ 6 m long N<sub>2</sub> delivery pipe was purposely incompletely purged before the experiment in order to deliver trace levels of oxygen to HIRAC. The initial Cl<sub>2</sub> concentration in these experiments was 5.6 × 10<sup>15</sup> molecule cm<sup>-3</sup> and hence is 1–2 orders of magnitude higher than [Cl<sub>2</sub>]<sub>0</sub> used in the kinetic experiments above in order to generate higher [Cl] and hence [CH<sub>3</sub>O]. The concentration of CH<sub>3</sub>O during the experiment was computed by using the FAGE calibration factor for methoxy radicals generated from the photolysis of methanol in N<sub>2</sub>,  $C_{\text{CH}_3\text{O}} = (5.1 \pm 2.2) \times 10^{-10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> (Sect. 3.1.1). The temporal profile of CH<sub>3</sub>O is shown in Fig. 8, together with a numerical

simulation of  $\text{CH}_3\text{O}(t)$  using a chemistry system described in the Supplementary Information. The best fit to the experimental  $\text{CH}_3\text{O}$  concentration profile was obtained for  $[\text{O}_2] = (5.4 \pm 0.6) \times 10^{15} \text{ molecule cm}^{-3}$ , i.e. around 0.02 % relative to  $\text{N}_2$ . The numerical simulations showed that  $\text{Cl}_2$  consumption was dominated by the reaction with  $\text{CH}_3$  radicals, present at a relatively high concentration, explaining the  $\sim 50\%$  decrease in  $[\text{CH}_3\text{O}]$  observed during its temporal measurement shown in Fig. 8. The Supplementary Information (Fig. S5) shows the concentration profiles of  $\text{Cl}_2$ ,  $\text{Cl}$ ,  $\text{CH}_3$  and  $\text{CH}_3\text{O}_2$  obtained by numerical simulations performed over  $\sim 2$  min.



**Figure 8.** Concentration of  $\text{CH}_3\text{O}$  as a function of time in HIRAC: red points are the experimental data and black points are generated by a numerical simulation.  $\text{CH}_3\text{O}$  radicals were formed as a product of the self-reaction of  $\text{CH}_3\text{O}_2$  species at 295 K and 1 bar of  $\text{N}_2$ , with  $\text{CH}_3\text{O}_2$  generated by the reaction of  $\text{Cl}$  atoms with  $\text{CH}_4$ , with the HIRAC black lamps being turned on at time zero. Oxygen was present in trace amounts, determined to be  $(5.4 \pm 0.6) \times 10^{15} \text{ molecule cm}^{-3}$  from comparison of the simulations to the experimental data. The chemical mechanism used in the numerical simulations is presented in the Supplementary Information.

These results demonstrate the capability to measure an absolute concentration of  $\text{CH}_3\text{O}$  radicals in a simulation chamber, with  $\text{CH}_3\text{O}$  representing a further model target species for the validation of chemical mechanisms for the chemical oxidation of VOCs. However, it is recognized that the experiments need to be performed at reduced  $[\text{O}_2]$ , and that  $[\text{O}_2]$  needs to be known *a priori* in order to test robustly the accuracy of the chemical mechanism and underlying kinetic parameters.

#### 4. Conclusions

Currently there is no measurement of the absolute concentration of  $\text{CH}_3\text{O}_2$  radicals in the atmosphere. In this work the FAGE technique has been extended by adding the capability to detect  $\text{CH}_3\text{O}_2$  and  $\text{CH}_3\text{O}$  radicals to the more typical measurement of  $\text{OH}$  and  $\text{HO}_2$  radicals. The method enables the speciated and sensitive detection of  $\text{CH}_3\text{O}_2$  radicals by converting  $\text{CH}_3\text{O}_2$  into  $\text{CH}_3\text{O}$  by reaction with  $\text{NO}$  and detecting the resultant  $\text{CH}_3\text{O}$  by LIF with excitation at *ca.* 298 nm. The limit of detection of the method obtained using the radical source commonly employed to provide accurate concentrations of  $\text{OH}$  with added  $\text{CH}_4$ , is  $3.8 \times 10^8 \text{ molecule cm}^{-3}$  for a signal-to-noise ratio of 2 and 5 min time resolution and reduces to  $1.1 \times 10^8 \text{ molecule cm}^{-3}$  for  $S/N = 2$  and 1

hour averaging time. Therefore, the method has the potential to be used in field measurements of the diurnal profiles of CH<sub>3</sub>O<sub>2</sub> in clean air with low NO<sub>x</sub> levels, such as remote continental environments and in the marine boundary layer. Further improvements of the FAGE sensitivity could be achieved via the increase in the laser repetition frequency above the current value of 5 kHz, a decrease in the detection chamber pressure (currently ~ 2.65 Torr), and the use of a shorter distance between the inlet sampling pinhole and the fluorescence detection axis (presently a long distance of ~ 580 mm). The method is also demonstrated for the direct detection of CH<sub>3</sub>O, in the absence of added NO to the fluorescence cell. The limit of detection for CH<sub>3</sub>O determined using the conventional radical source for  $S/N = 2$  and 5 min averaging time is  $3.0 \times 10^8$  molecule cm<sup>-3</sup>.

Additional investigations into the FAGE sensitivity for CH<sub>3</sub>O<sub>2</sub> were carried out in the HIRAC simulation chamber at Leeds, by studying the kinetics of the second-order decays of CH<sub>3</sub>O<sub>2</sub> by its self-reaction. The second-order decays of CH<sub>3</sub>O<sub>2</sub> were analysed by fixing the observed rate coefficient to the IUPAC recommendation,  $k_{\text{obs}} = (4.8 \pm 1.1) \times 10^{-13}$  cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup>, (Atkinson et al., 2006) in the fitting routine to extract the FAGE sensitivity factor for CH<sub>3</sub>O<sub>2</sub>,  $C_{\text{CH}_3\text{O}_2}$ . The obtained value,  $C_{\text{CH}_3\text{O}_2} = (5.6 \pm 0.9) \times 10^{10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup>, agrees well with the result found using the conventional radical source,  $C_{\text{CH}_3\text{O}_2} = (4.1 \pm 0.7) \times 10^{10}$  counts cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> mW<sup>-1</sup> (uncertainties quoted to 1  $\sigma$ ). The two values have overlapping error limits at 1  $\sigma$  level.

In addition to the quantitative detection of CH<sub>3</sub>O<sub>2</sub>, experiments were carried out to measure CH<sub>3</sub>O generated as a product by the CH<sub>3</sub>O<sub>2</sub> self-reaction in HIRAC. Oxygen was present at a significantly lower concentration to reduce the consumption rate of CH<sub>3</sub>O by reaction with O<sub>2</sub> in order to enable the measurement. Good agreement between the experimental data and [CH<sub>3</sub>O] generated by numerical simulations using a model describing the chemical system was obtained, demonstrating the capability to quantitatively measure CH<sub>3</sub>O. As well as CH<sub>3</sub>O<sub>2</sub>, a measurement of CH<sub>3</sub>O will be useful as a further model target in future mechanistic studies of atmospherically relevant chemical systems within HIRAC.

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